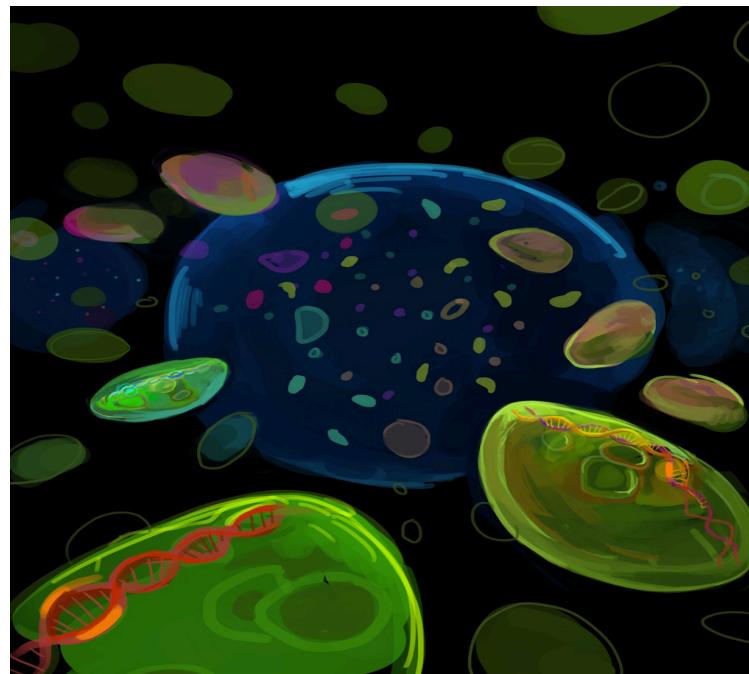


# *Prochlorococcus*

An adventure in Microbial Oceanography  
(or the power of the model system approach)



Carly Sankar, MIT

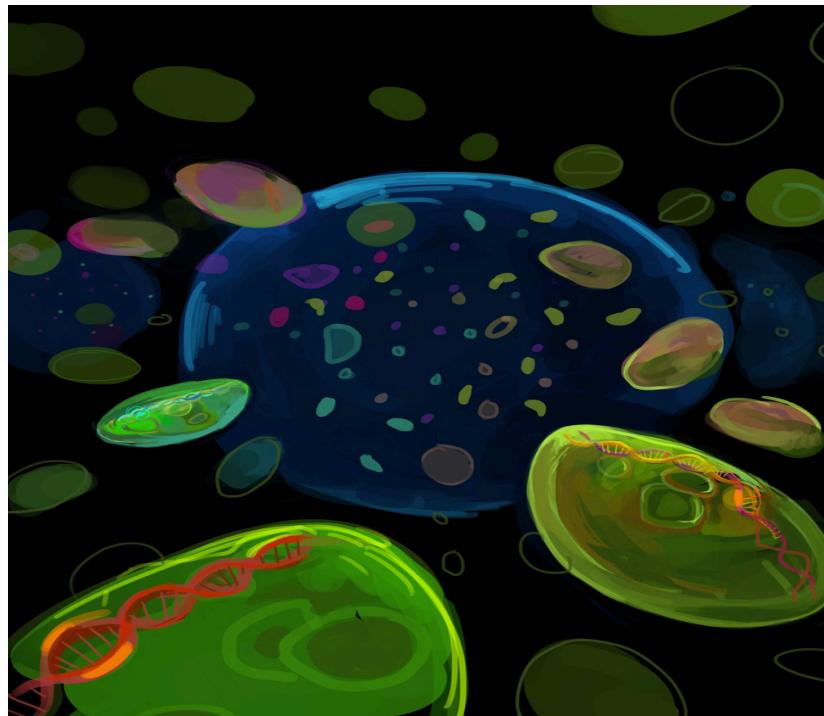
Penny Chisholm

C-MORE Summer Course

June 4, 2014

# *Prochlorococcus*

An adventure in Microbial Oceanography  
(or the power of the model system approach)



Carly Sanker, MIT

A self-centered presentation!

C-MORE Summer Course

June 4, 2014

# Overview

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- ◆ History
- ◆ The Cell
- ◆ Niche Dimensions of *Prochlorococcus*
  - Light and Temperature
  - Genomics and Niche Dimensions:
    - Phosphorus
    - Nitrogen
    - Iron
- ◆ The Community
- ◆ Marine Vesicles
- ◆ Integrative Systems Biology

# ...The BIG discovery - began the paradigm shift

Waterbury et. al. 1979

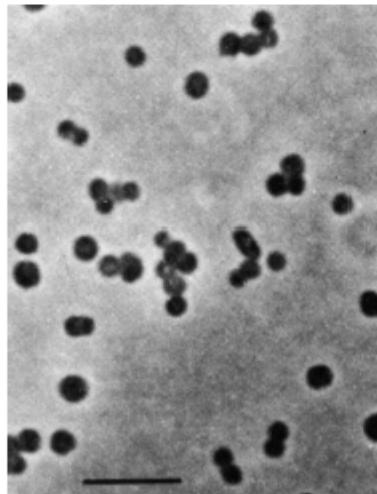


Fig. 1 Phase contrast photomicrograph of *Synechococcus* sp. strain Syn-48 illustrating general cell morphology (scale bar, 5.0  $\mu\text{m}$ ).

