

Ecology and physiology of planktonic archaea

Alyson Santoro

Horn Point Laboratory

University of Maryland Center for Environmental Science

C-MORE Microbial Oceanography Course 2014

Talk outline

Who are the marine archaea?

Tracking down the MGI archaea

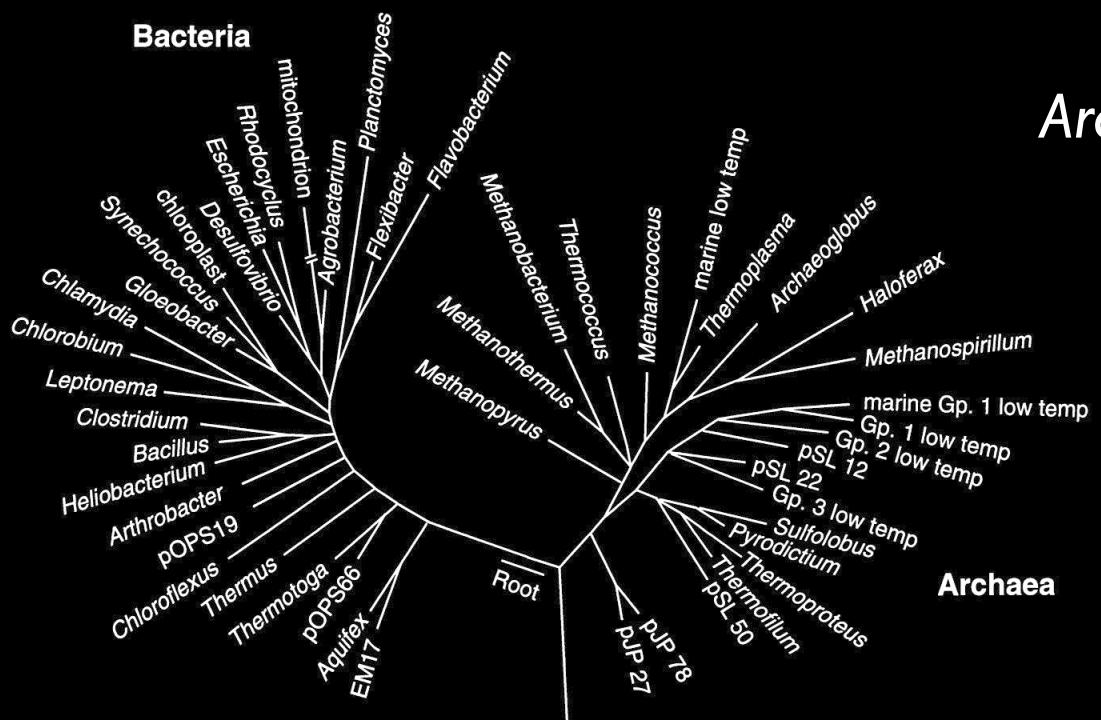
Taming wild archaea to investigate their physiology

The elusive Euryarchaeota

Who are the marine archaea?

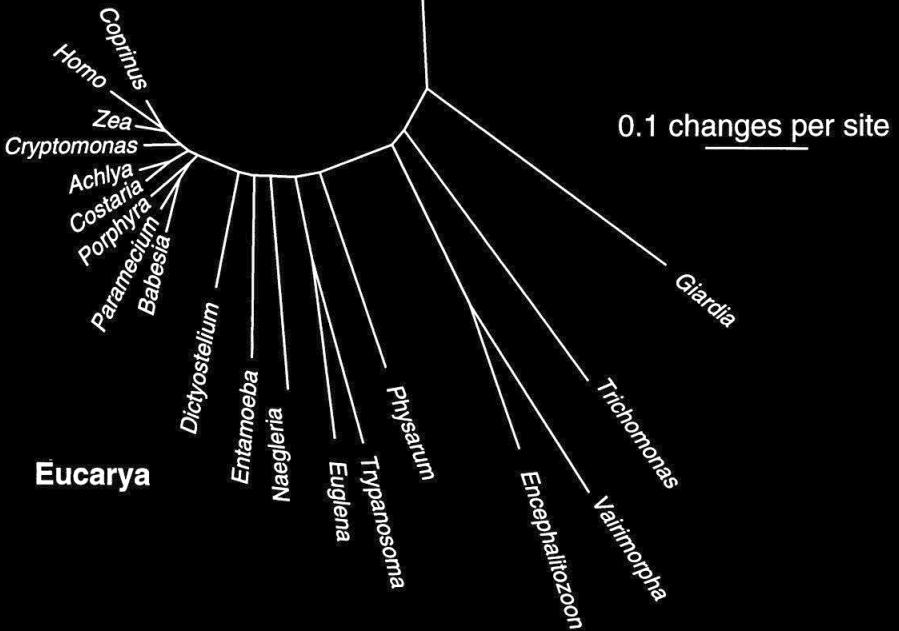
Bacteria

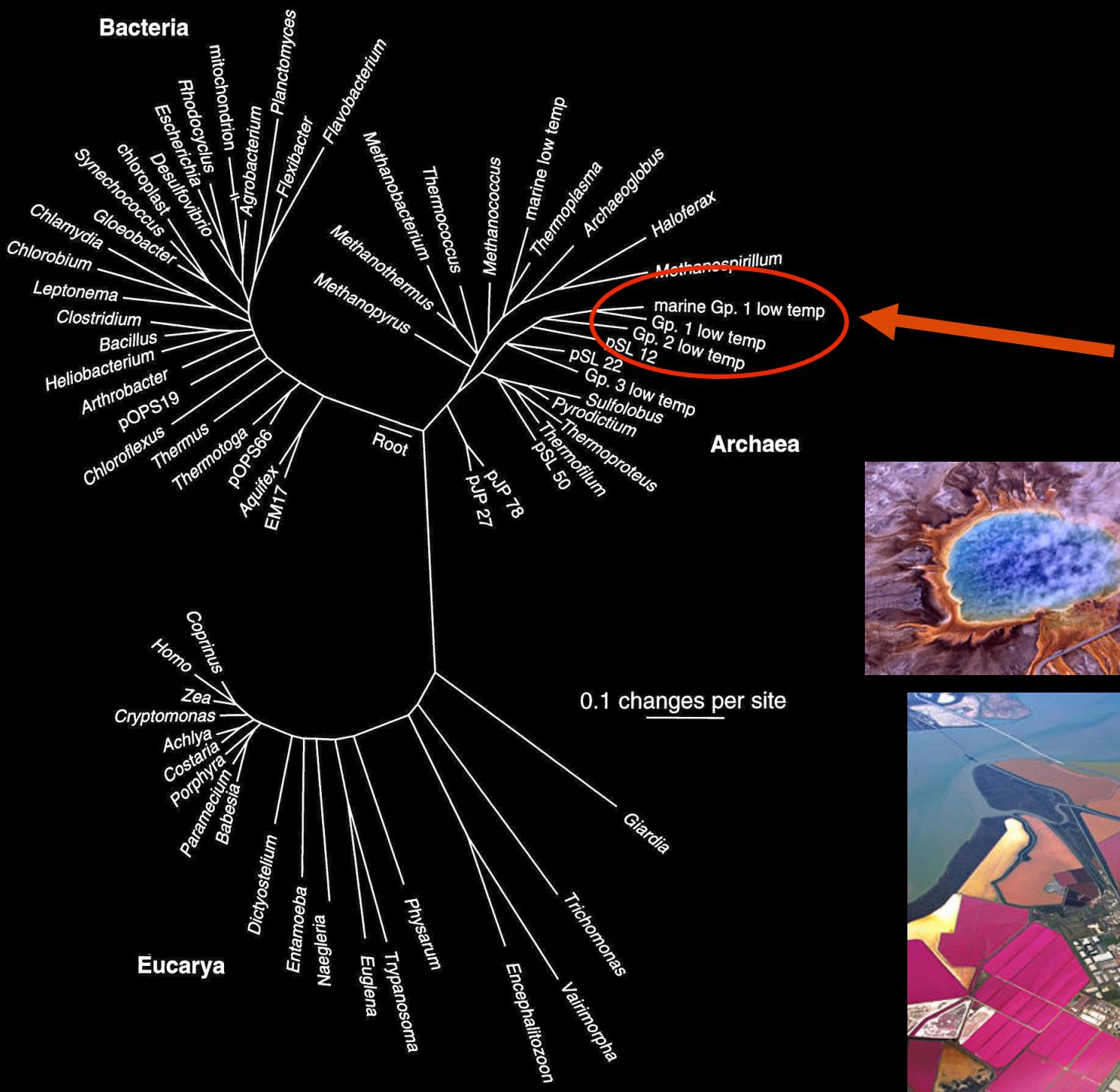
Archaea



Eucarya

Eucarya

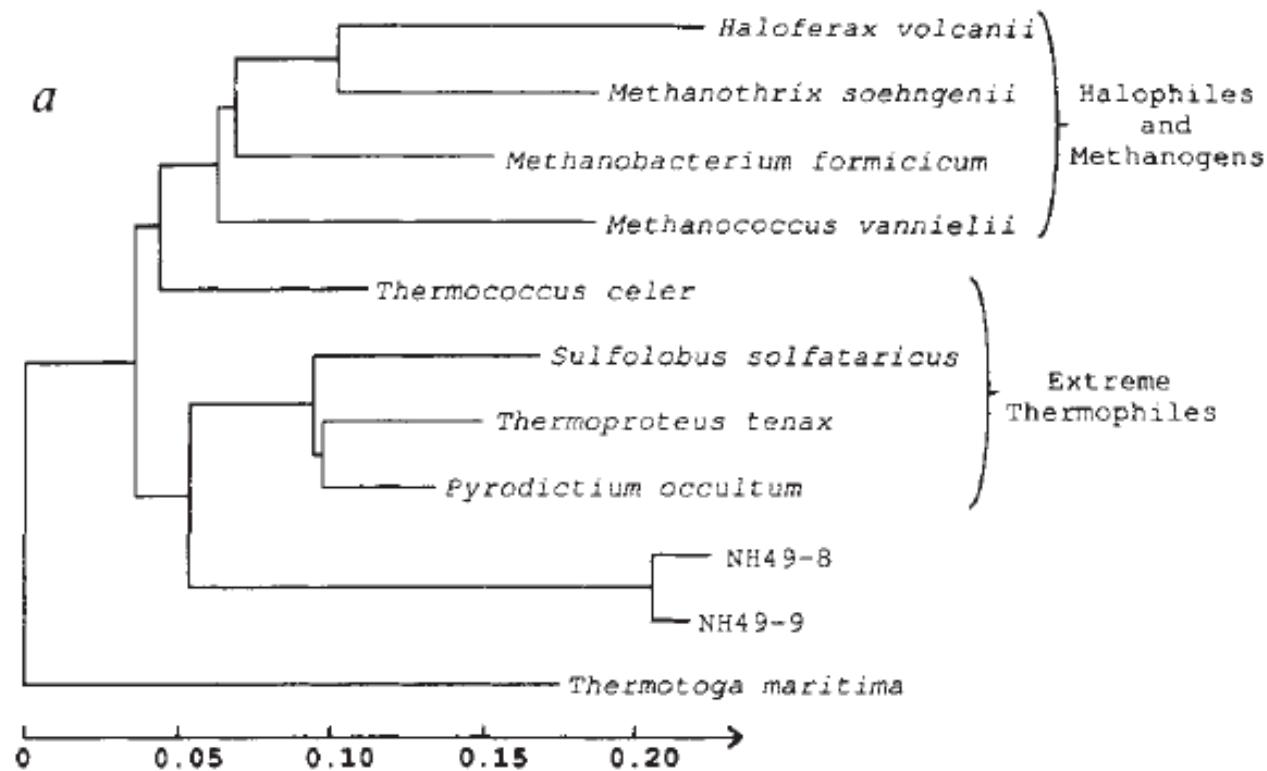




Novel major archaeabacterial group from marine plankton

Jed A. Fuhrman, Kirk McCallum* & Alison A. Davis

Department of Biological Sciences, University of Southern California,
Los Angeles, California

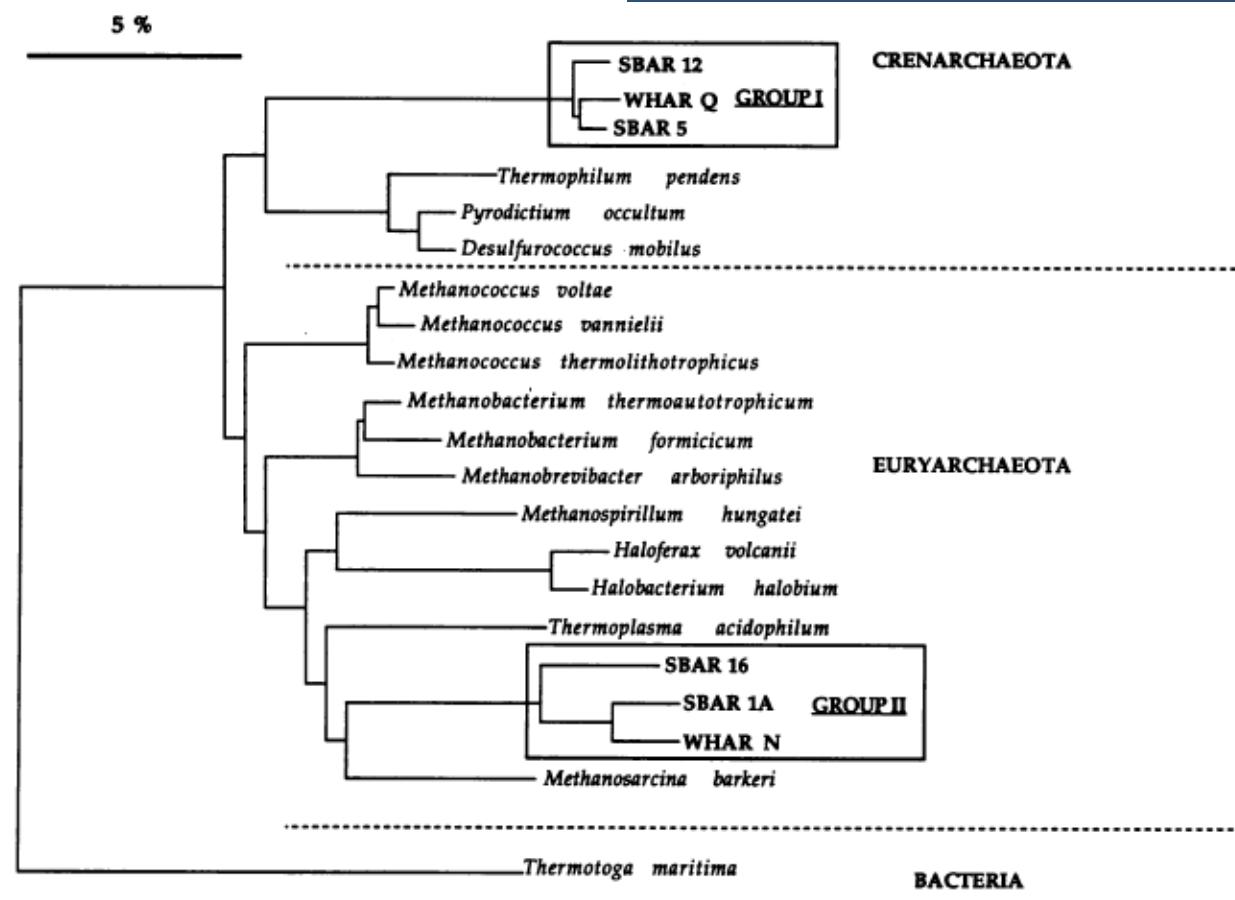


Archaea in coastal marine environments

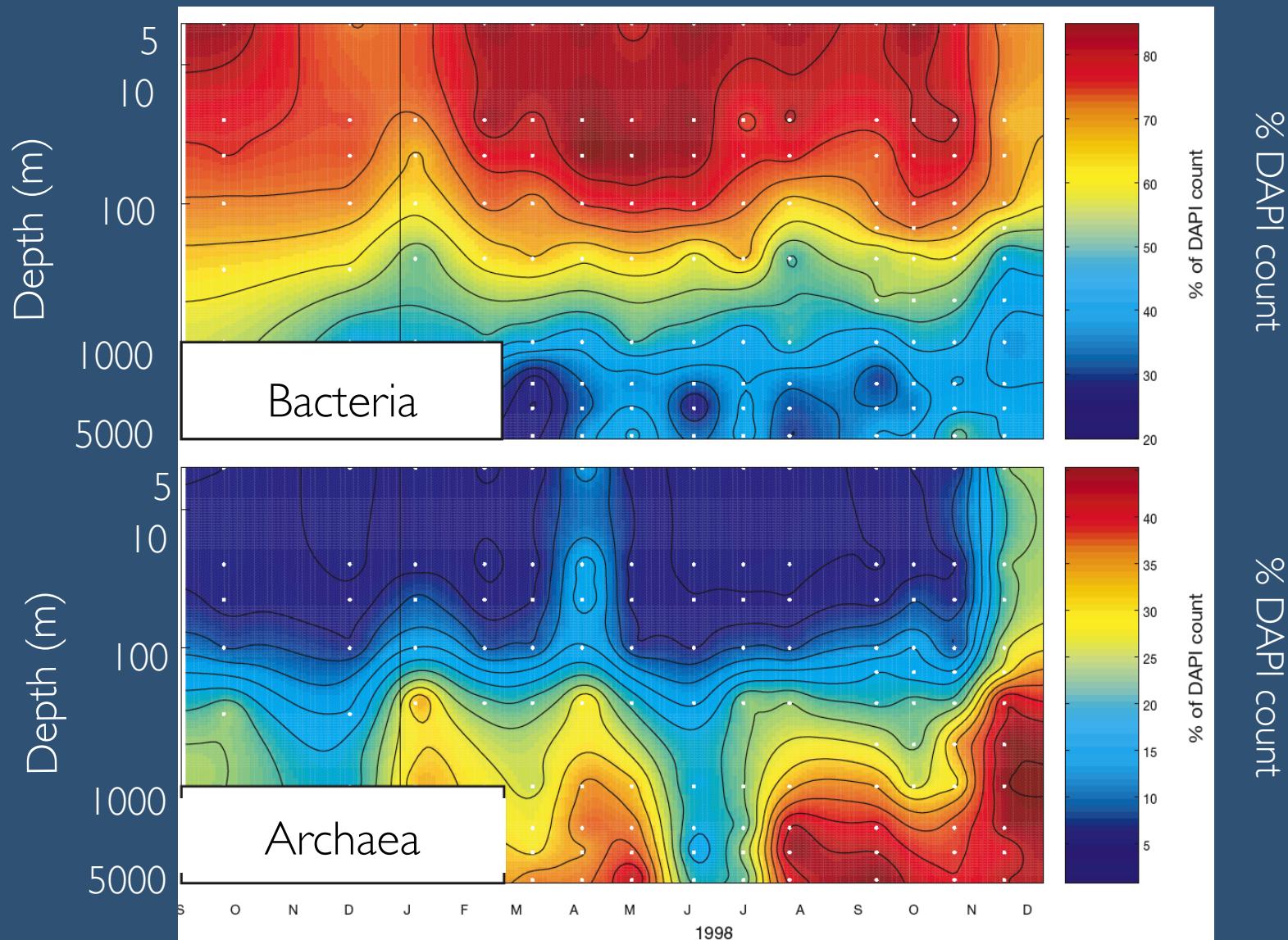
(archaeabacteria/phylogeny/bacterioplankton/molecular ecology)

EDWARD F. DELONG*

Biology Department, Woods Hole Oceanographic Institution, Woods Hole, MA 02543

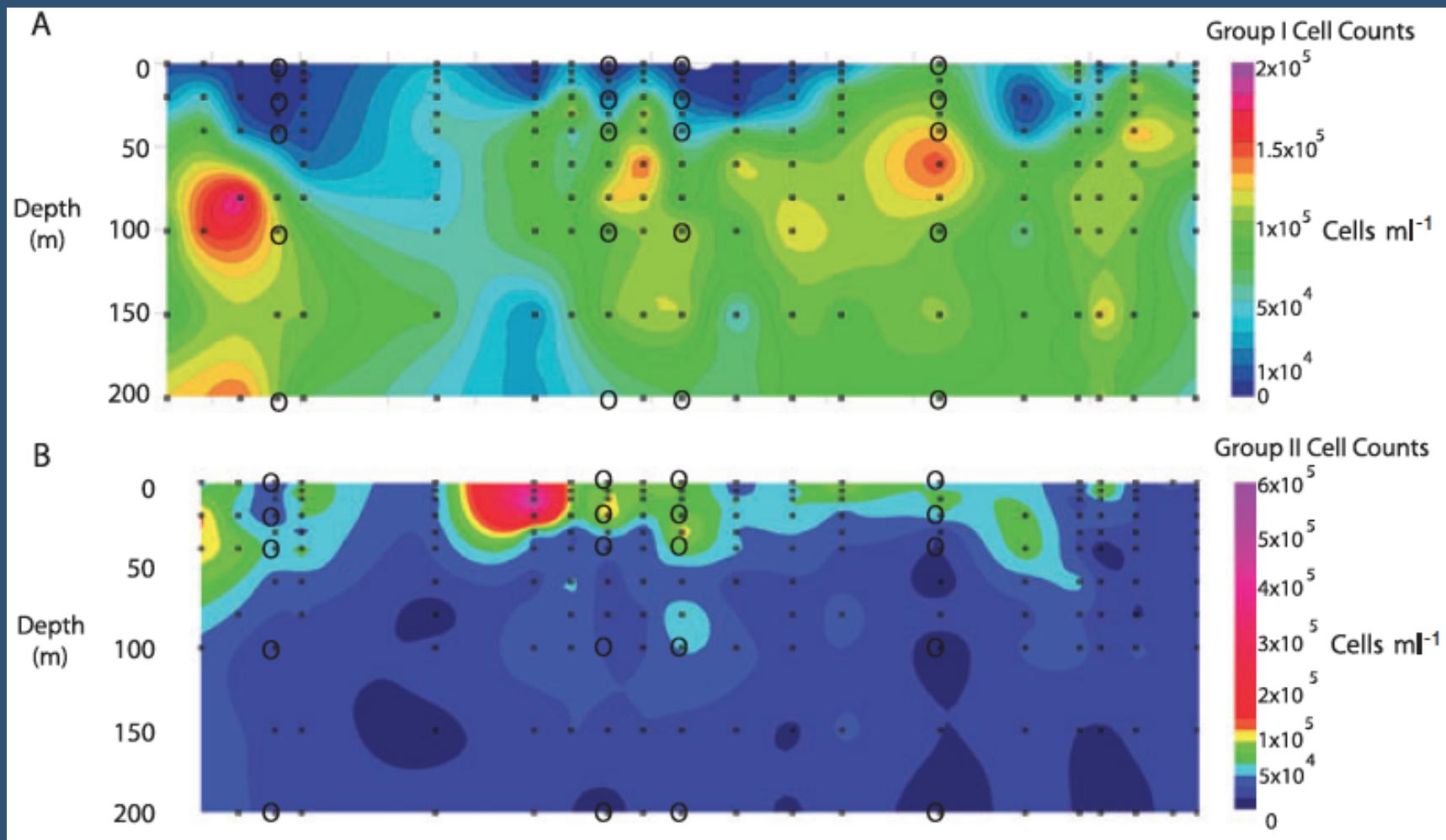


Mesophilic archaea are the most abundant cells in the deep ocean (and possibly the planet).



modified from Karner et al., Nature, 2001

Contrasting distribution of MGI and MGII



MGI archaea: Tracking down the most abundant cells in the deep ocean

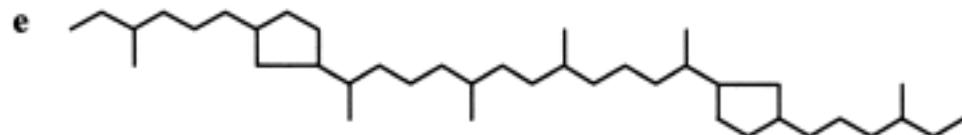
MGI archaea are autotrophic?

Origins of lipid biomarkers in Santa Monica Basin surface sediment: A case study using compound-specific $\Delta^{14}\text{C}$ analysis

A. PEARSON,^{1,*} A. P. McNICHOL,² B. C. BENITEZ-NELSON,^{1,†} J. M. HAYES,² and T. I. EGLINTON¹

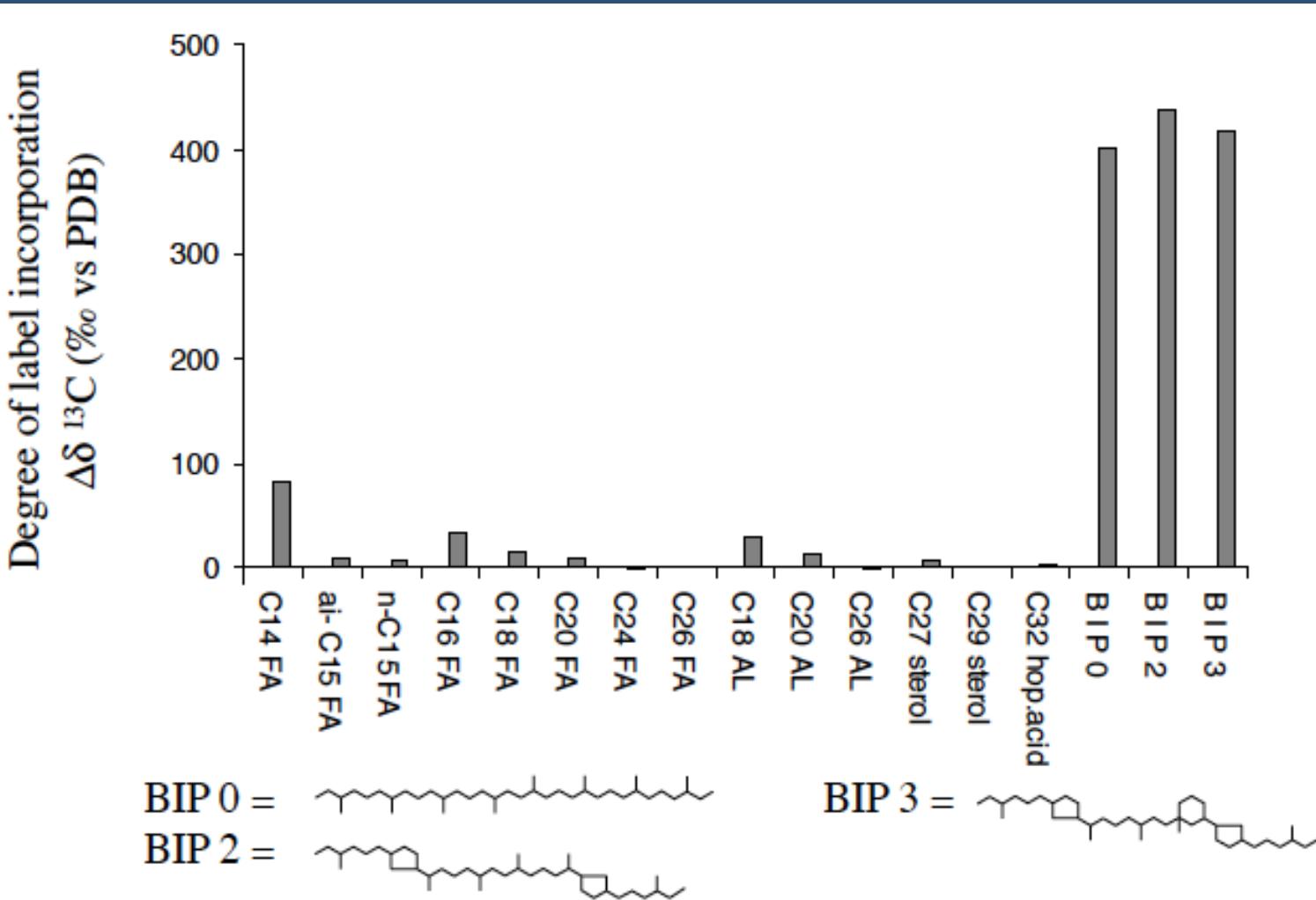
¹Department of Marine Chemistry and Geochemistry, Woods Hole Oceanographic Institution, Woods Hole, MA 02543 USA

²National Ocean Sciences Accelerator Mass Spectrometry (NOSAMS) Facility, Department of Marine Geology and Geophysics, Woods Hole Oceanographic Institution, Woods Hole, MA 02543, USA



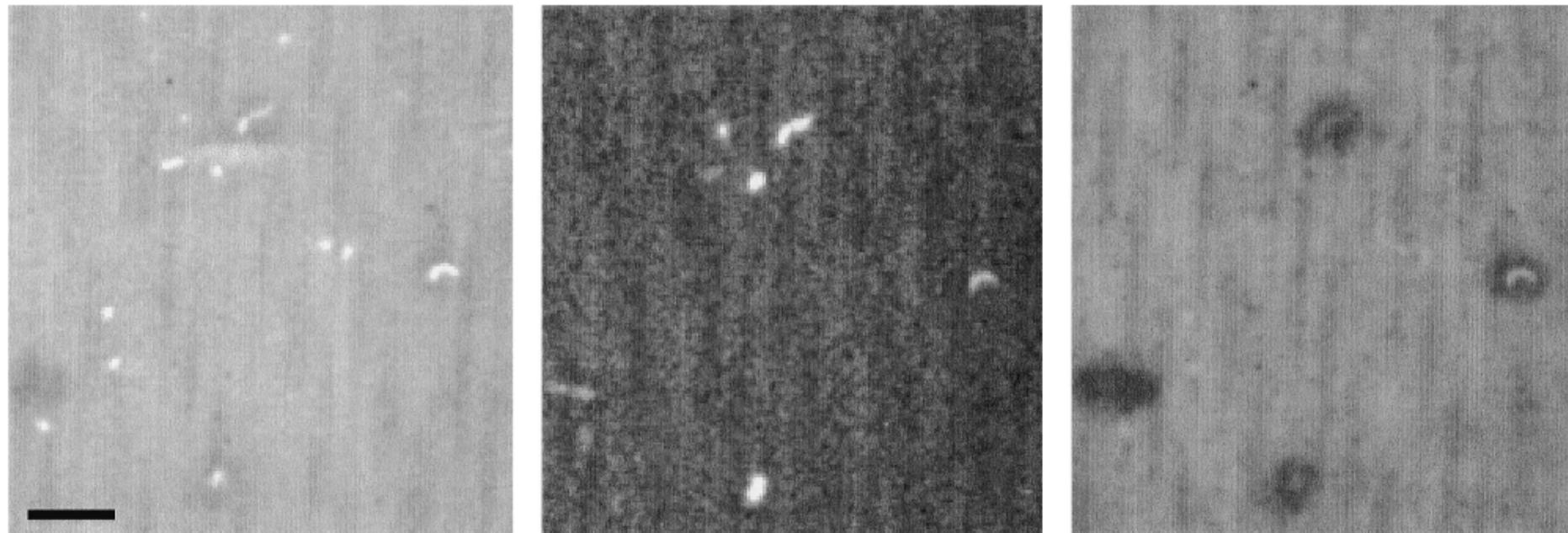
“The $\Delta^{14}\text{C}$ data are consistent with chemoautotrophic growth below the euphotic zone.”

MGI archaea are autotrophic?



Wuchter et al. 2003

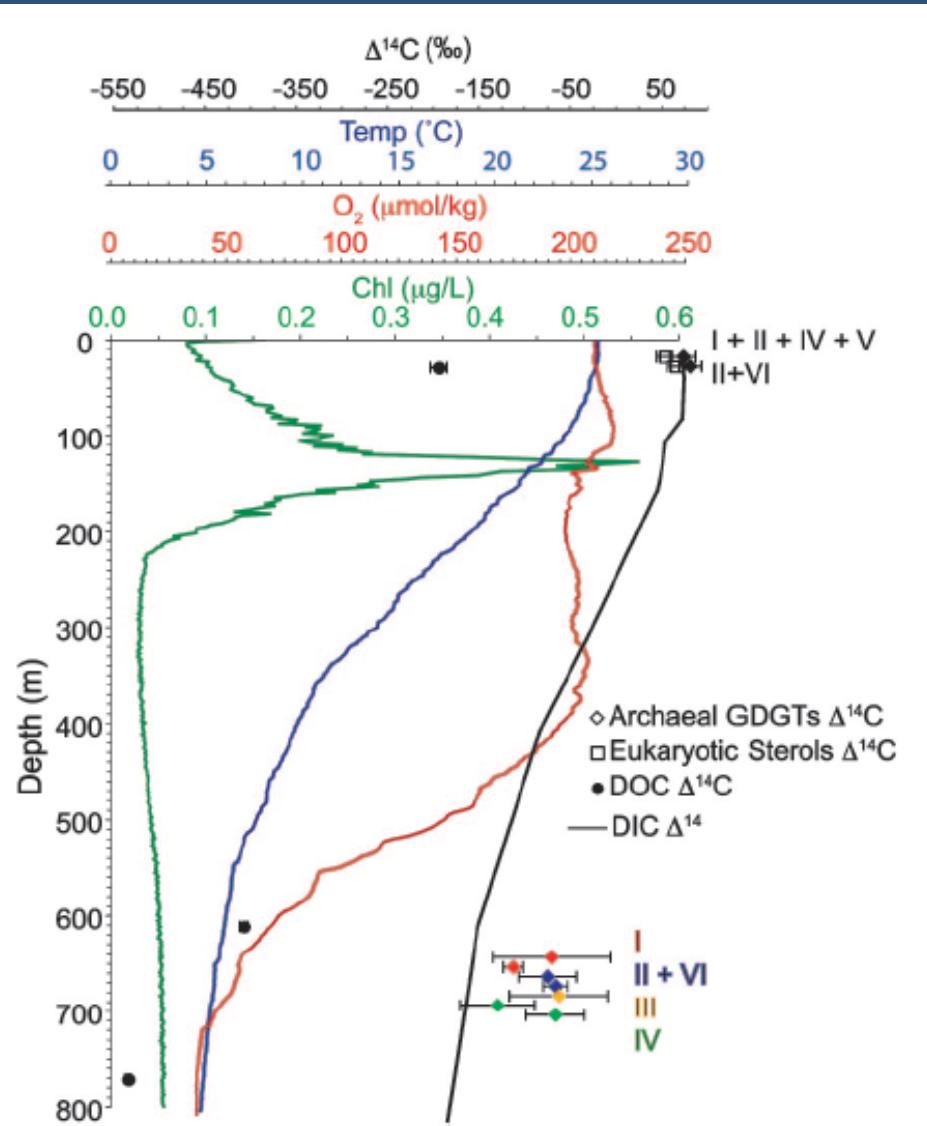
MGI archaea are heterotrophic?



Uptake of ^{14}C labeled amino acids into archaeal cells.

Ouverney and Fuhrman 2000

MGI archaea are mixotrophic?



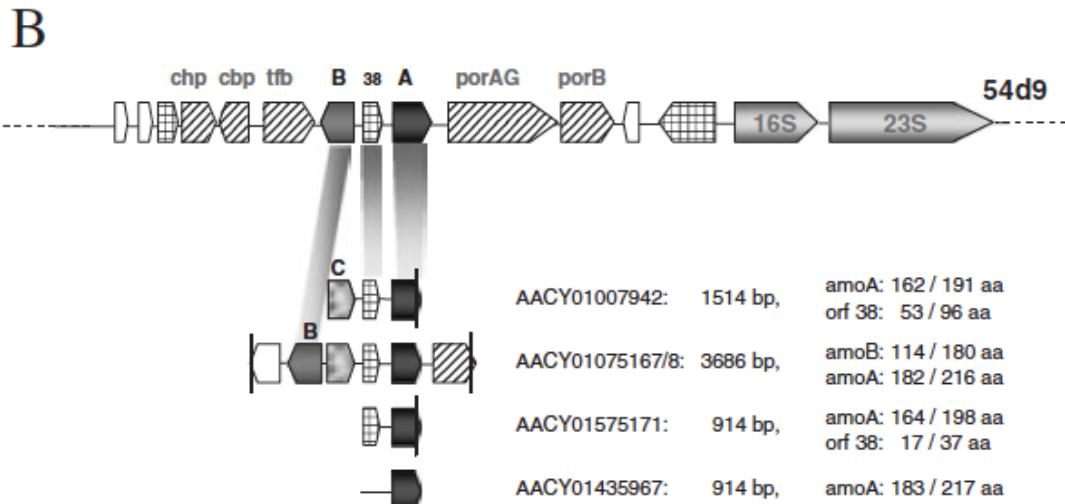
Suggests ~80% of C is coming from autotrophy, 20% from heterotrophy.

Ingalls et al. 2006

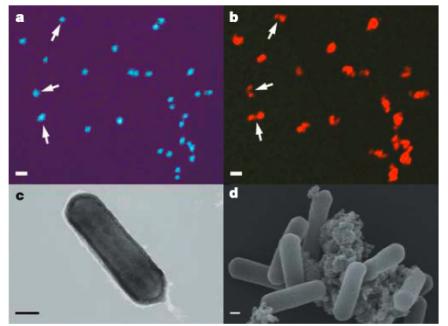
Environmental Genome Shotgun Sequencing of the Sargasso Sea

J. Craig Venter,^{1*} Karin Remington,¹ John F. Heidelberg,¹ Daniel R. Eisen,¹ Michael H. Hickey,¹ Robert L. Durkin,¹ Daniel J. Relman,¹ and J. William Tyson²

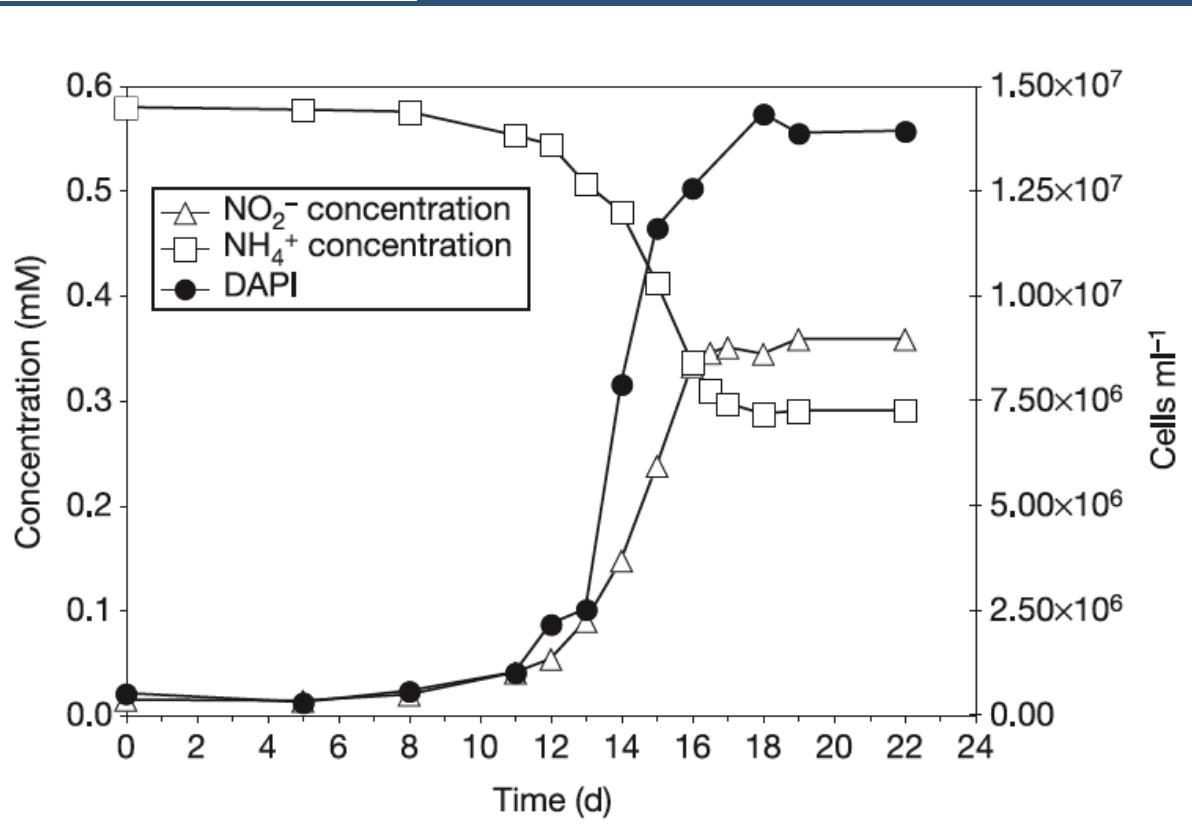
We are a long way from a full understanding of the biology of the organisms here, but this relatively small genome demonstrates areas where important insights may be gained. For example, because it has been believed that only members of the bacterial domain were capable of oceanic nitrification, it is interesting to note that an ammonium monooxygenase gene was found on an archaeal-associated scaffold within our data set. Though ultraviolet (UV) light in the



Venter et al. 2003; Treusch et al. 2005



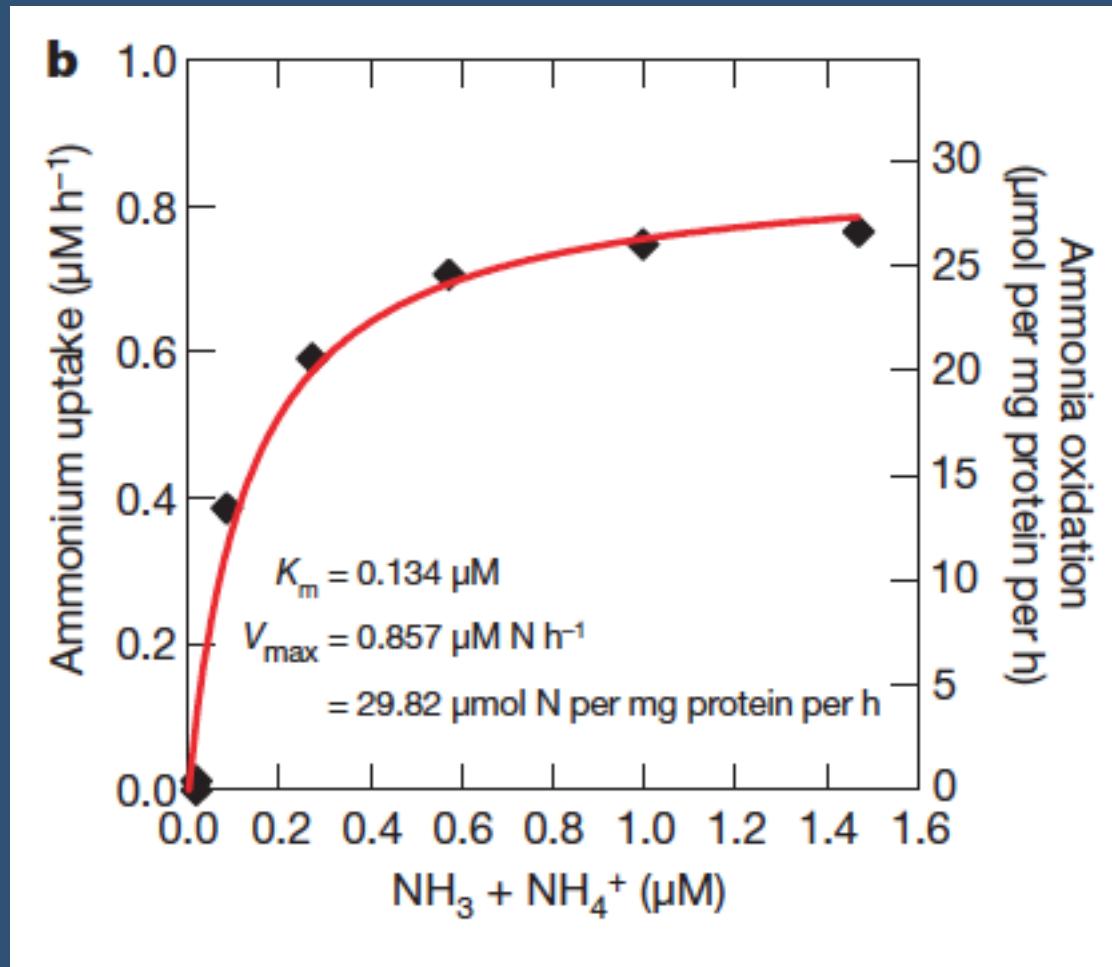
The first culture of a marine group I archaea, isolated from gravel at the Seattle Aquarium, *Nitrosopumilus maritimus*.