## Widespread occurrence of a unicellular, marine, planktonic, cyanobacterium

IN marked contrast to their freshwater counterparts, marine planktonic cyanobacteria are restricted to a few nostocalean genera, of which only *Trichodesmium* is capable of forming extensive water blooms<sup>1-3</sup>. We report here the widespread occurrence of a small, marine, chroococcacean cyanobacterium belonging to the genus *Synechococcus*.

Natural water samples were filtered through 0.2  $\mu\text{m}$  Nuclepore filters, counterstained with Irgalan black<sup>4</sup>. The filters were examined with a Zeiss Standard microscope equipped with Neofluar objectives and an epifluorescent illumination system containing a 100-W halogen lamp, a BP 450–500 excitation filter, a LP 528 barrier filter and a FT 510 chromatic beam splitter. Using this system, phycoerythrin-containing cyanobacteria fluoresce orange and can be distinguished from phytoplankters that fluoresce red.

Phycoerythrin-rich unicellular cyanobacteria were observed at seven stations in the Arabian Sea in January 1977, at three stations off the coast of Peru in March 1978, in Slope Water north of the Gulf Stream in April 1978, and periodically in Woods Hole Harbor. In the relatively rich waters of the Arabian Sea and off the coast of Peru, the population varied from  $10^4$  to  $10^5$  cells  $\text{ml}^{-1}$  within the euphotic zone (Table 1). The greatest number of cells was found within the top 20 m of the water column, with occasional cells being observed as deep as 400 m. In contrast, the surface sample collected from Slope Water north

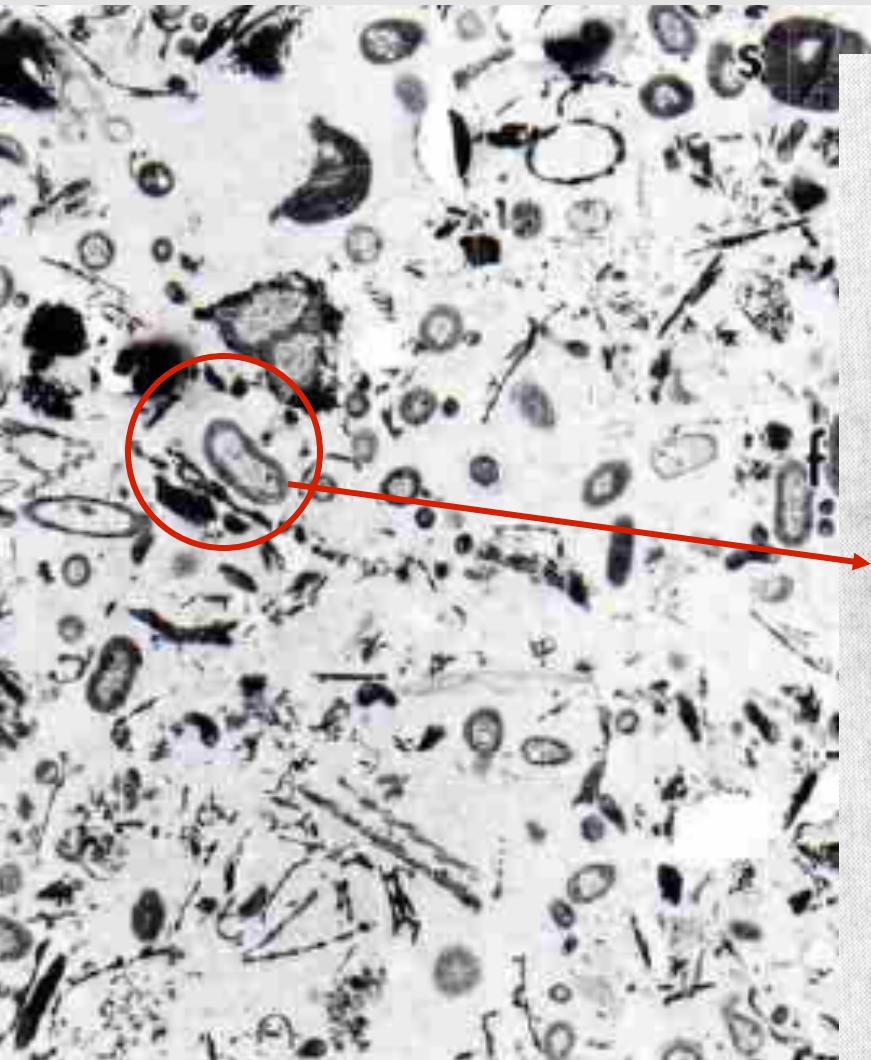
JOHN B. WATERBURY  
STANLEY W. WATSON  
ROBERT R. L. GUILLARD  
LARRY E. BRAND

Department of Biology,  
Woods Hole Oceanographic Institution,  
Woods Hole, Massachusetts 02543

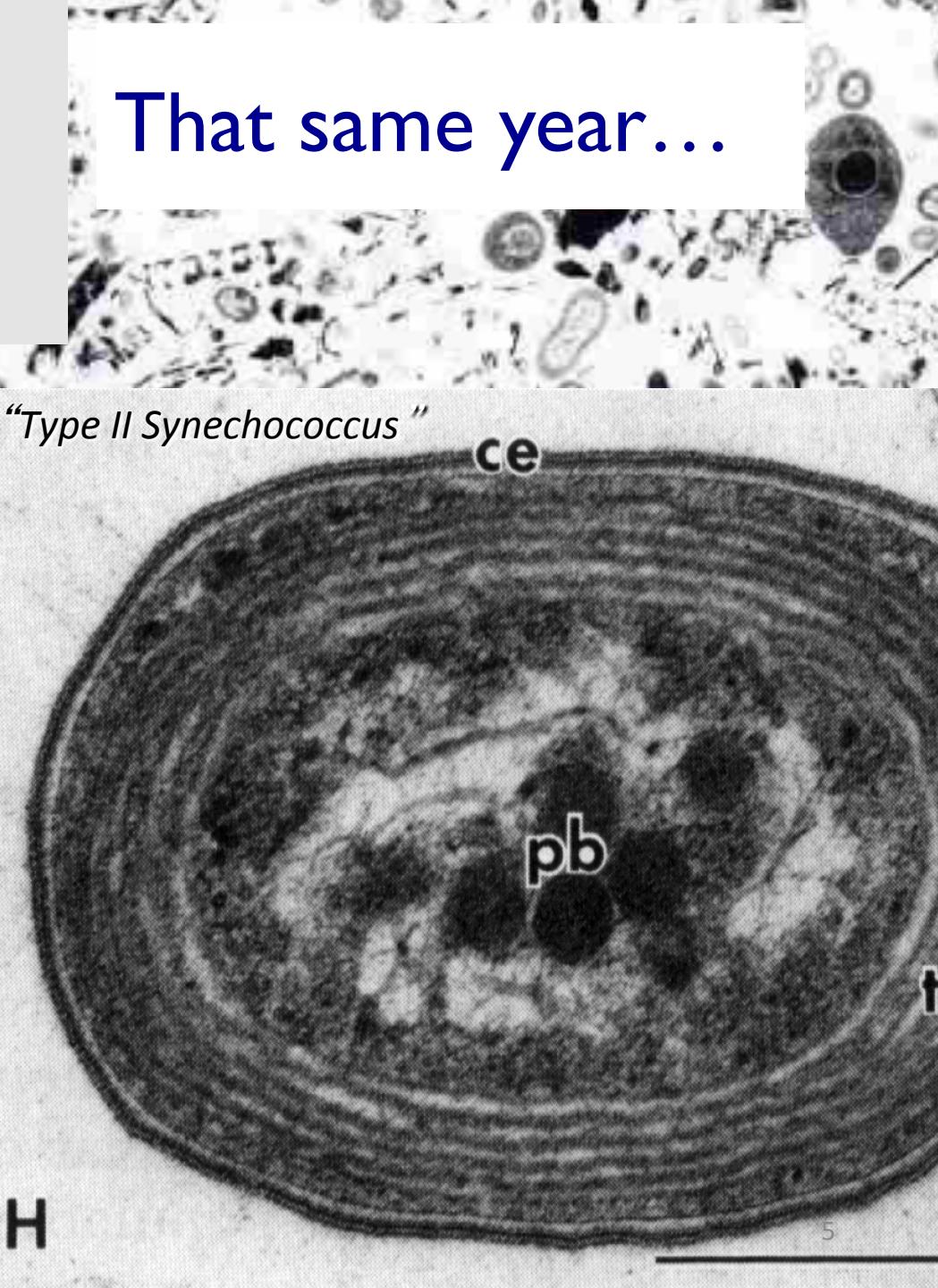
© Macmillan Journals Ltd 1979

*Johnson and Seiburth 1979*

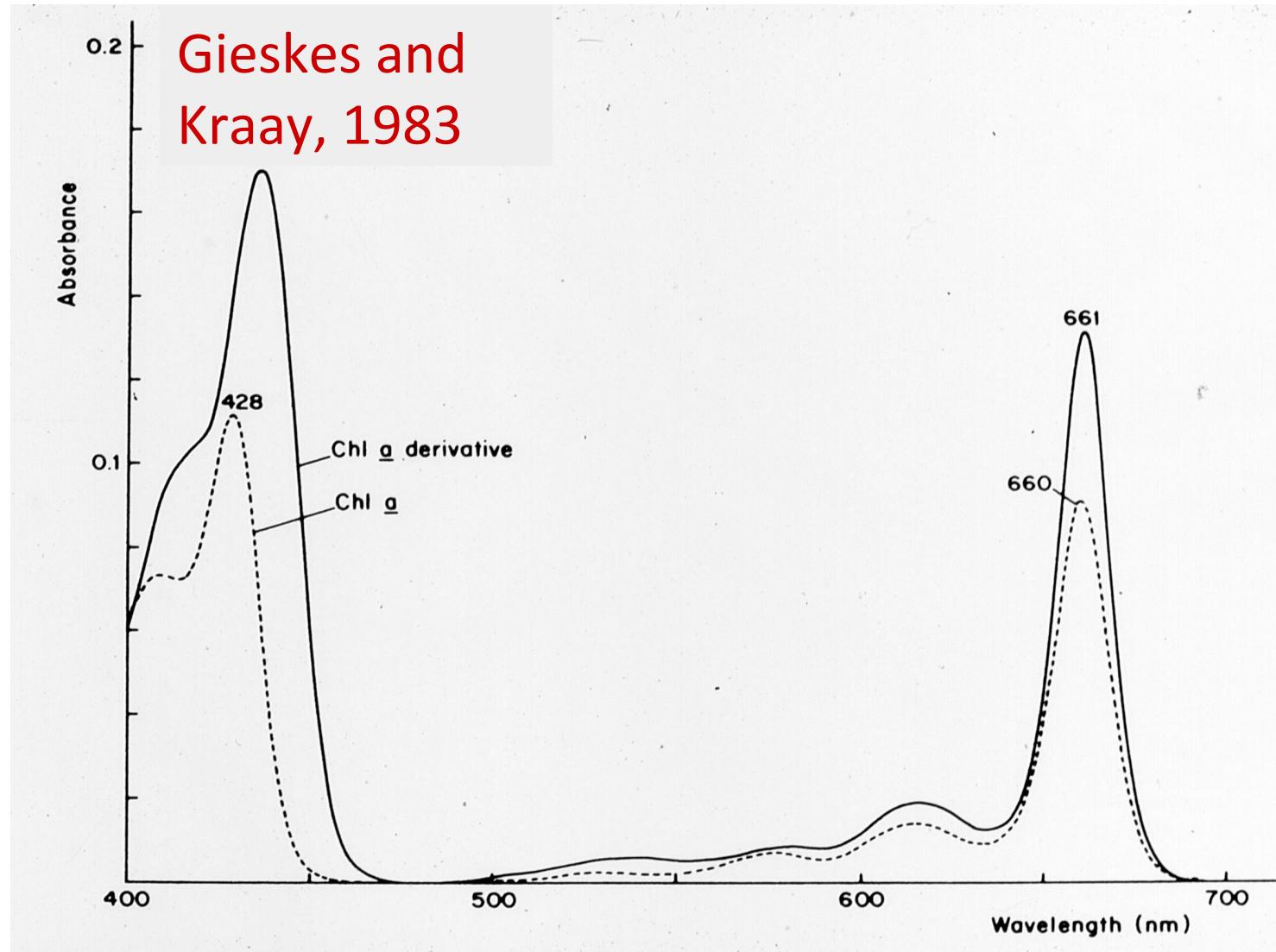
Chroococcoid cyanobacteria in the sea:  