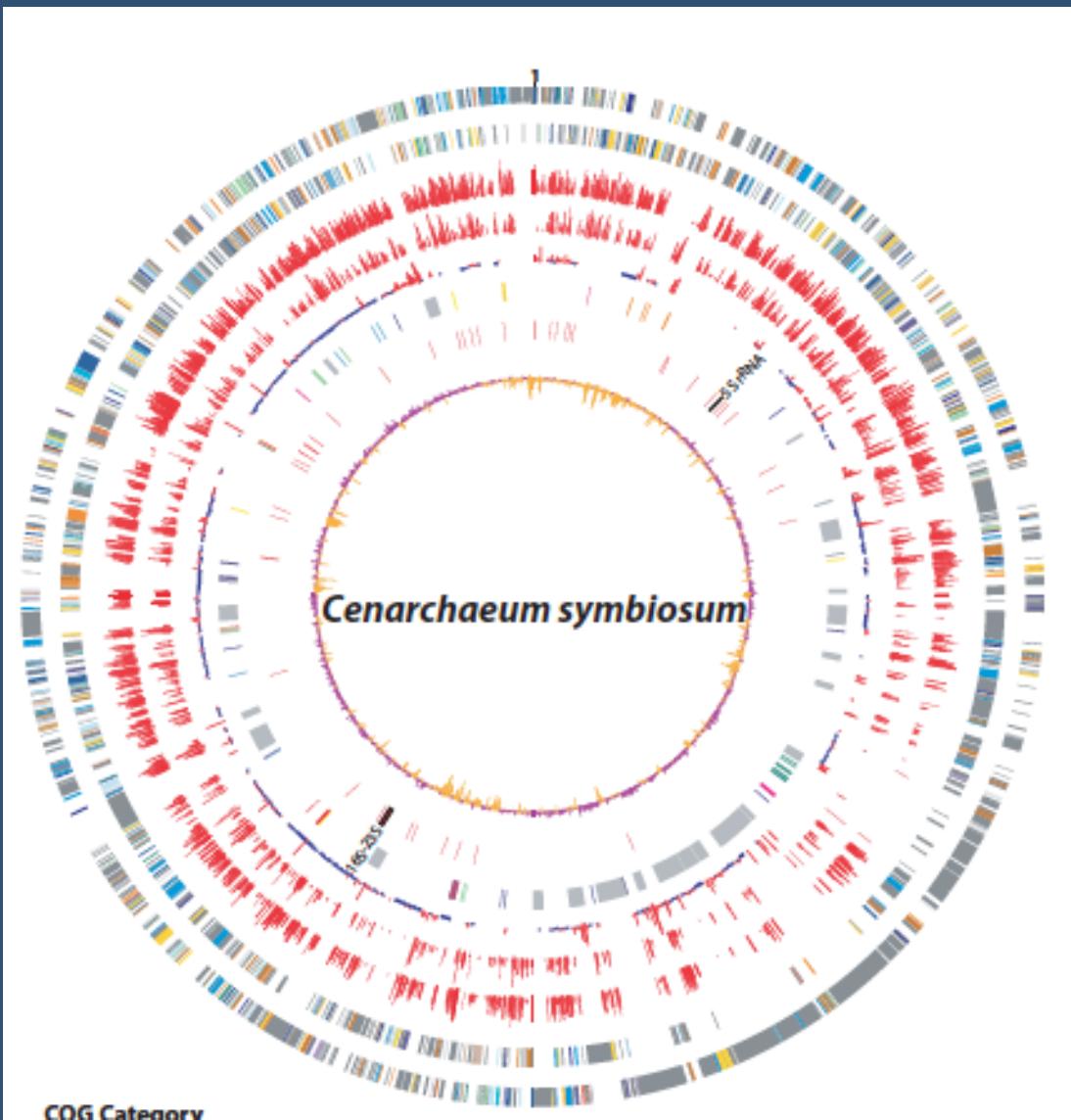
Showed that MGI archaea can oxidize NH_3

Grow as chemolithoautotrophs

N. maritimus has a high affinity for ammonium

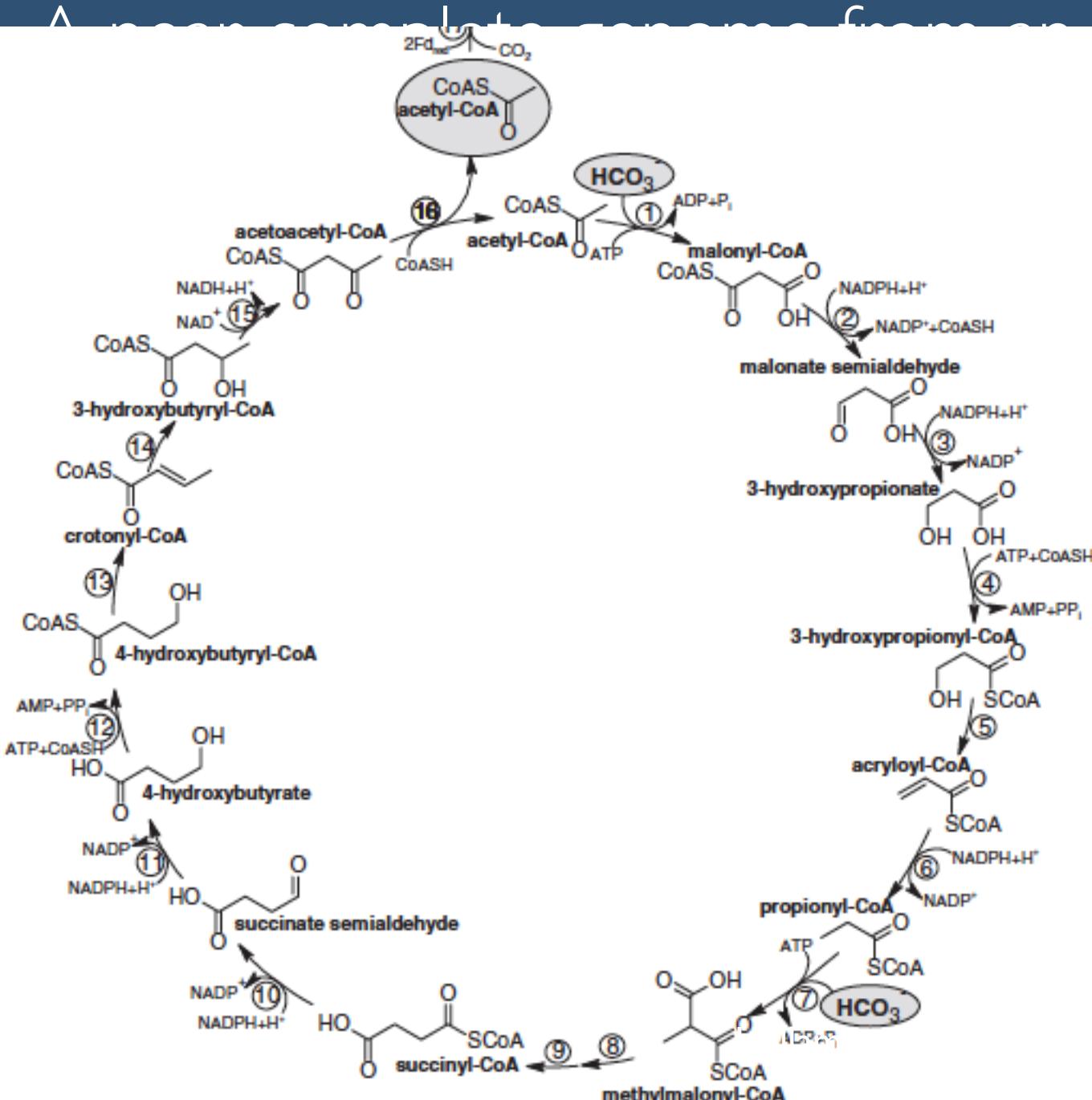


A near-complete genome from an uncultivated archaea



Hallam et al. 2006; Berg et al. 2007

t al. 2007

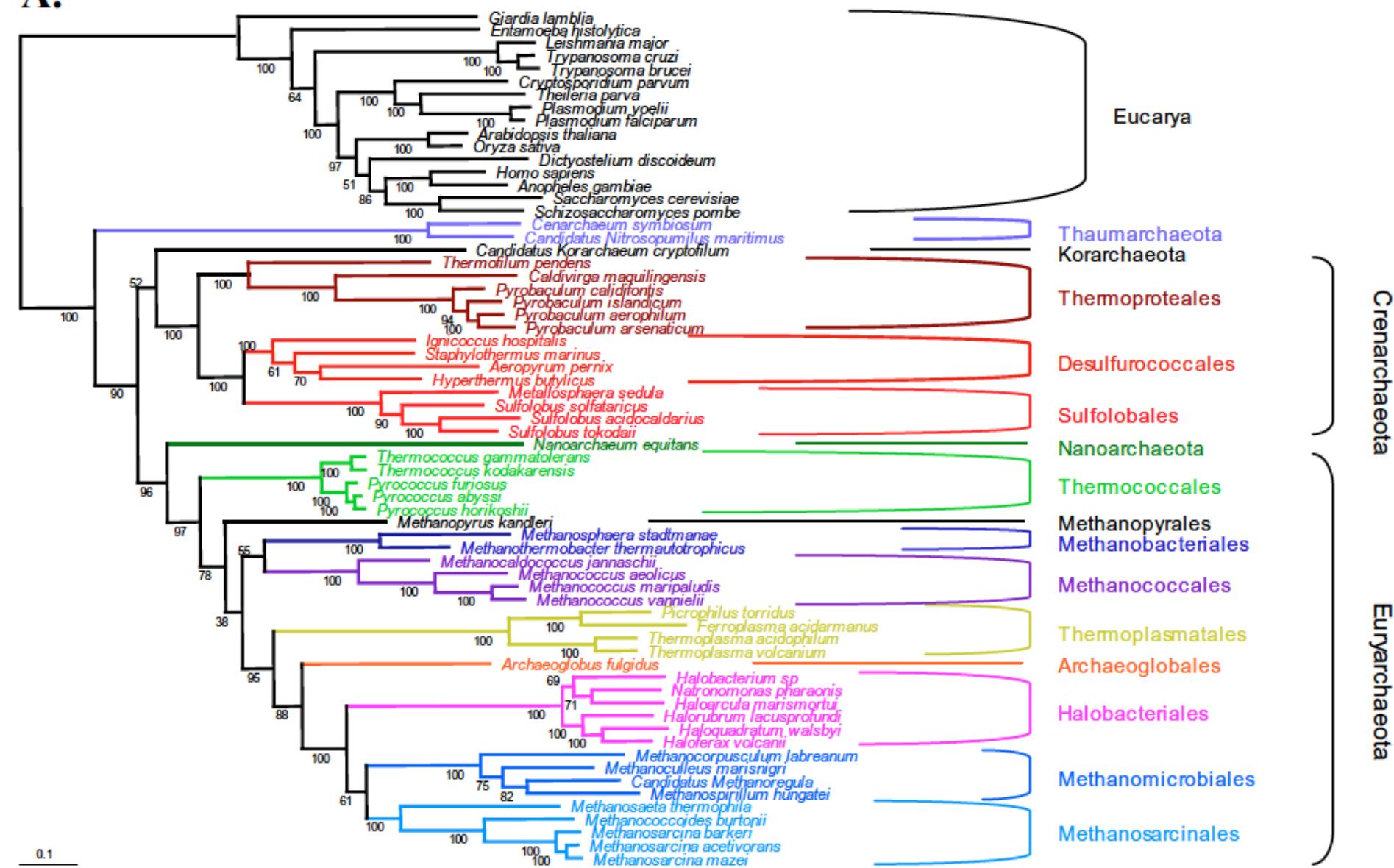


Thaumarchaea fix carbon using an extremely energy efficient pathway

Table 4. Comparison of the energy efficiency of aerobic autotrophic CO₂ fixation cycles

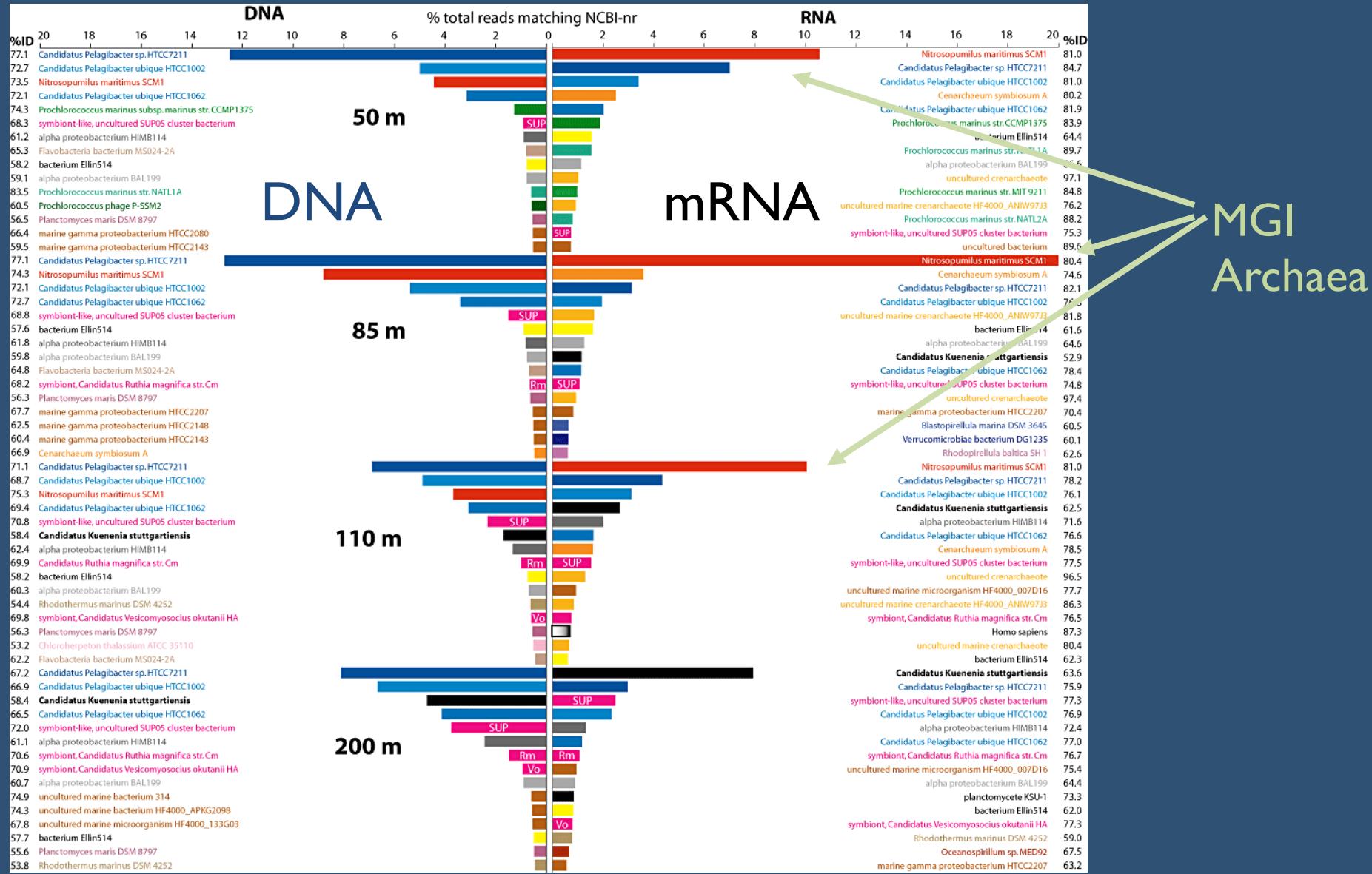
1 mole precursor or 1 g biomass	Calvin–Benson cycle*	3-Hydroxypropionate bicycle	Crenarchaeal HP/HB cycle	Thaumarchaeal HP/HB cycle
Acetyl-CoA	7	7	6	4
Pyruvate	7	7	9–10 [†]	5
Phosphoenolpyruvate	8	9	10–11 [†]	7
Oxaloacetate	8	9	9–10 [†]	6
2-Oxoglutarate	15	16	15–16 [†]	10
Biomass (1 g)	0.12*	0.13	0.13–0.15 [†]	0.09

The numbers represent moles of high-energy anhydride bonds of ATP required to form 1 mole of central precursor metabolite or of the main precursors for the synthesis of 1 g of dry biomass. For the details of calculations, see *SI Appendix, SI Text*. The estimated amount of the central metabolic precursors



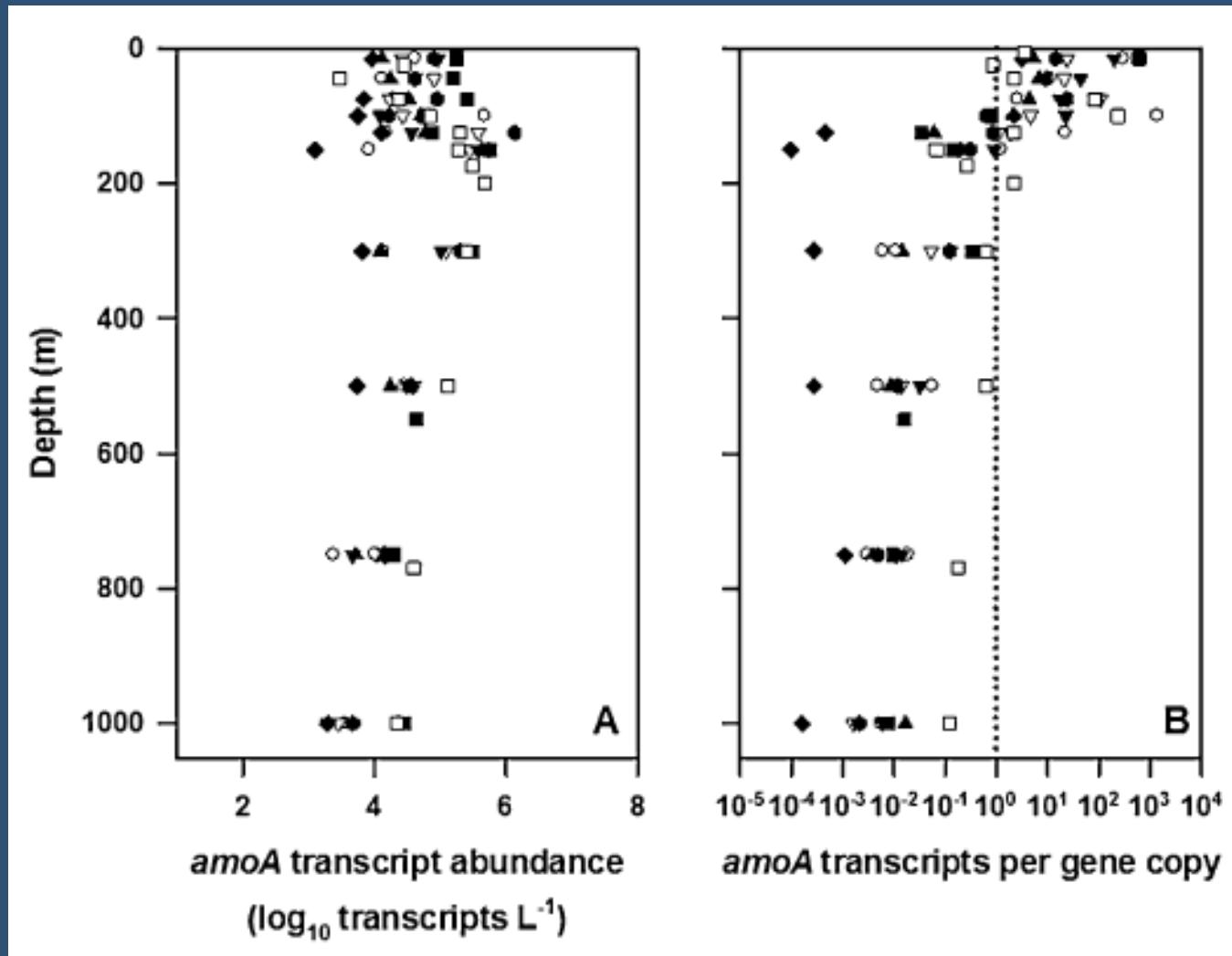
Brochier-Armanet et al. 2008; Walker et al. 2010; Kelly et al. 2010