A ubiquitous and diverse phototrophic biomass.  
*Limnology and Oceanography* 24(5):928-935.



That same year...

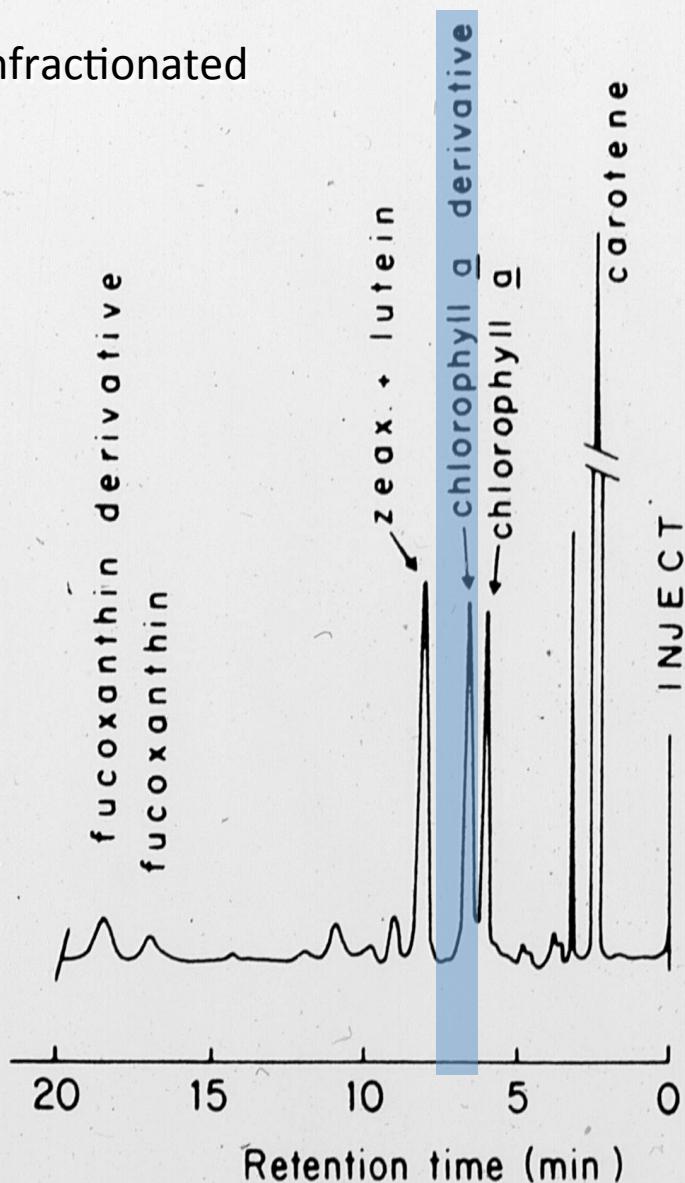


Meanwhile, a few years later, a new pigment was discovered...



And that pigment is enriched in the < 1  $\mu\text{m}$  fraction

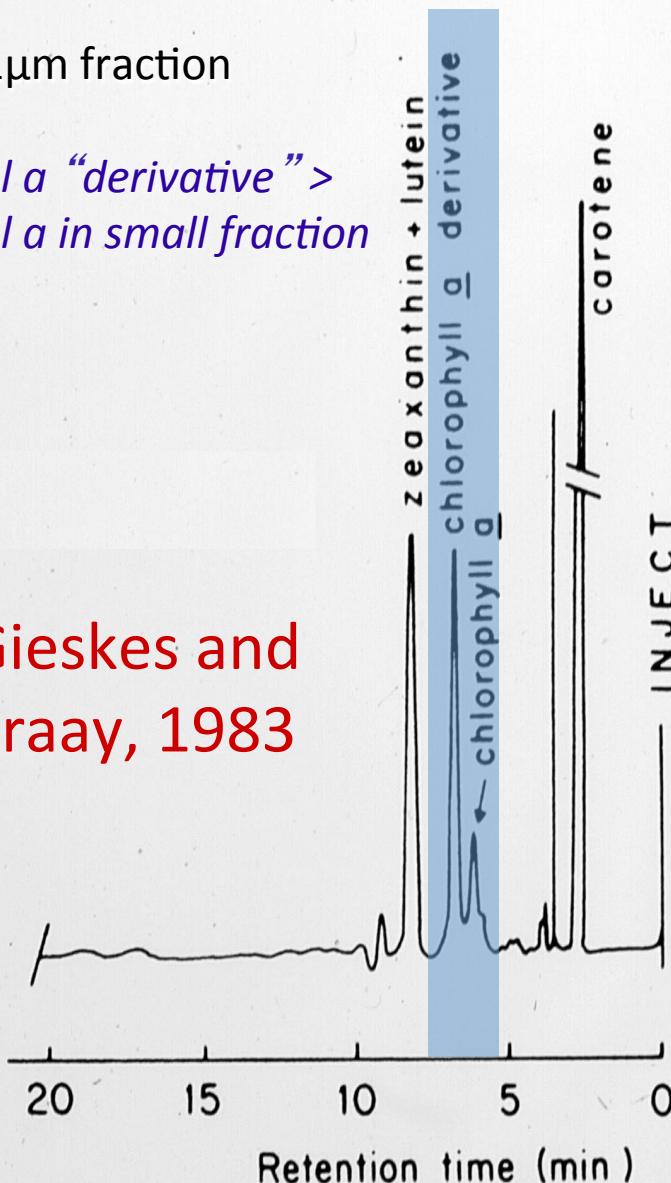
unfractionated



< 1  $\mu\text{m}$  fraction

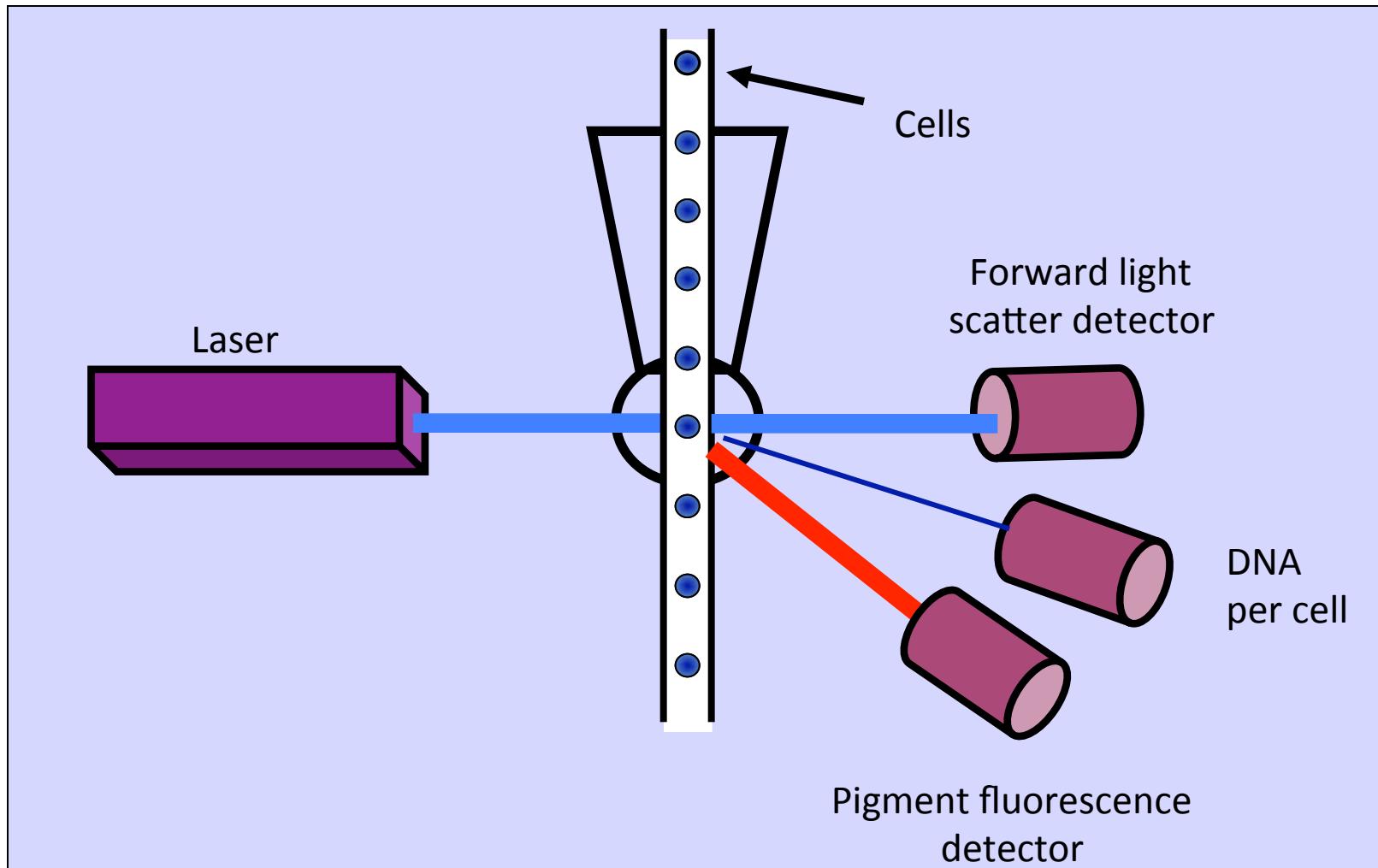
*Chl a “derivative” >  
Chl a in small fraction*