Archaea are active in low oxygen waters

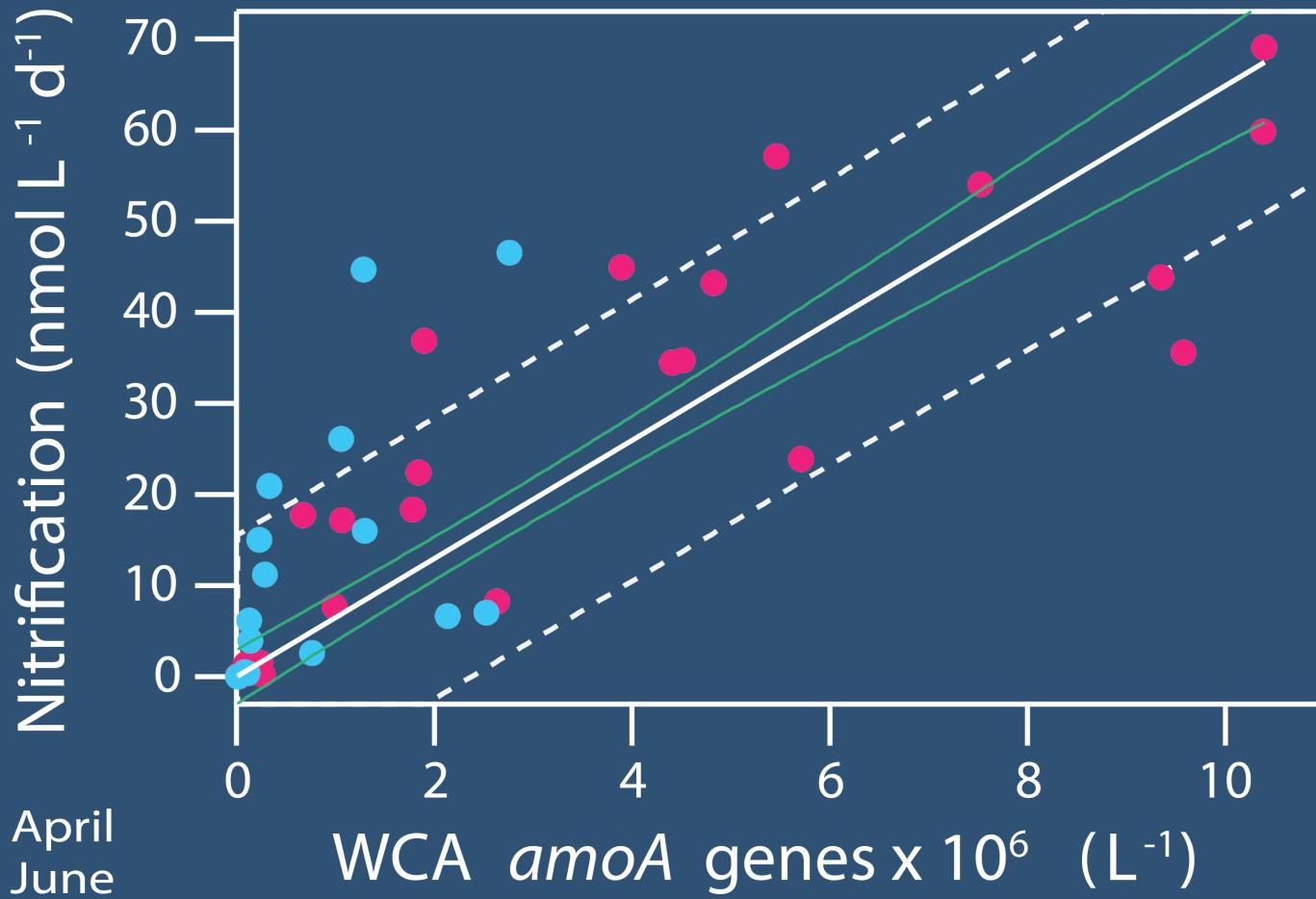


Stewart et al. 2011

Abundant *amoA* transcripts in surface waters at Station ALOHA

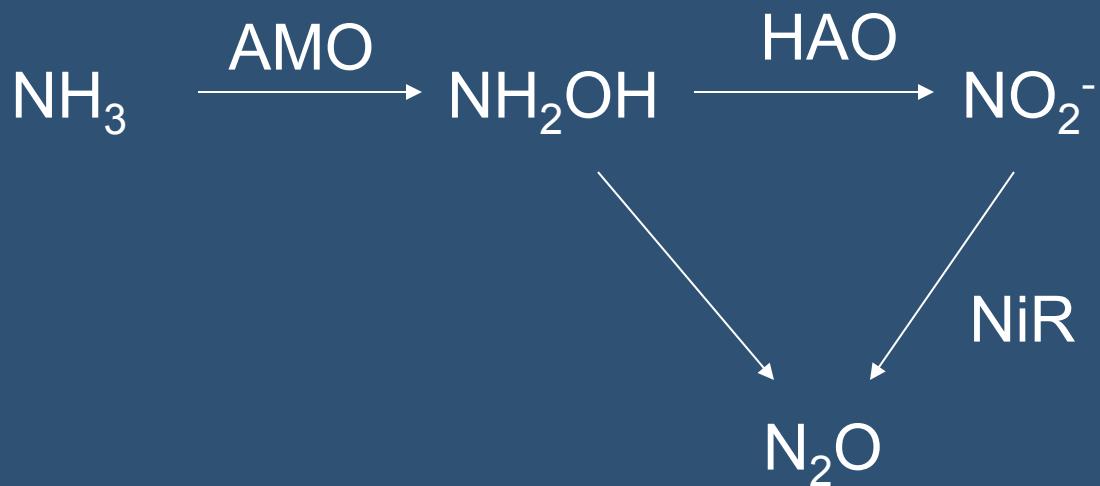


Abundance of the shallow ecotype correlates with nitrification rates



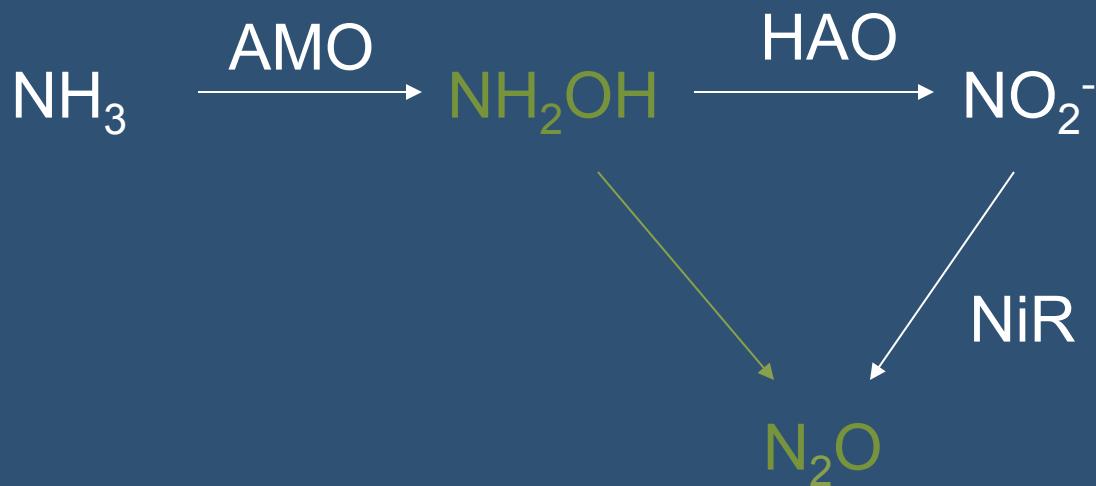
from Smith et al. 2014, ISMEJ

AOB make nitrous oxide (N_2O)

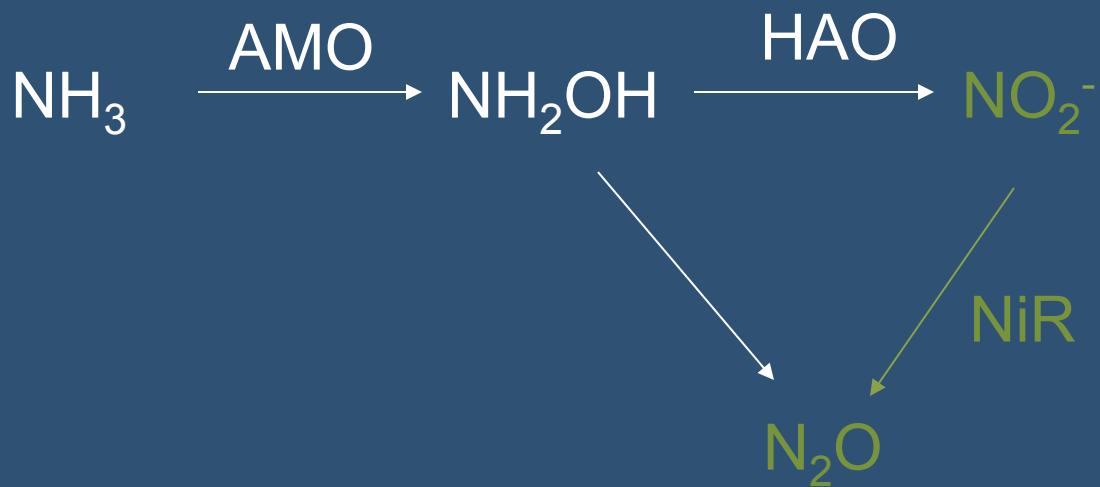


AOB = ammonia-oxidizing bacteria

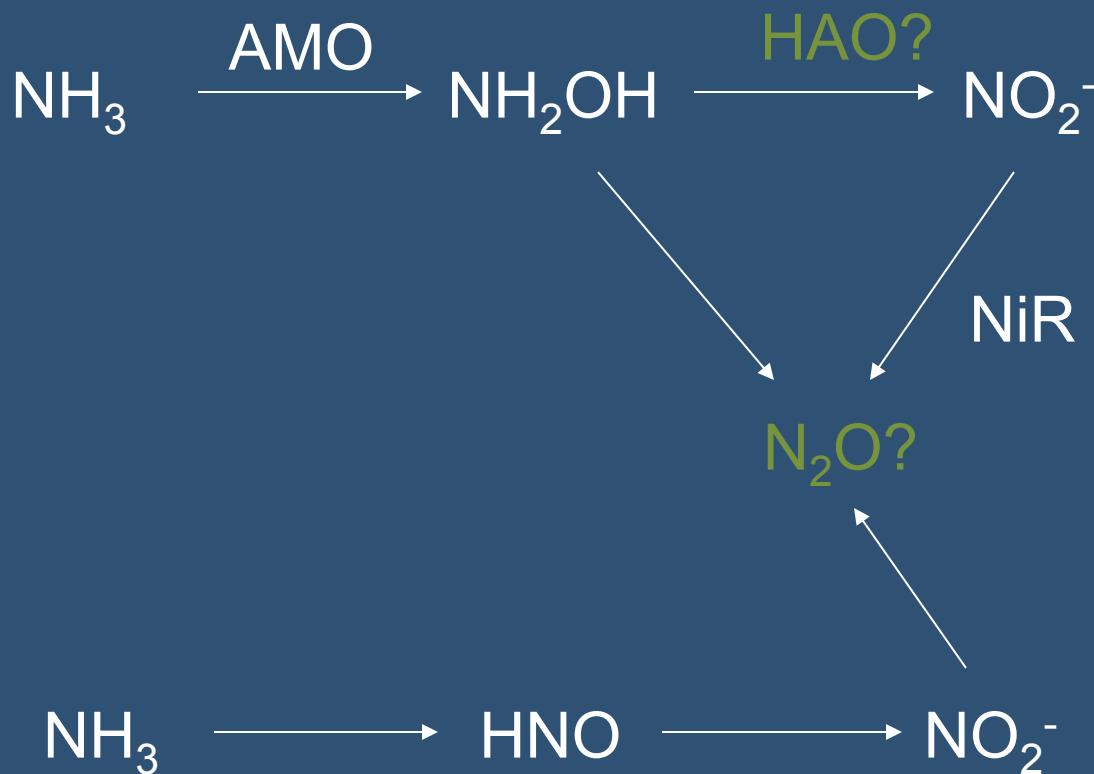
AOB can make nitrous oxide in two ways



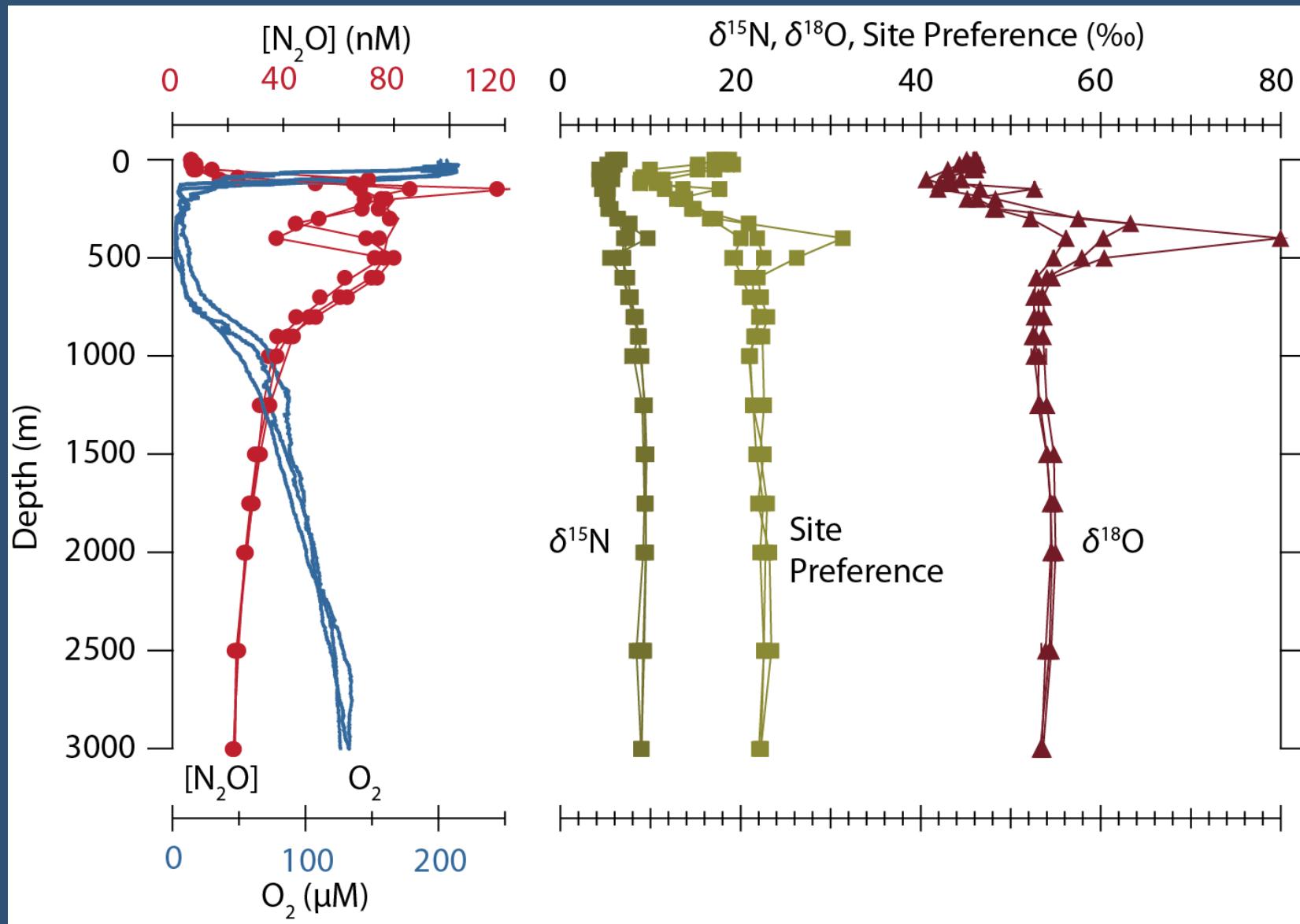
AOB can make nitrous oxide in two ways



Can archaea make N₂O?



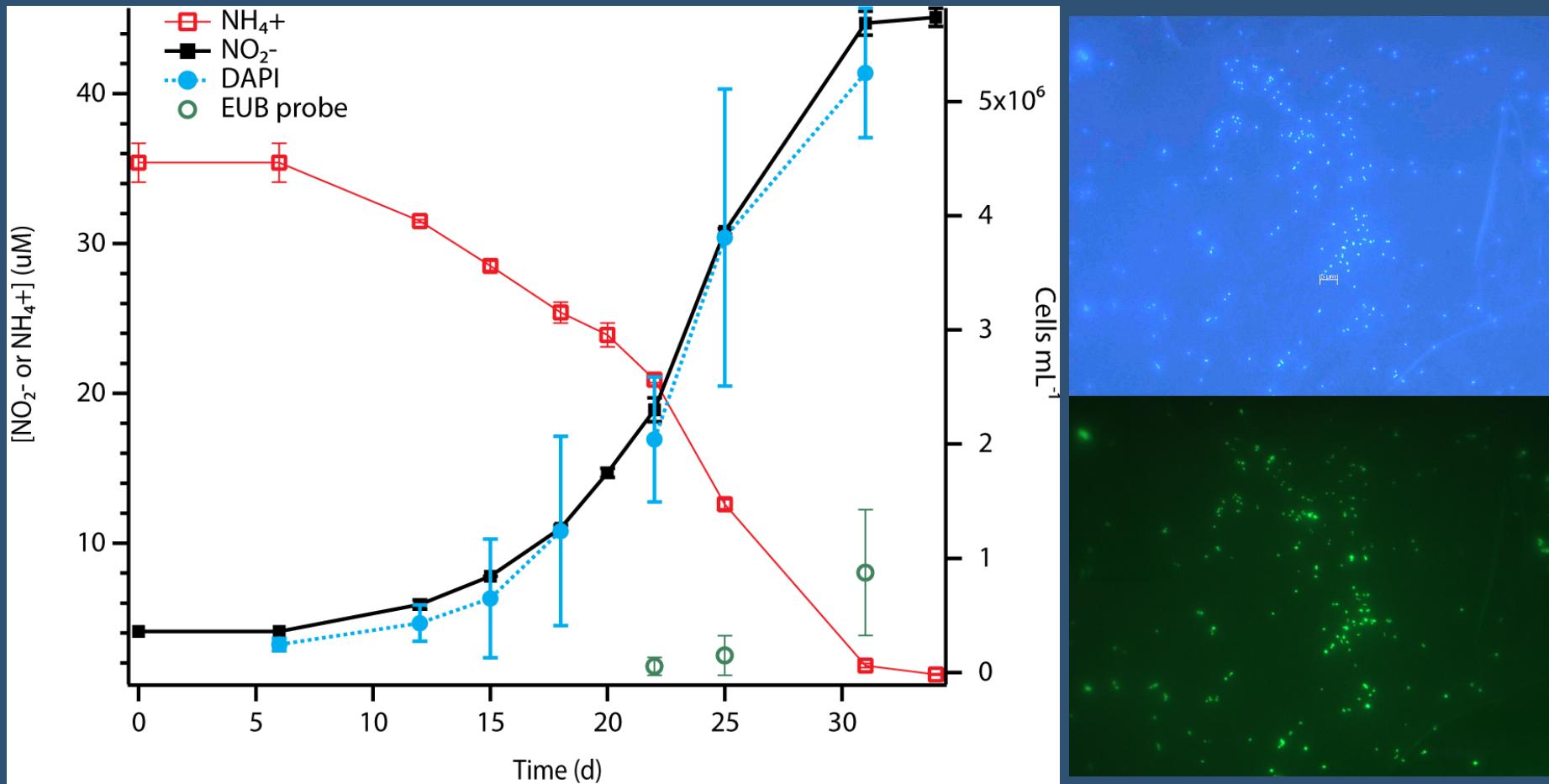
Interpreting N_2O profiles requires some information about microbial physiology



	Anammox	AOB	AOA
e- transport intermediate oxidase	Fe	Fe	Cu
	HAO/HZO	HAO	?
C-fixation	acetyl-CoA	CBB	3-Hp/4-Hb
NO ₂ ⁻ reduction	<i>nirS</i>	<i>nirK</i>	<i>nirK</i>

Taming wild archaea

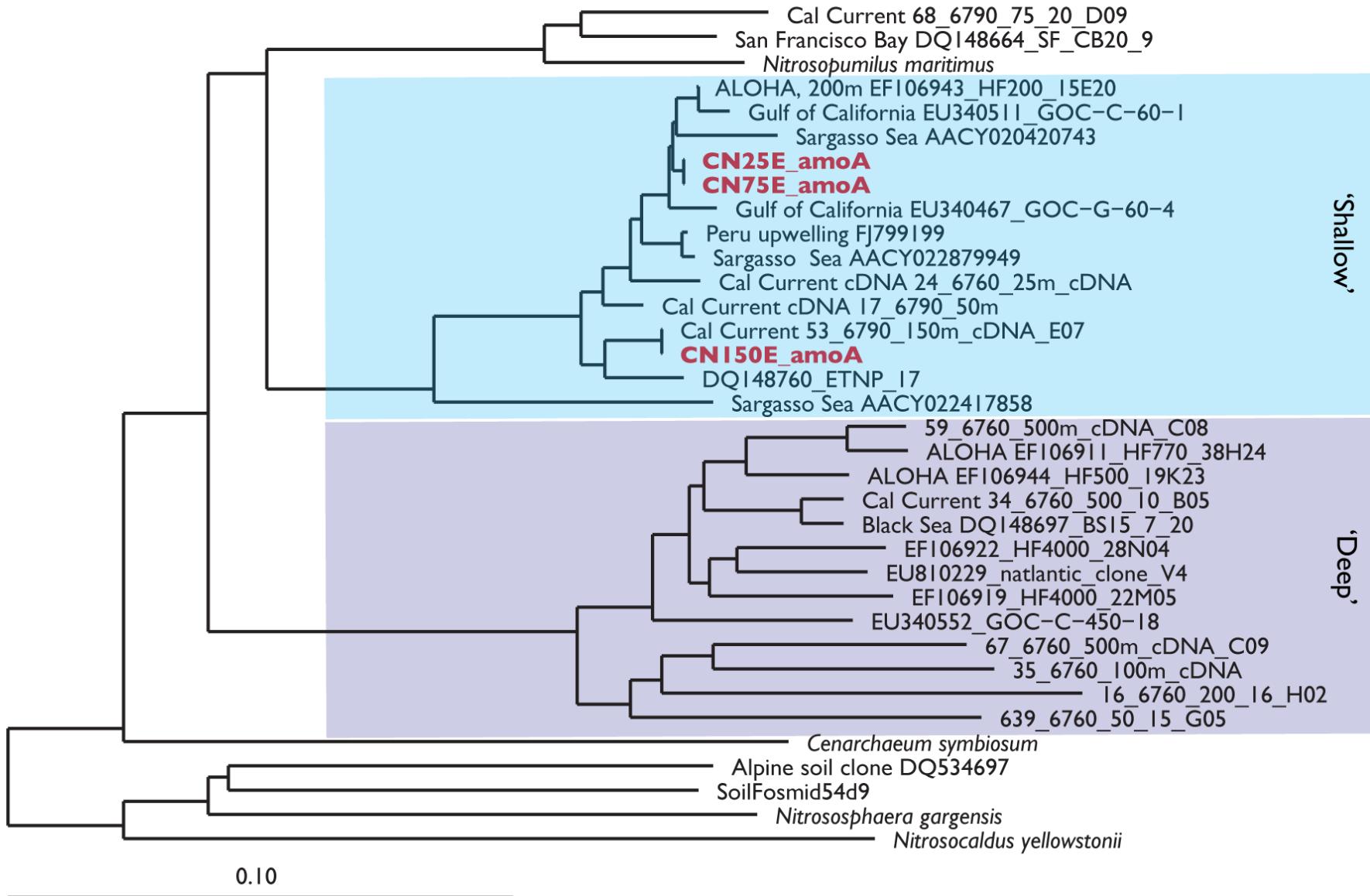
Enrichment ‘CN25’ from the North Pacific



Growth rate = 0.17 d^{-1} , doubling time $\sim 4 \text{ d}$

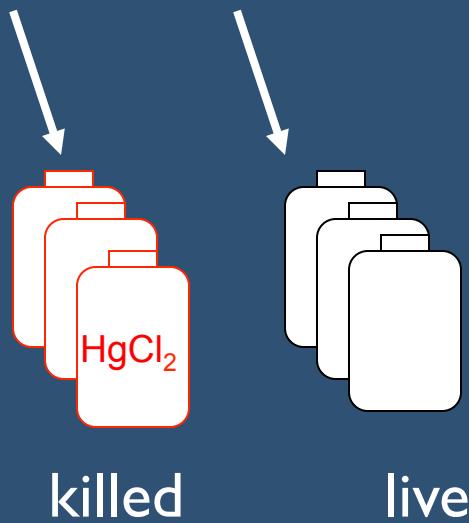
Santoro and Casciotti, 2011; ISMEJ

CN25 is the ‘shallow’ AOA ecotype





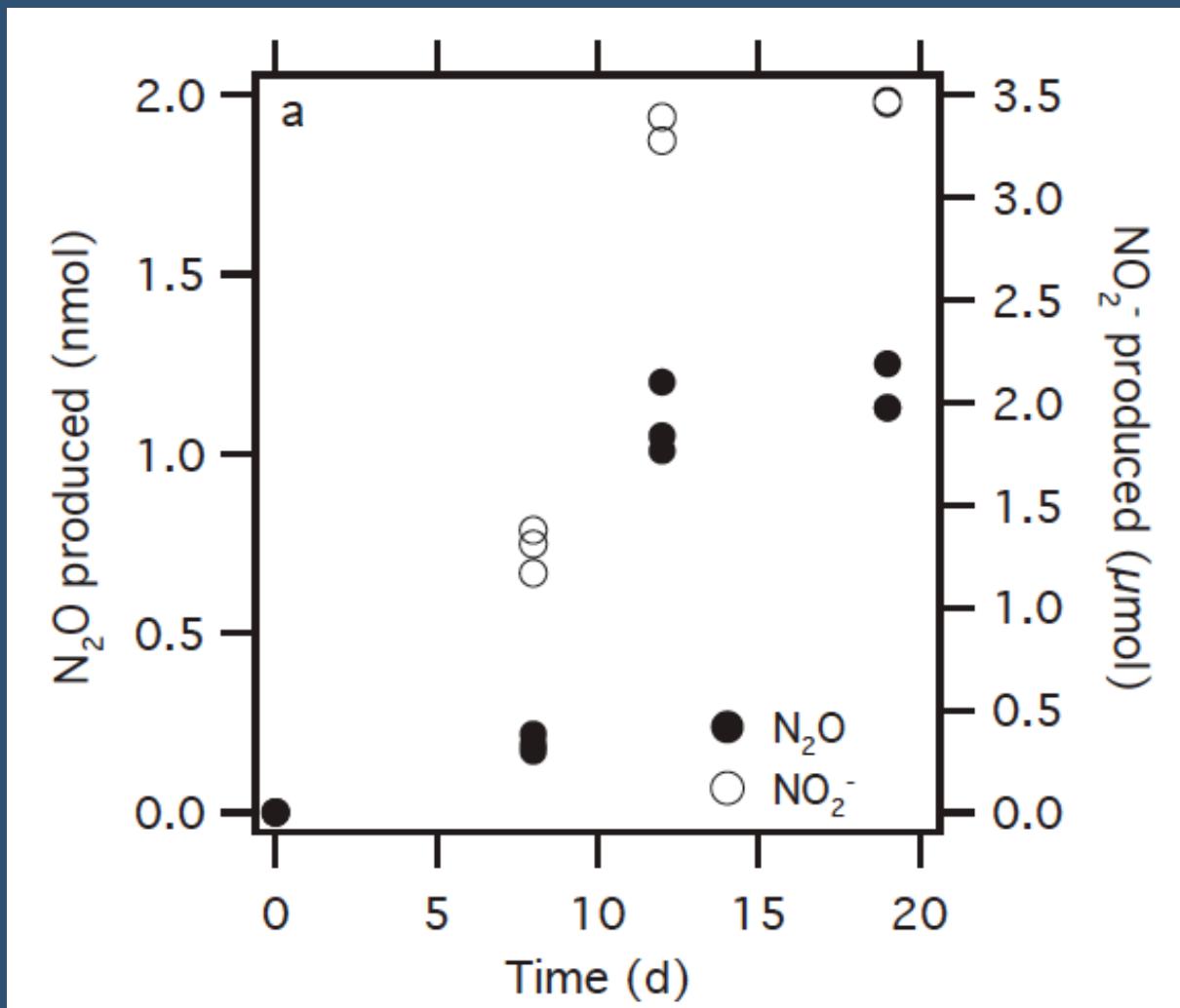
Measuring N₂O production



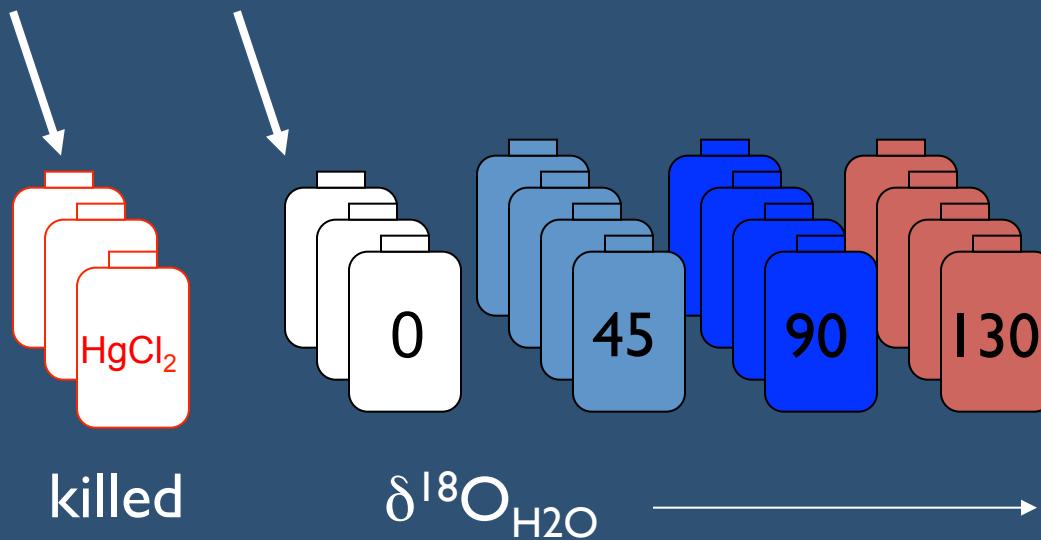
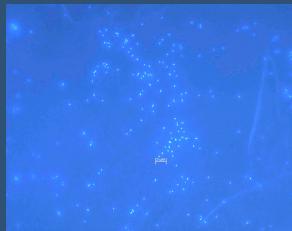
6 weeks →

N₂O produced =
live - killed

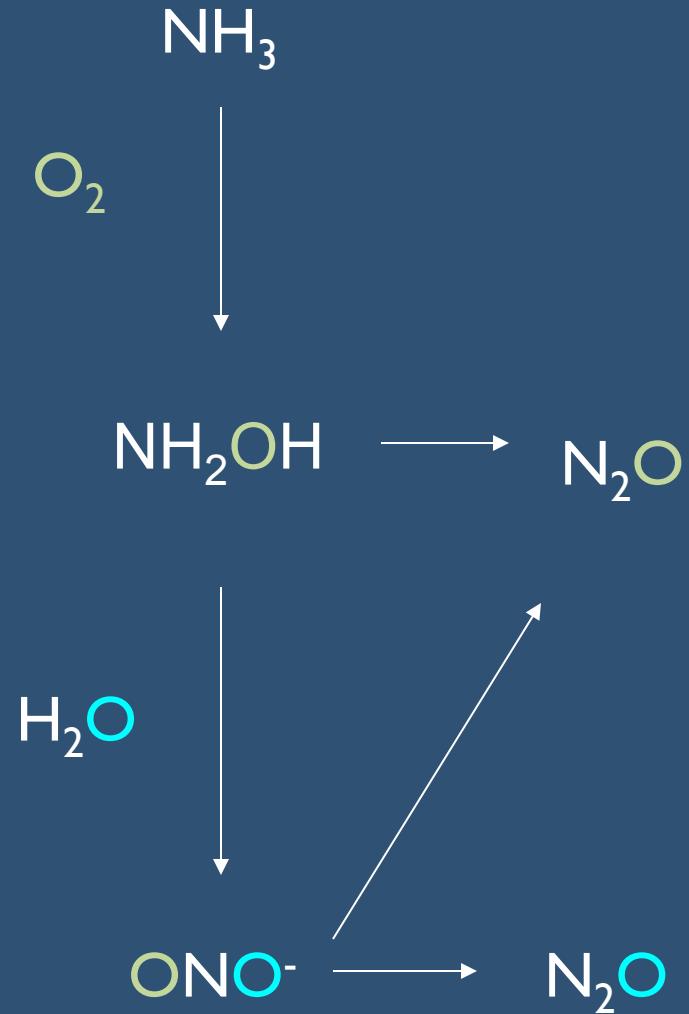
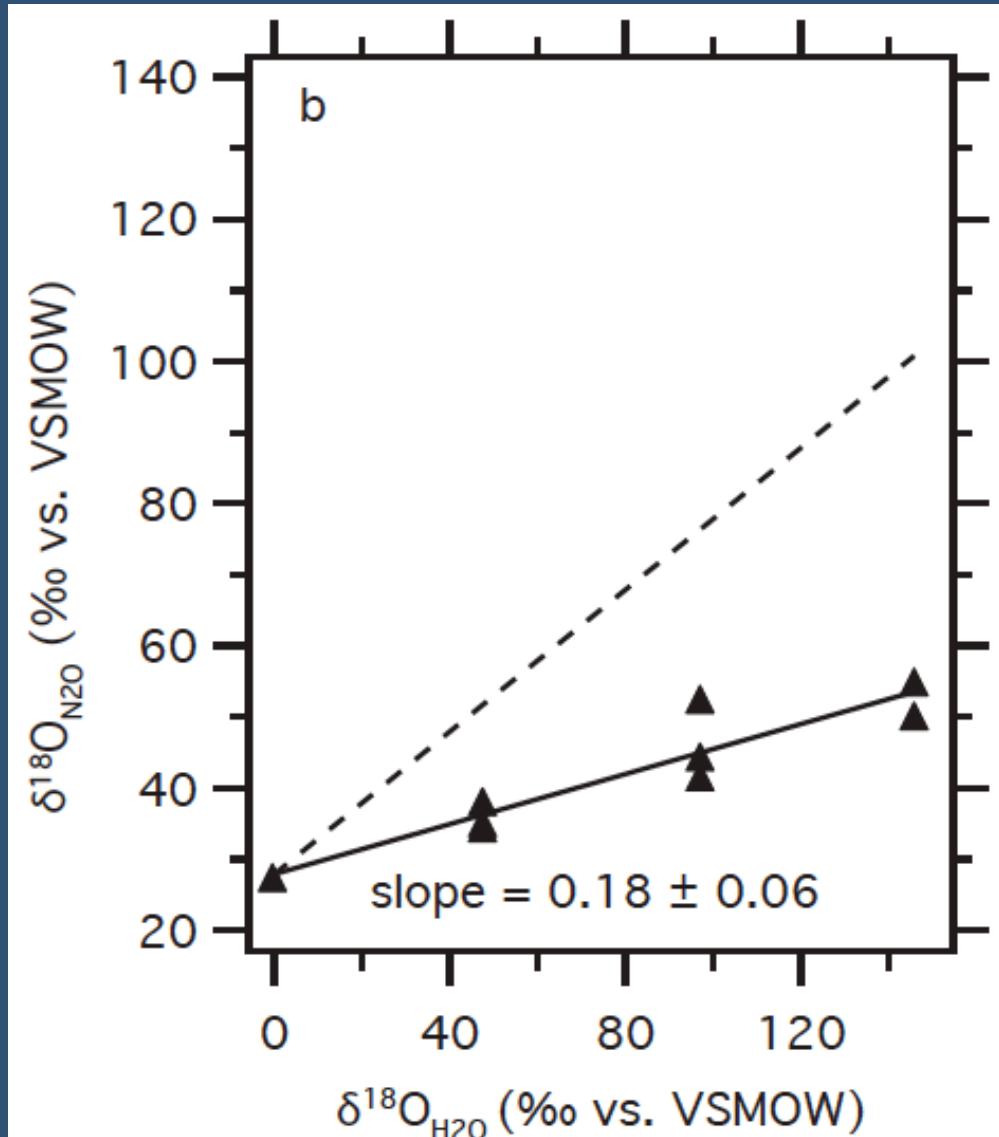
Marine archaea do make N_2O