Gieskes and  
Kraay, 1983

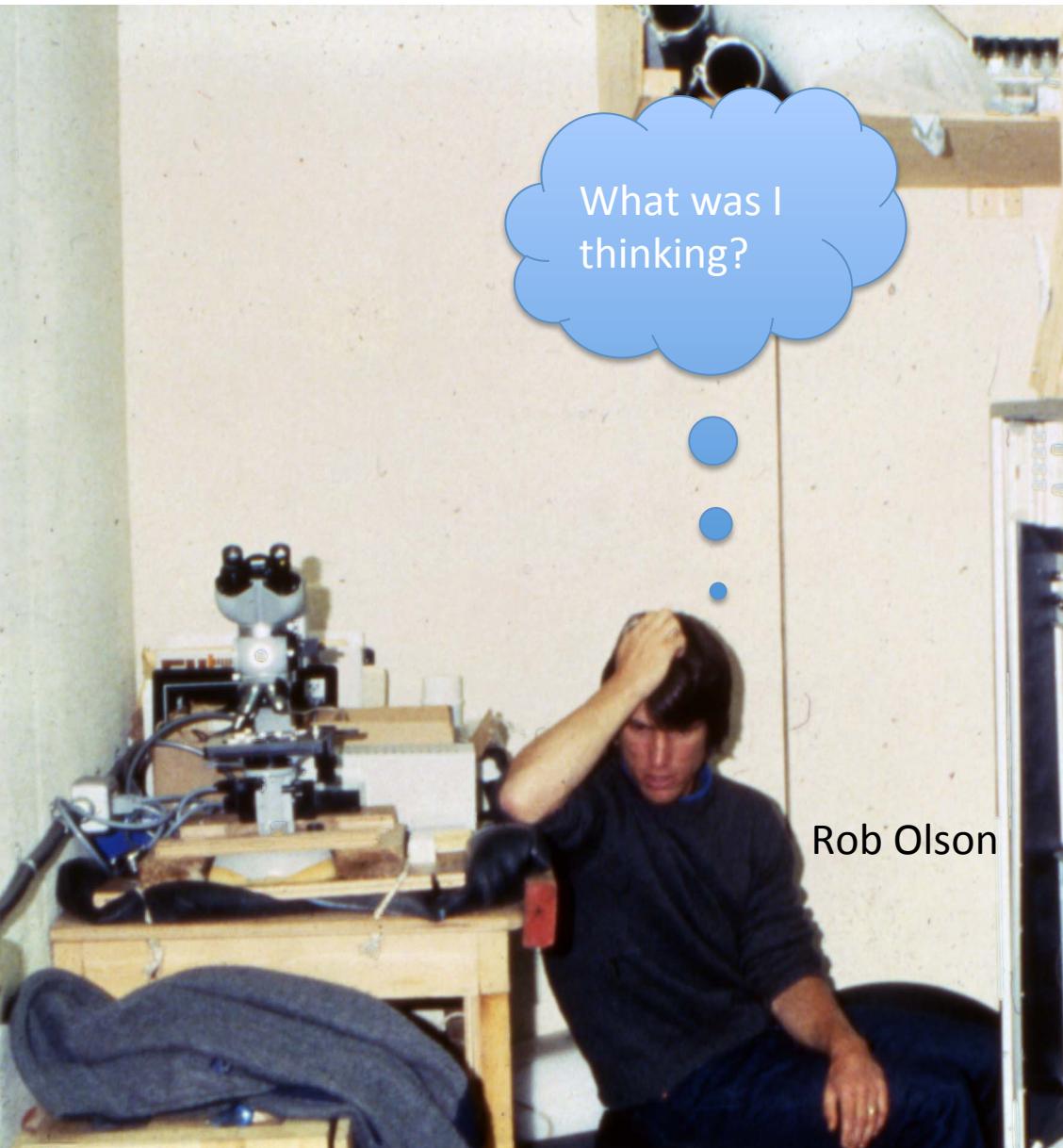


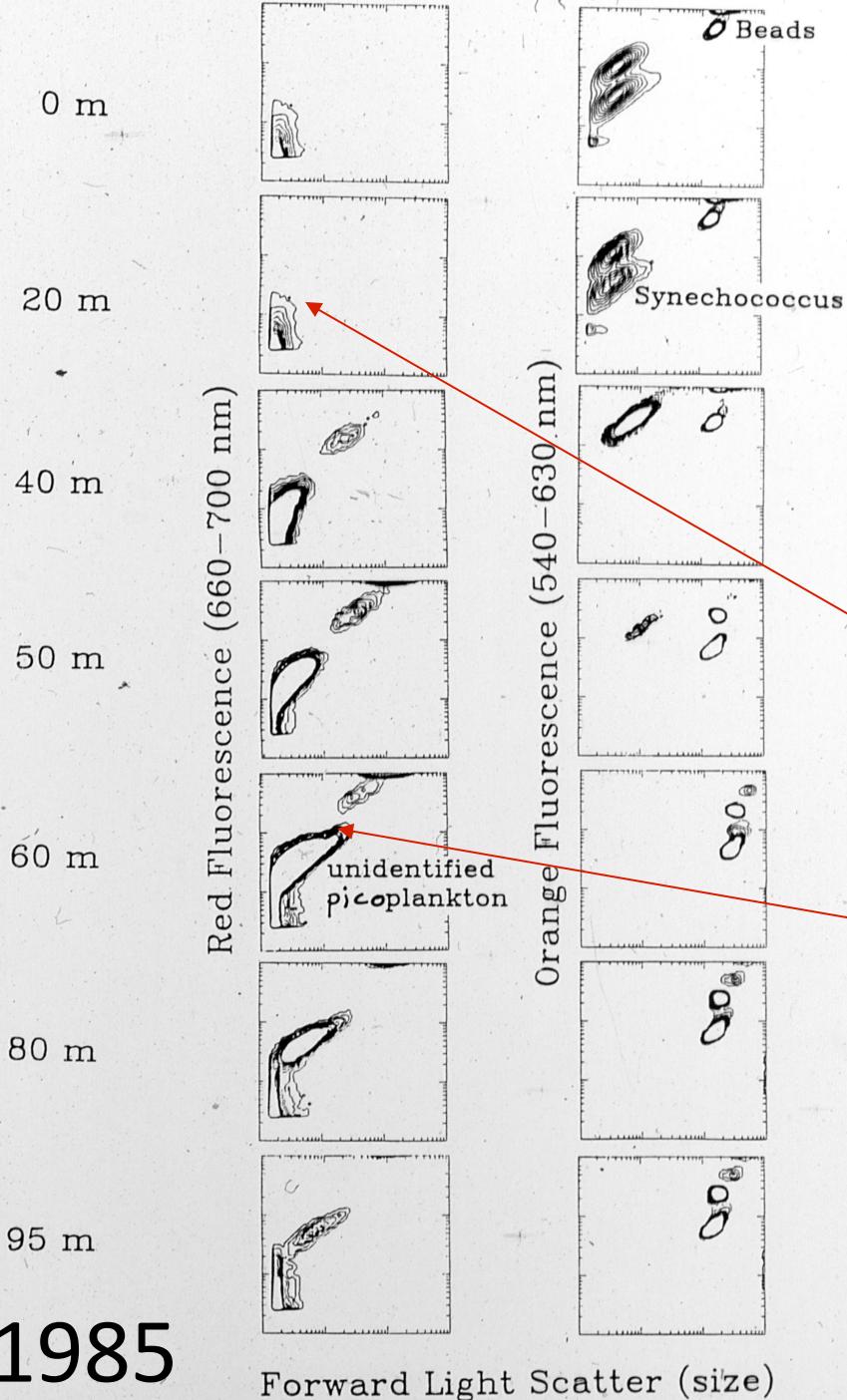
# Enter Flow Cytometry.....