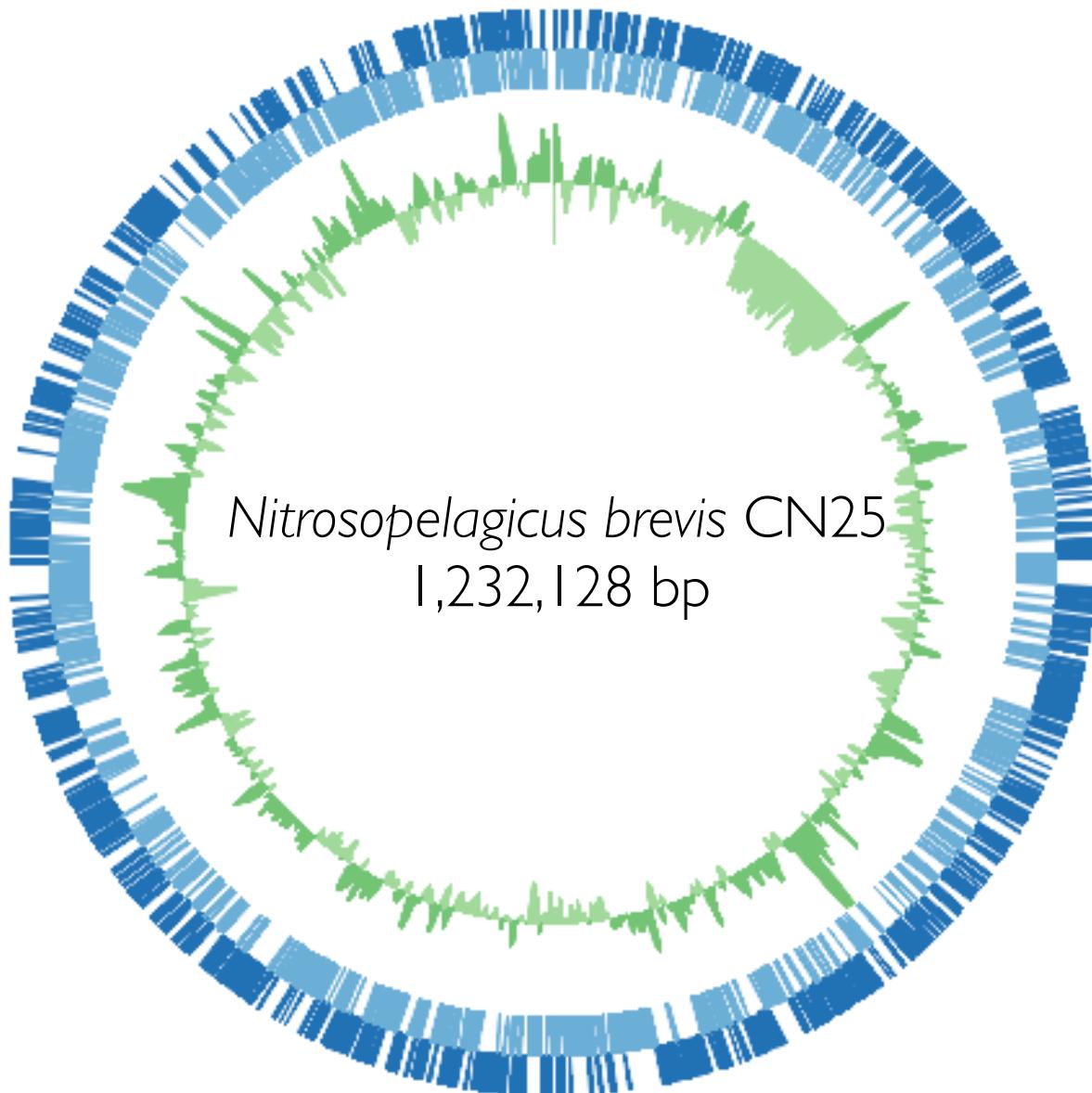
Determining the source of O atoms

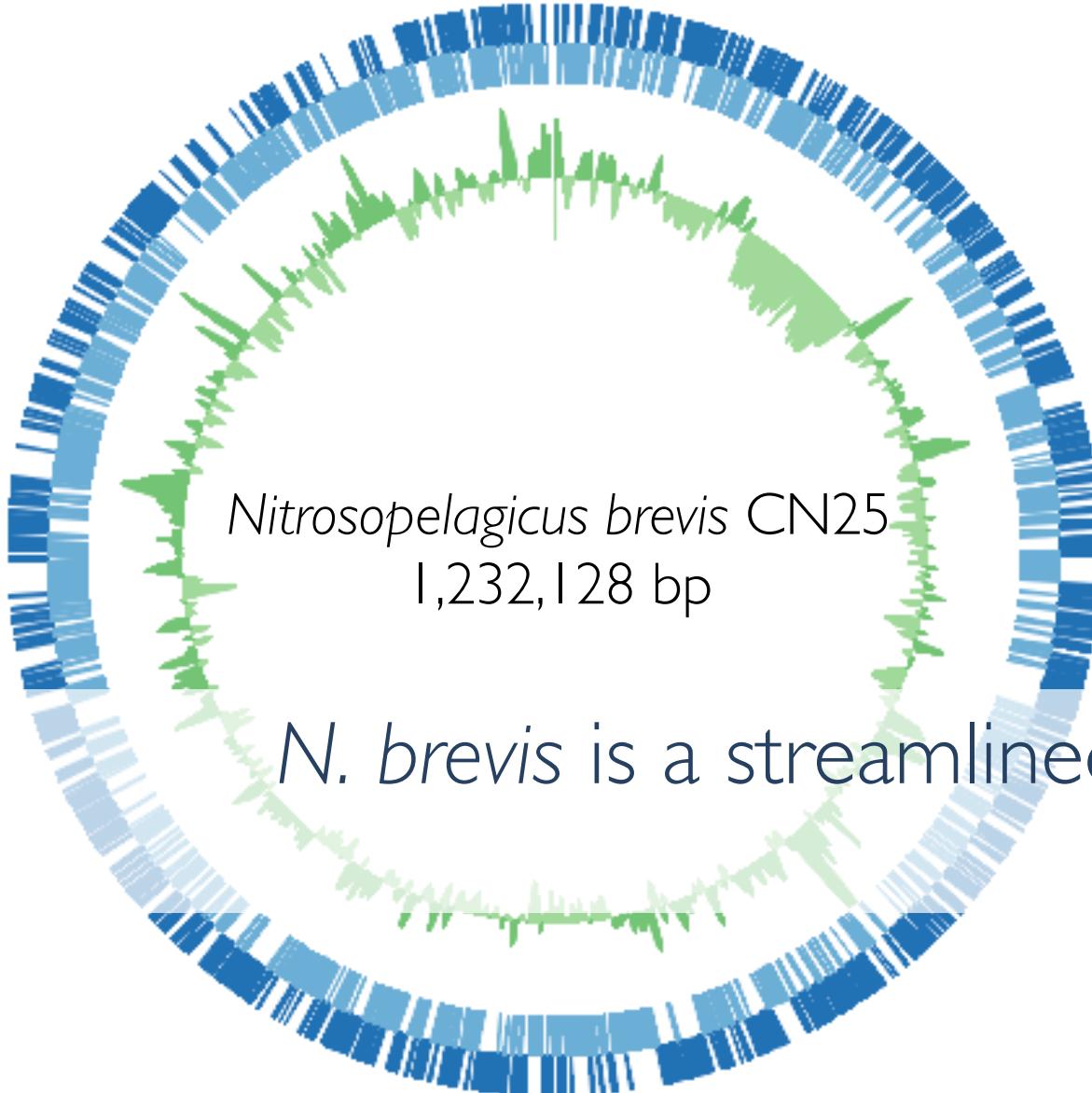


Most of the O in N_2O comes from O_2









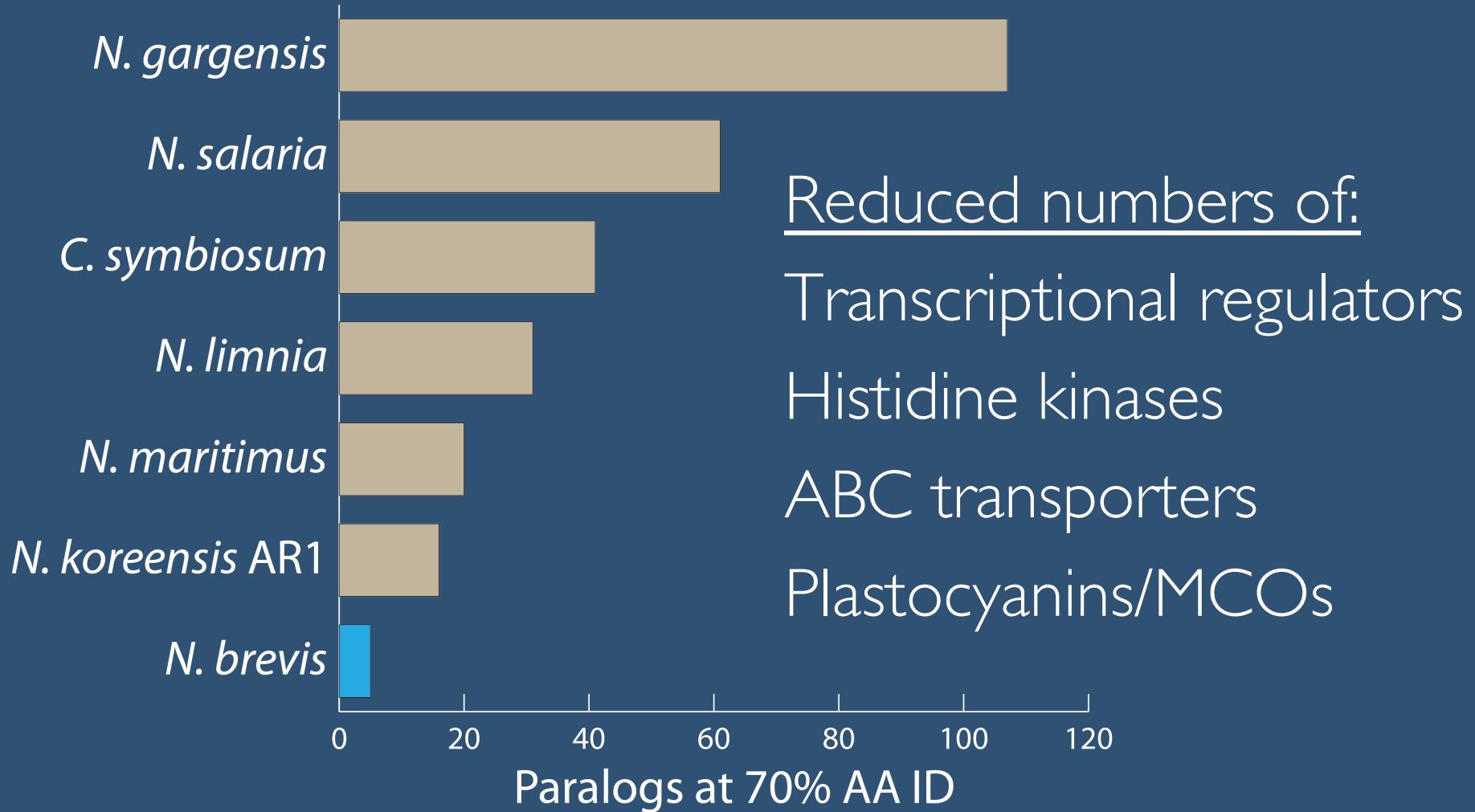
Nitrosopelagicus brevis CN25
1,232,128 bp

N. brevis is a streamlined archaeal genome.

Small genomes are features of other model oligotrophic microbes

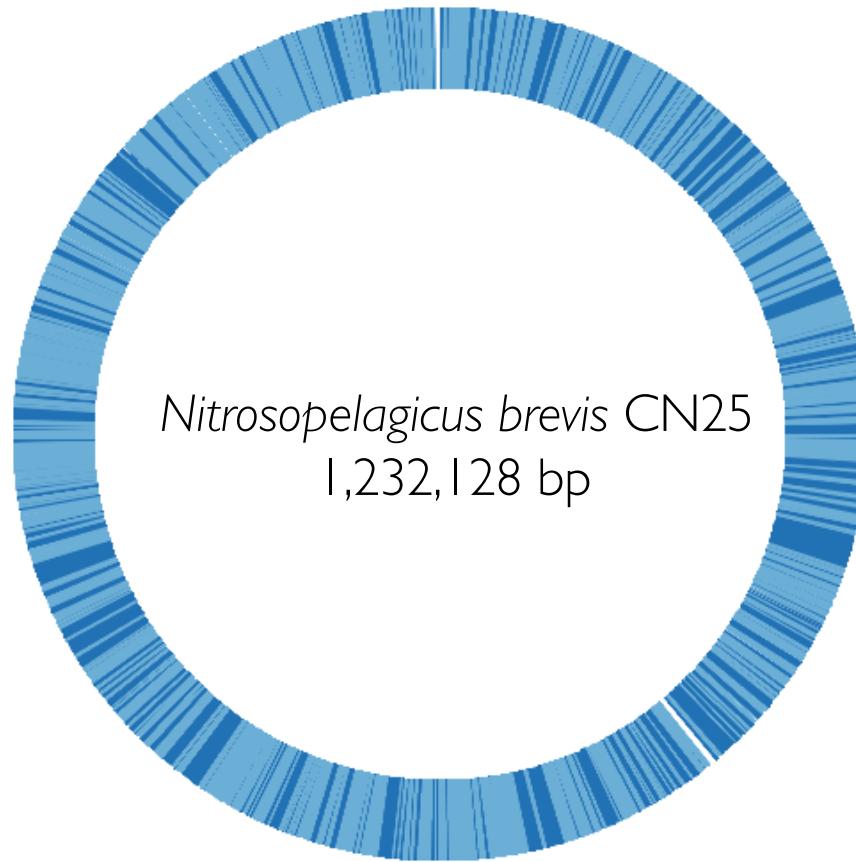
	Size (Mbp)	%GC	# of genes
<i>N. brevis</i>	1.23	33	1501
<i>N. maritimus</i>	1.65	34	1842
<i>Prochlorococcus marinus</i> AS9601	1.67	31	1988
<i>Pelagibacter ubique</i> HTCC1062	1.31	30	1394
<i>Methylophilales</i> sp. HTCC2181 (OM43)	1.30	38	1377
SAR86	1.7	33	1712

N. brevis cuts out redundancy

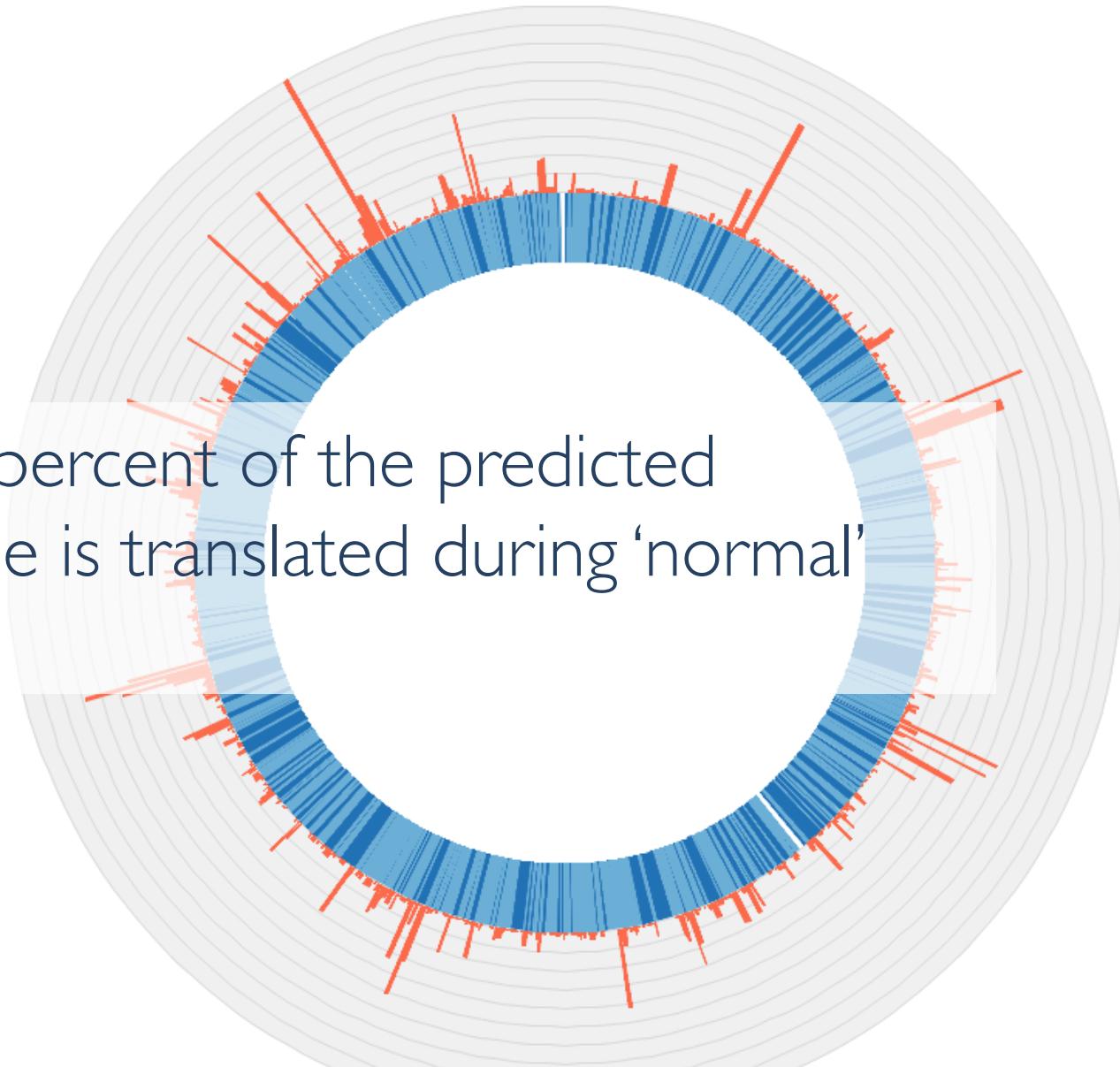


Despite its small genome, *N. brevis* makes its own vitamins and cofactors:

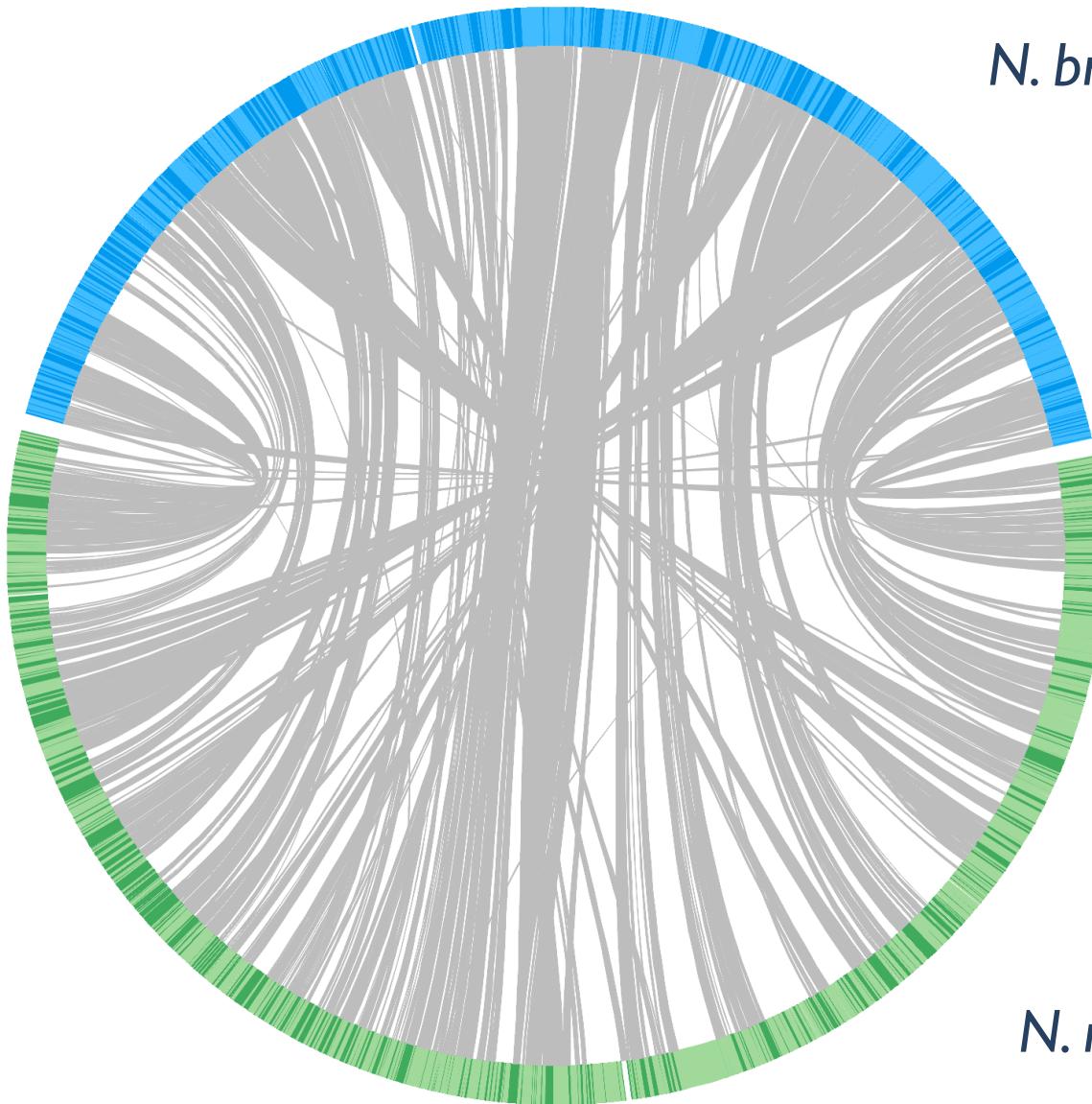
- Vitamin B₁₂
- Vitamins B₁, B₂, B₅, B₇
- Cofactor F₄₂₀
- All amino acids



Seventy percent of the predicted proteome is translated during 'normal' growth.



Proteome data: Mak Saito, Matt McIlvin, Dawn Moran (WHOI)



N. brevis

N. maritimus

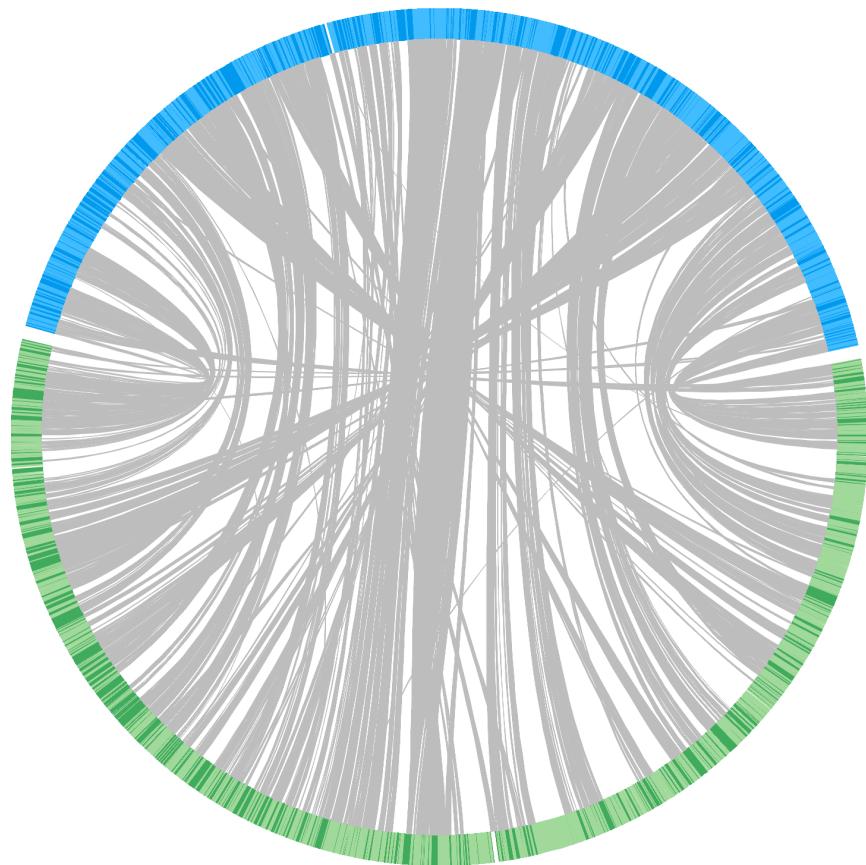
Full genome comparison with other thaumarchaea underscores significant differences

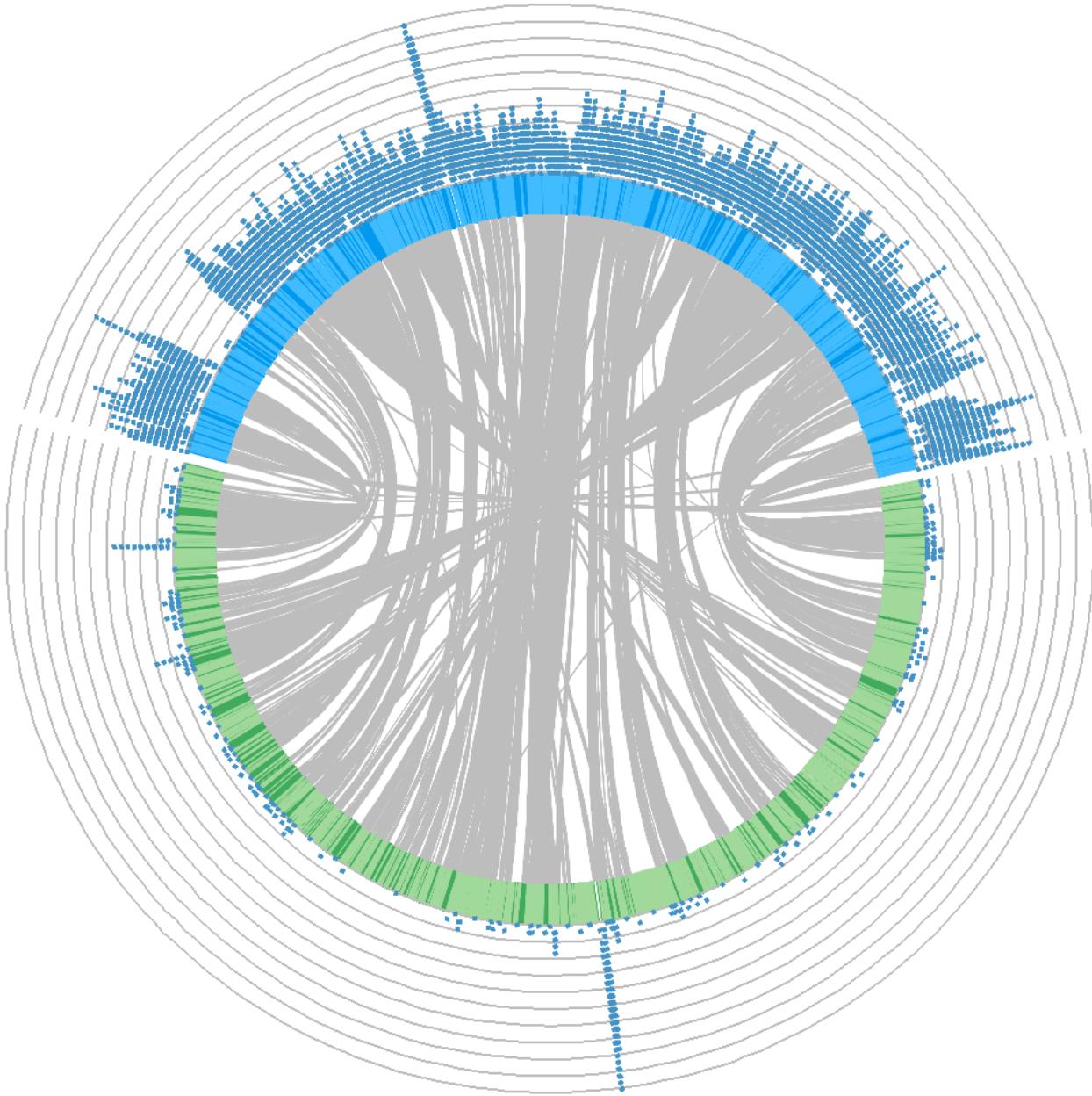
	<i>Cenarchaeum</i>	<i>N. limnia</i>	<i>N. koreensis</i>	<i>N. gargensis</i>	<i>N. maritimus</i>	<i>N. brevis</i>
<i>Cenarchaeum</i>	100					
<i>N. limnia</i>	59	99				
<i>N. koreensis</i>	58	78	99			
<i>N. gargensis</i>	53	63	62	99		
<i>N. maritimus</i>	64	76	82	38	100	
<i>N. brevis</i>	57	66	69	34	75	100

Full genome comparison with other thaumarchaea underscores significant differences

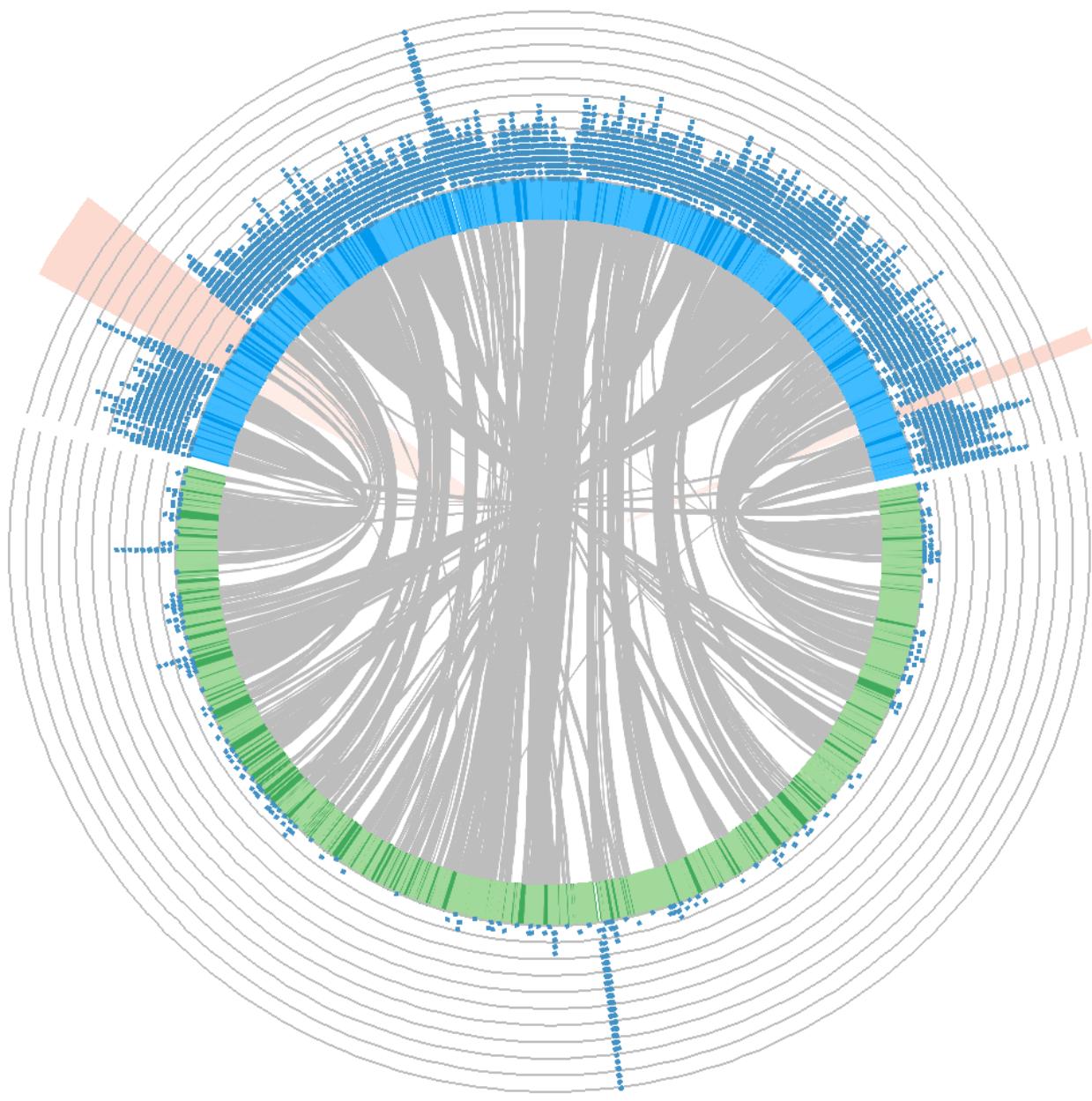
	<i>Cenarchaeum</i>	<i>N. limnia</i>	<i>N. koreensis</i>	<i>N. gargensis</i>	<i>N. maritimus</i>	<i>N. brevis</i>
<i>Cenarchaeum</i>	100					
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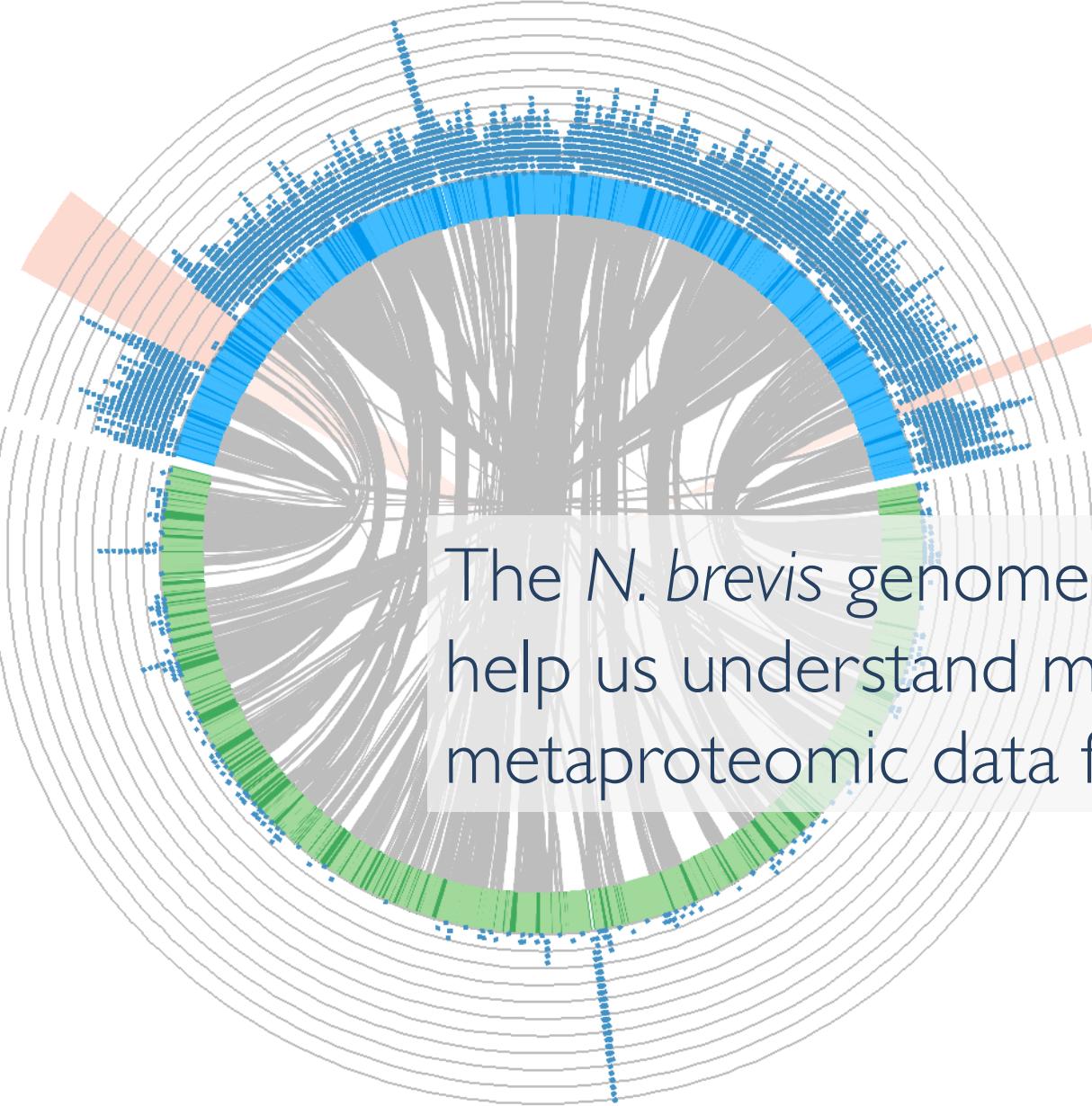
N. brevis averages only 75% ortholog identity with *N. maritimus*.





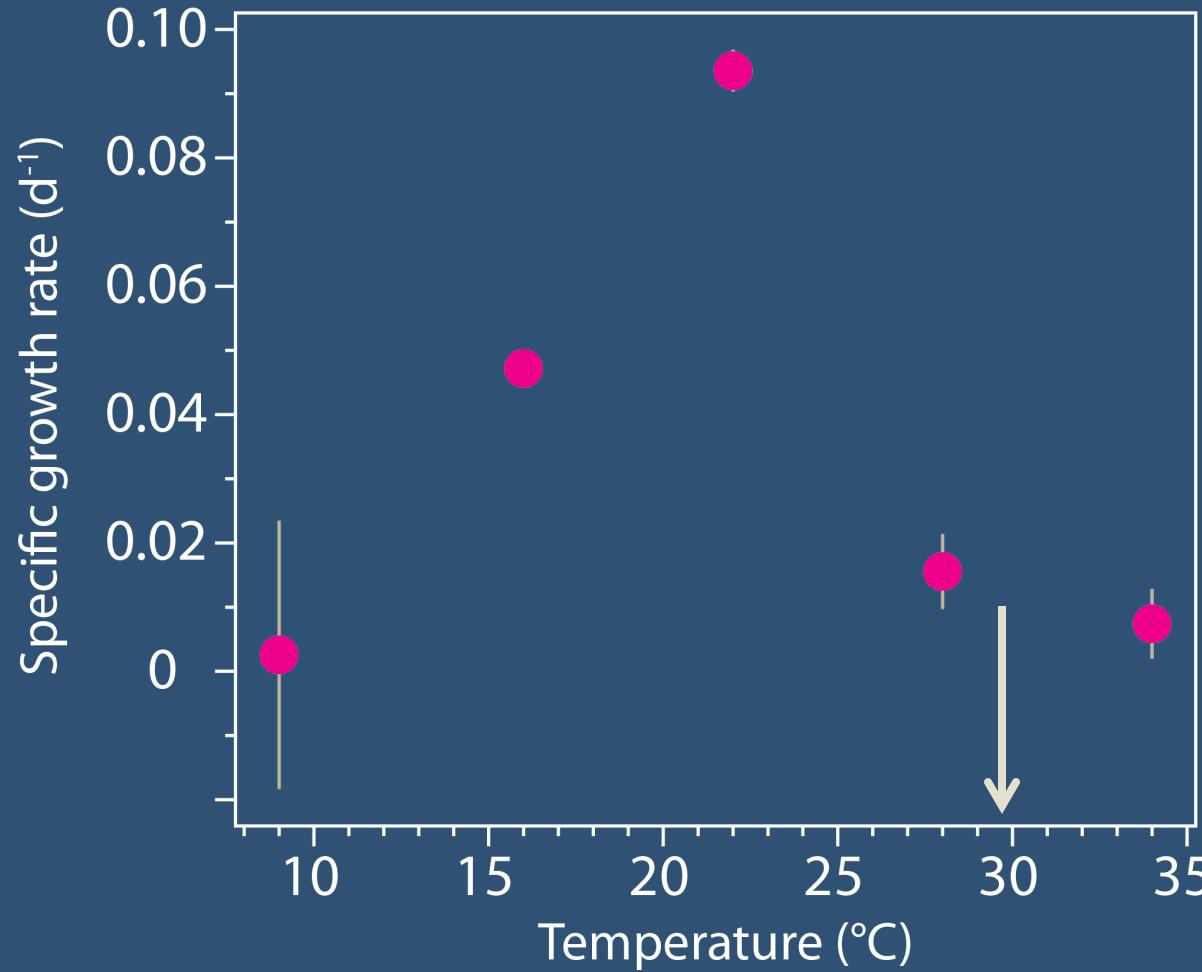
GOS data
mapped at
90%
nucleotide ID



A circular phylogenetic tree diagram showing the relationships between various isolates of *N. brevis*. The tree is composed of numerous small, colored line segments radiating from a central point, forming concentric circles. The colors used include shades of blue, green, orange, and grey. The tree is set against a background of fine, radial grey lines.

The *N. brevis* genome is a new tool to
help us understand metagenomic and
metaproteomic data from the ocean.

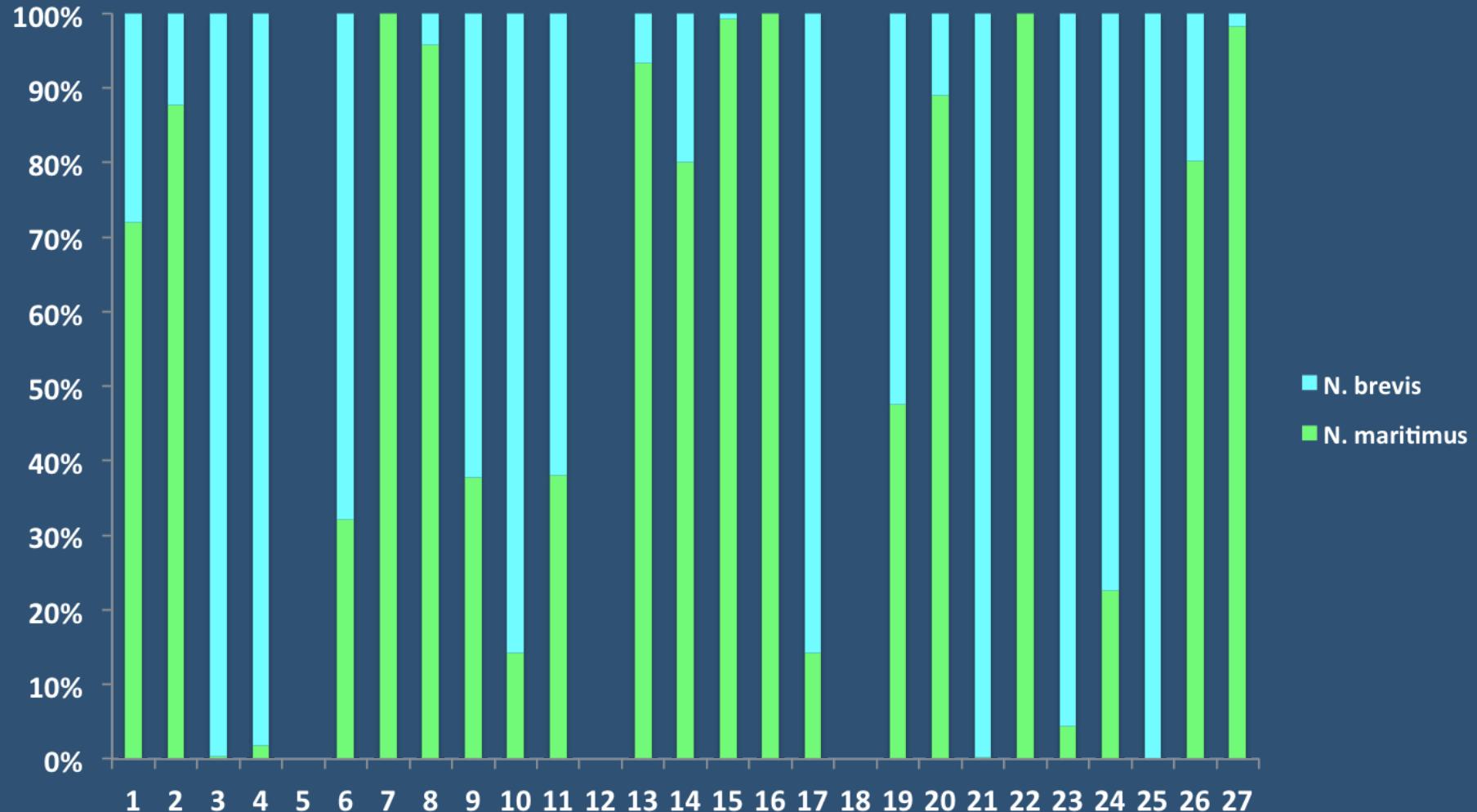
N. brevis temperature optimum is similar
to surface ocean temperatures



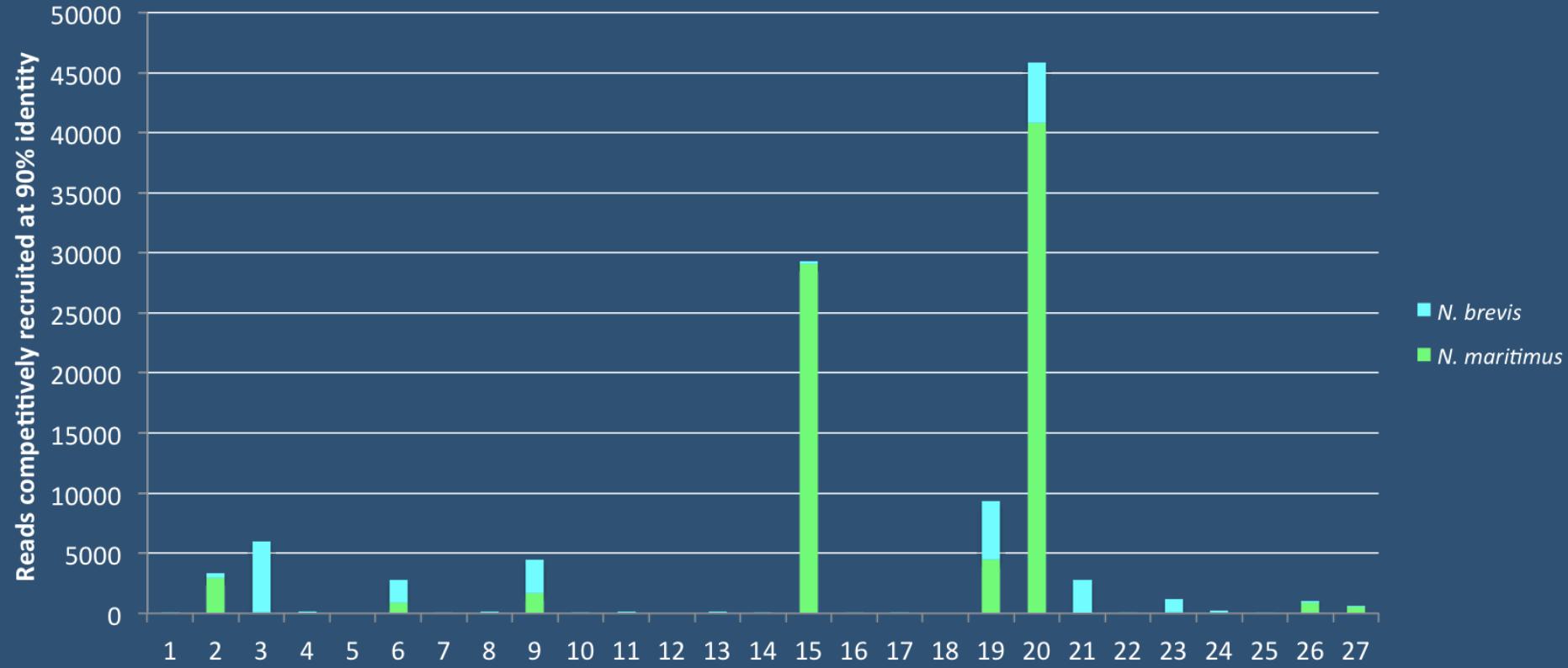
Data: Paul Carini

The GOS data only tell part of the story

The GOS data only tell part of the story



N. maritimus ‘blooms’ in hydrothermal vent plumes and coastal waters



Does 'high' ammonium flux favor *N. maritimus*?

The ISME Journal (2012), 1–11
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www.nature.com/ismej



ORIGINAL ARTICLE

Genome-enabled transcriptomics reveals archaeal populations that drive nitrification in a deep-sea hydrothermal plume

Brett J Baker¹, Ryan A Lesniewski^{1,4} and Gregory J Dick^{1,2,3}

¹Department of Earth and Environmental Sciences, University of Michigan, Ann Arbor, MI, USA;

²Department of Ecology and Evolutionary Biology, University of Michigan, Ann Arbor, MI, USA and

³Center for Computational Medicine and Bioinformatics, University of Michigan, Ann Arbor, MI, USA

www.nature.com/ismej

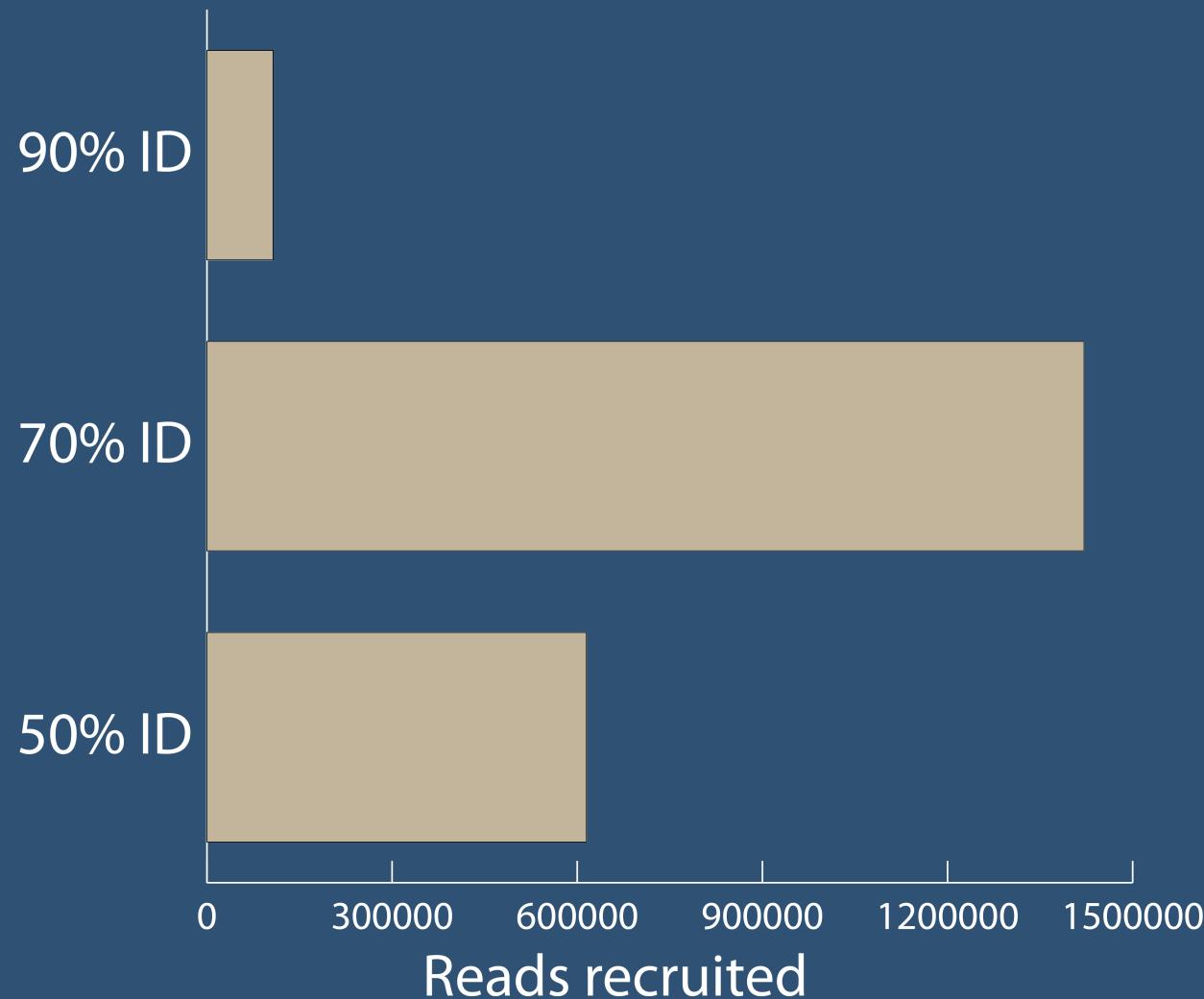
ORIGINAL ARTICLE

Metatranscriptomic analysis of ammonia-oxidizing organisms in an estuarine bacterioplankton assemblage

James T Hollibaugh¹, Scott Gifford¹, Shalabh Sharma, Nasreen Bano and Mary Ann Moran¹

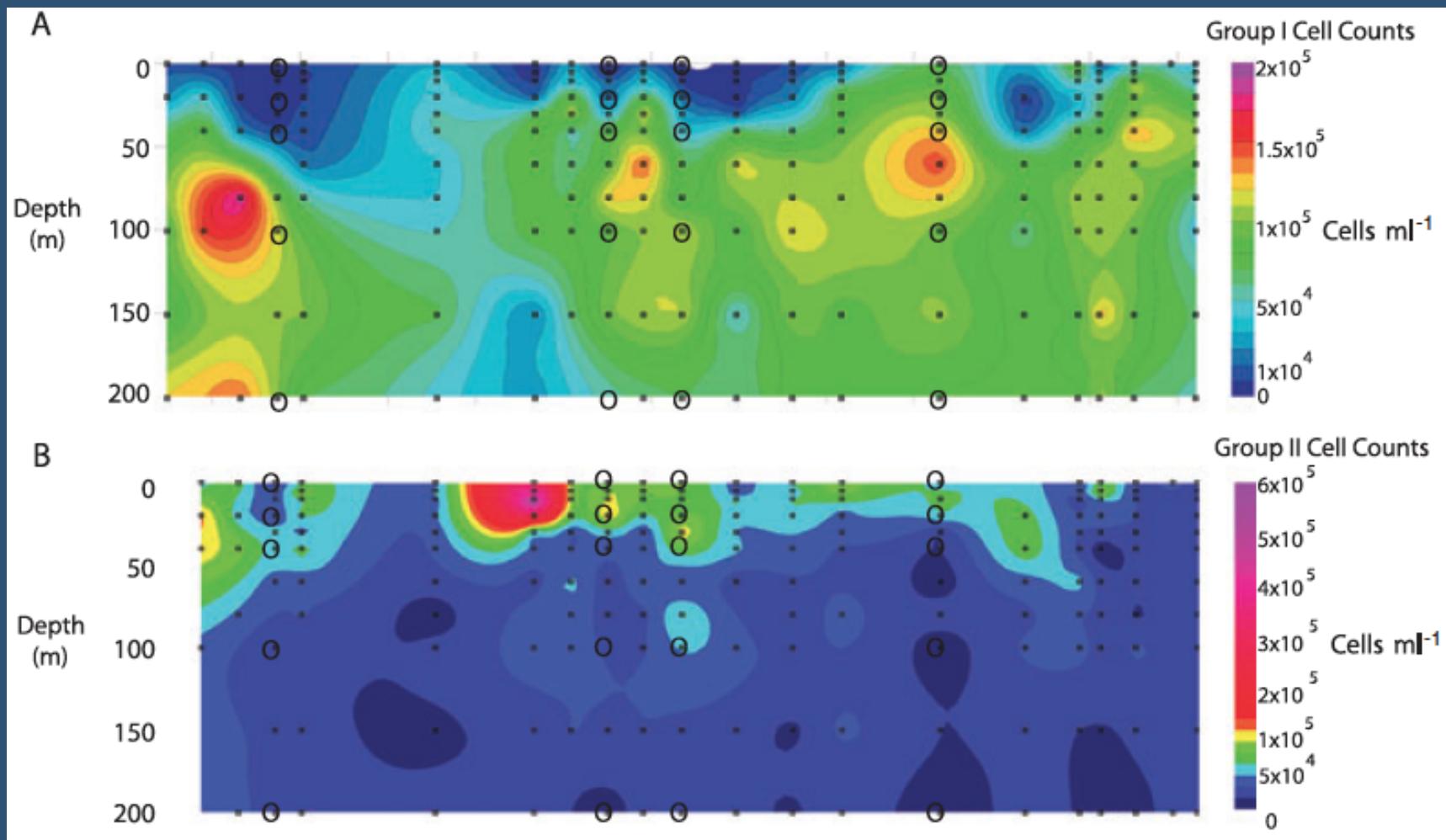
¹Department of Marine Sciences, University of Georgia, Athens, GA, USA

There are a lot of thaumarchaea in the ocean
that aren't *N. brevis* or *N. maritimus*

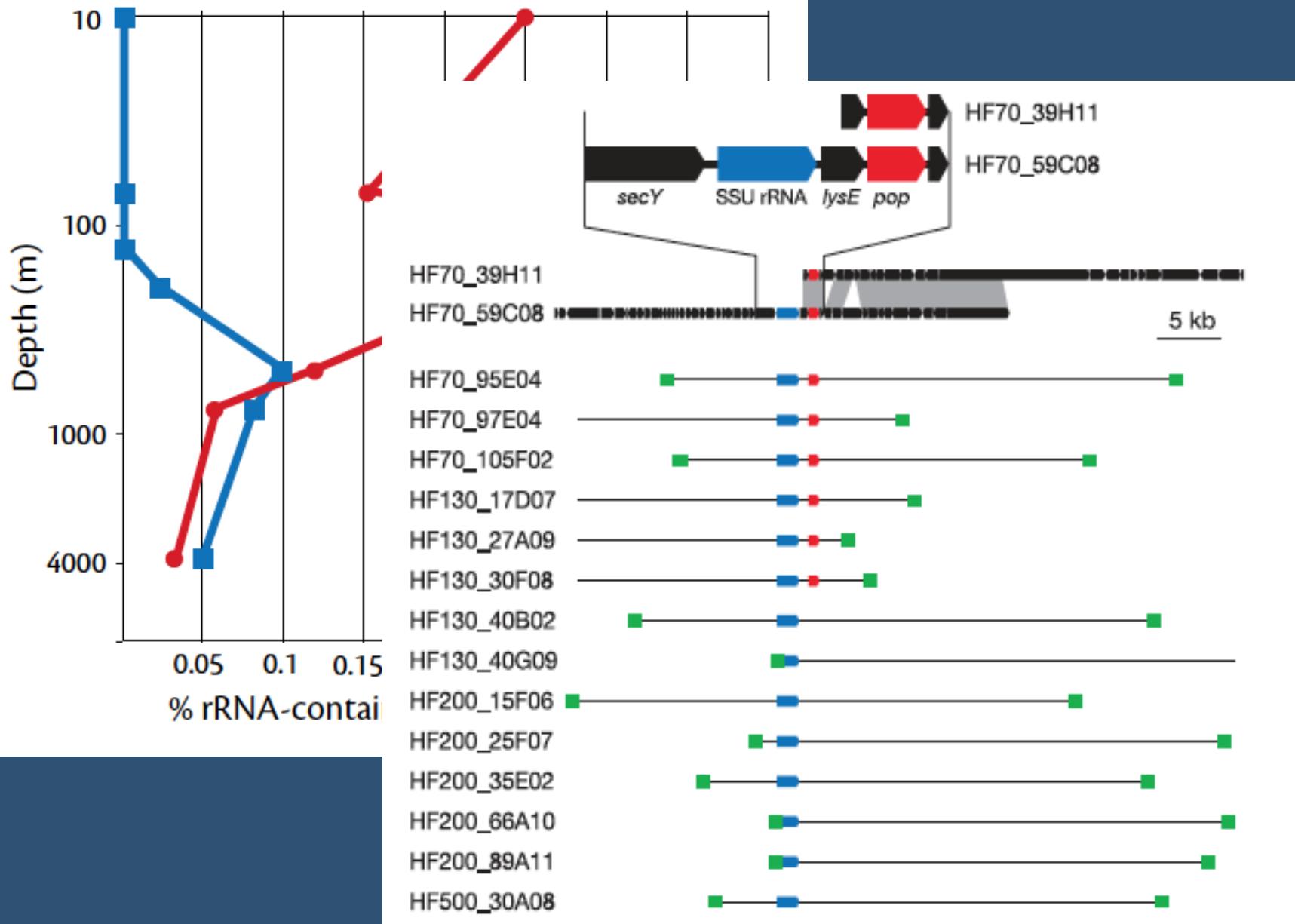


The elusive Euryarchaeota

Contrasting distribution of MGI and MGII



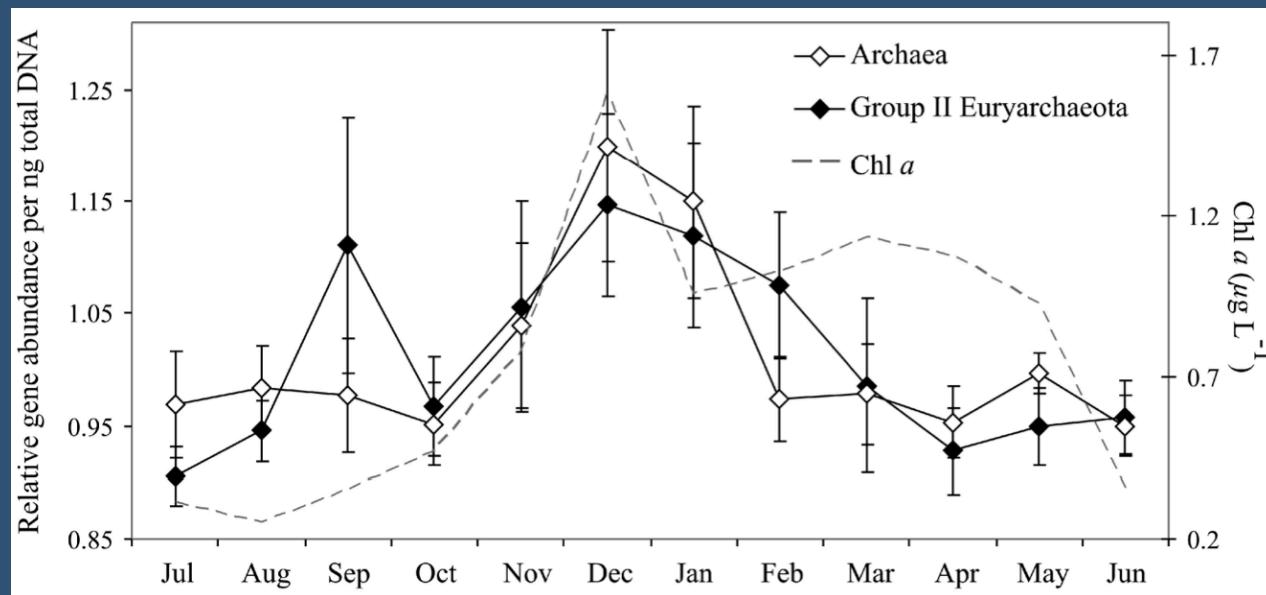
C



Frigaard et al. 2006; Delong et al. 2007

Marine group II Euryarchaeota (MGI) abundant and uncultured

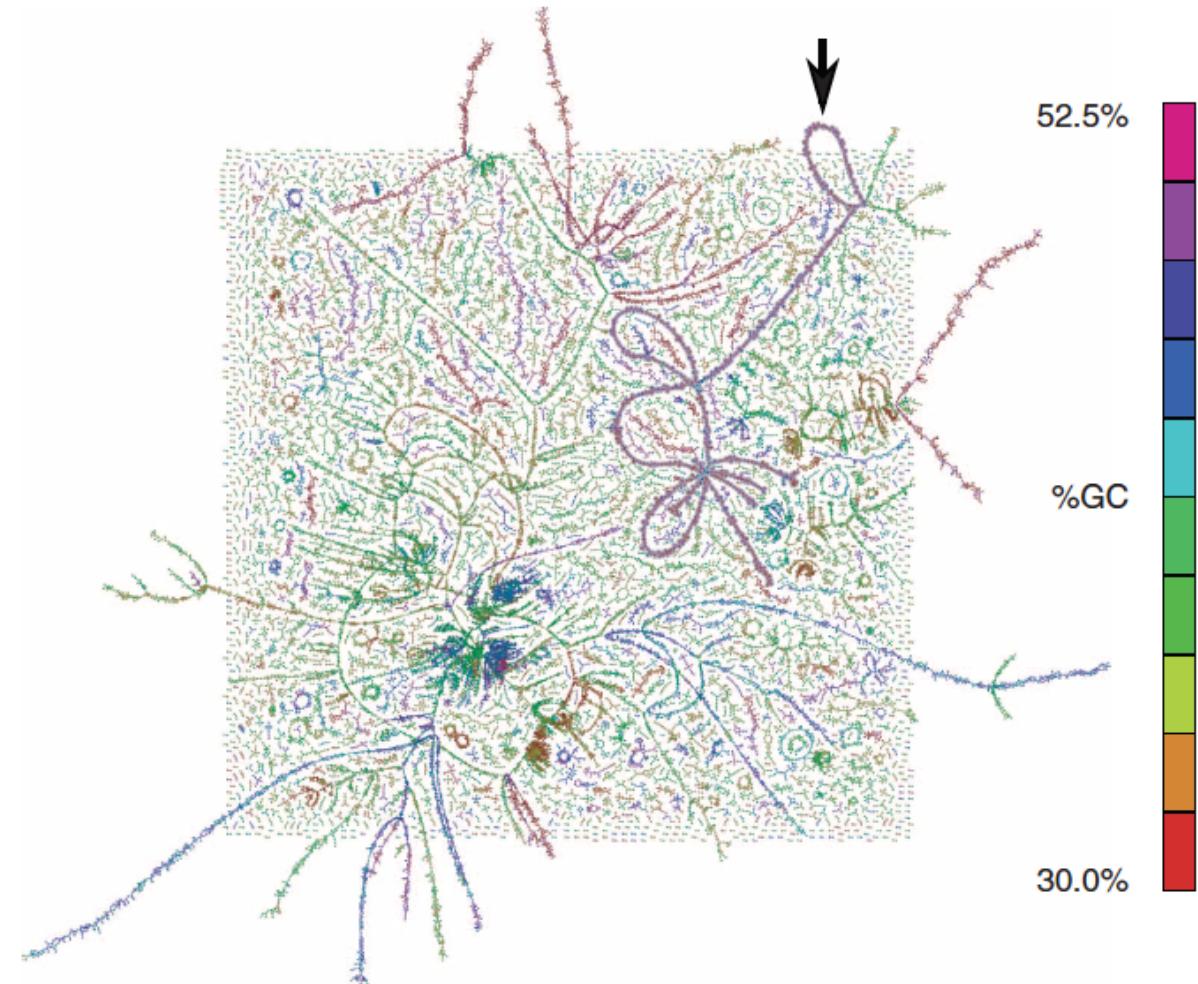
- Can reach abundances of $>10^4$ cells mL⁻¹, found in coastal, open ocean, and Arctic habitats
- Contain proteorhodopsin, potential photoheterotrophs
- Have been found to correlate with Chl *a* concentrations



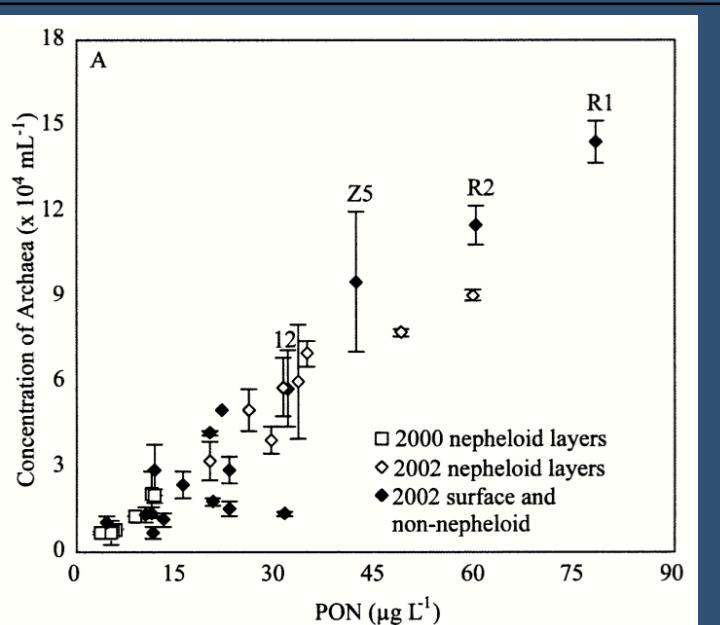
Galand et al. 2010

Untangling Genomes from Metagenomes: Revealing an Uncultured Class of Marine Euryarchaeota

Vaughn Iverson, Robert M. Morris, Christian D. Frazar, Chris T. Berthiaume,
Rhonda L. Morales, E. Virginia Armbrust*



Archaea have been detected on POM and PON



Wells *et al.*, 2006 L&O

Heterogeneous archaeal communities in the particle-rich environment of an arctic shelf ecosystem

Pierre E. Galand ^{a,*}, Connie Lovejoy ^a, Jérémie Pouliot ^a, Warwick F. Vincent ^b

^a Département de Biologie et Québec-Océan, Université Laval, Québec, QC G1V 0A6, Canada

^b Département de Biologie et Centre d'Études Nordiques, Université Laval, Québec, QC G1V 0A6, Canada

Received 10 April 2007; received in revised form 11 November 2007; accepted 7 December 2007

Available online 14 December 2007

Archaeaplankton in the Columbia River, its estuary and the adjacent coastal ocean, USA

Byron C. Crump ^{*}, John A. Baross

School of Oceanography, University of Washington, Seattle, WA 98195, USA

Received 7 September 1999; received in revised form 13 December 1999; accepted 13 December 1999

Questions:

What are MGII doing on particles?

Are there interactions between MGII and picoeukaryotes?

Is there a link between POM and MGII ecology?

Collaborators:

Mak Saito

Chris Dupont

Culture and genomic work:

NSF Bio OCE

UMCES start up funds

WHOI Ocean Life

Institute

Proteomics and MGII:

Gordon and Betty
Moore Foundation



Acknowledgements

Collaborators:

Jason Smith (MBARI)
Jarred Swalwell (UW)
Susanne Wilken (MBARI)
Heather Wilcox (MBARI)
Alexandra Worden (MBARI)
Patrick Keeling (UBC)
Tom Richards (U. Exeter)

Cruise support:

Western Flyer: Captain and crew
Tim Pennington (MBARI)
Marguerite Blum (MBARI)
Francisco Chavez (MBARI)
Valeria Jimenez (UBC)
Noriko Okamoto (UBC)



Thank you!!!