...for the study of single cells



# And someone with the courage to take it to sea...

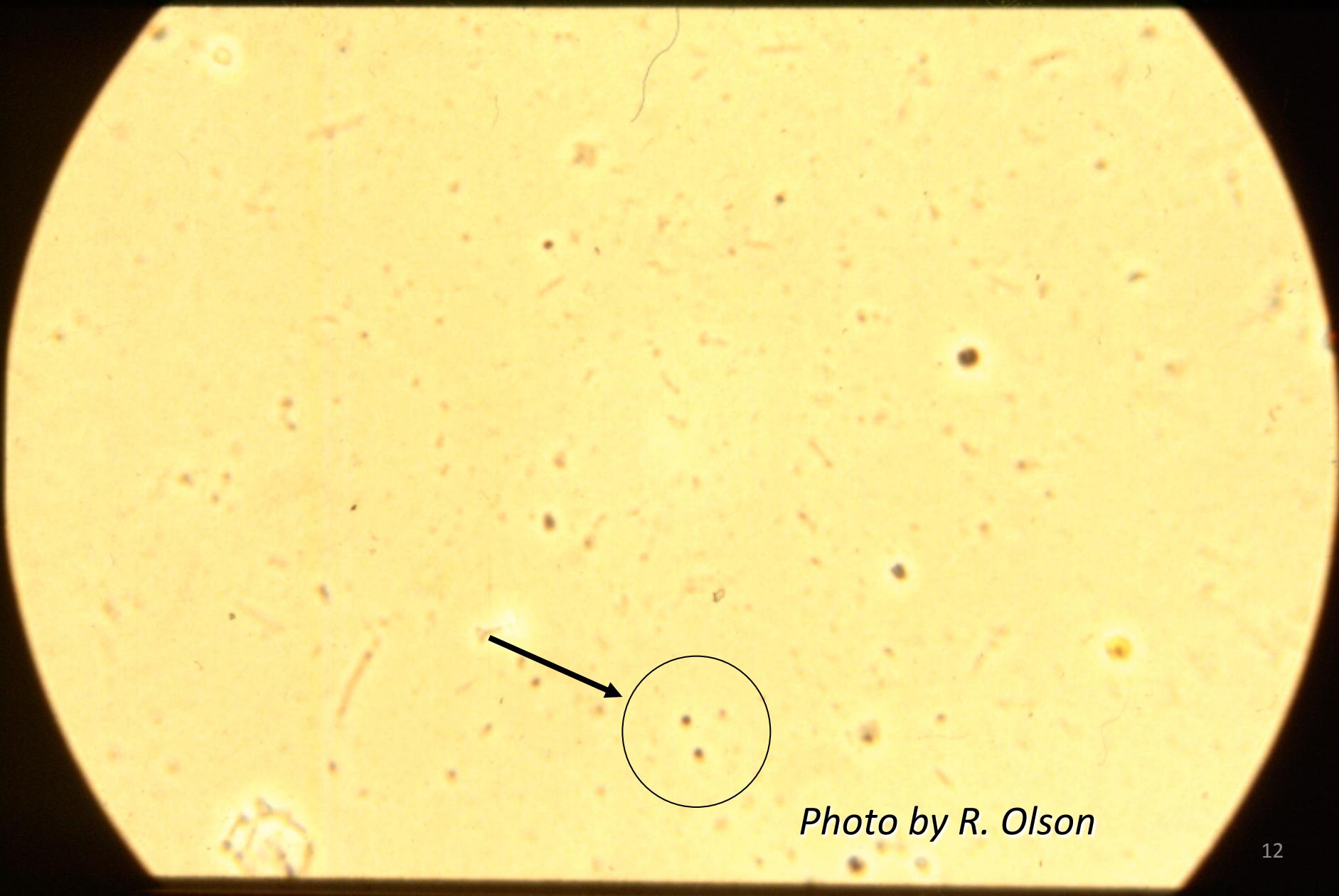




We set out to study  
*Synechococcus*,  
and noticed something else...

Based on their fluorescence excitation/emission spectrum - suspected chl b, typical of green algae.  
Called them "**Little Greens**"



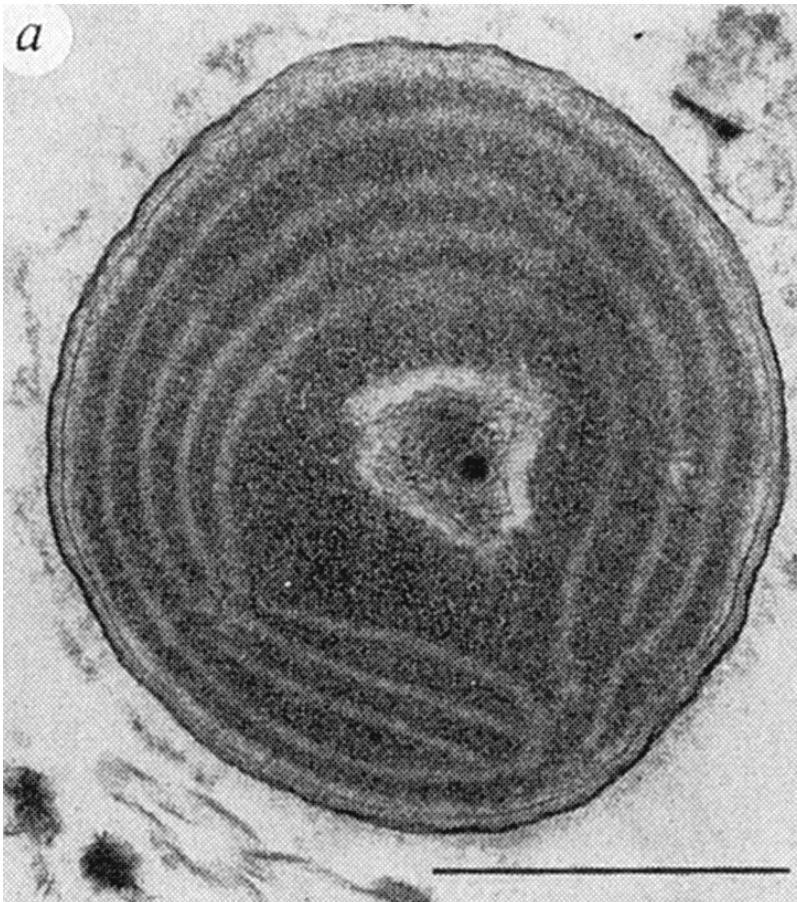


*Photo by R. Olson*

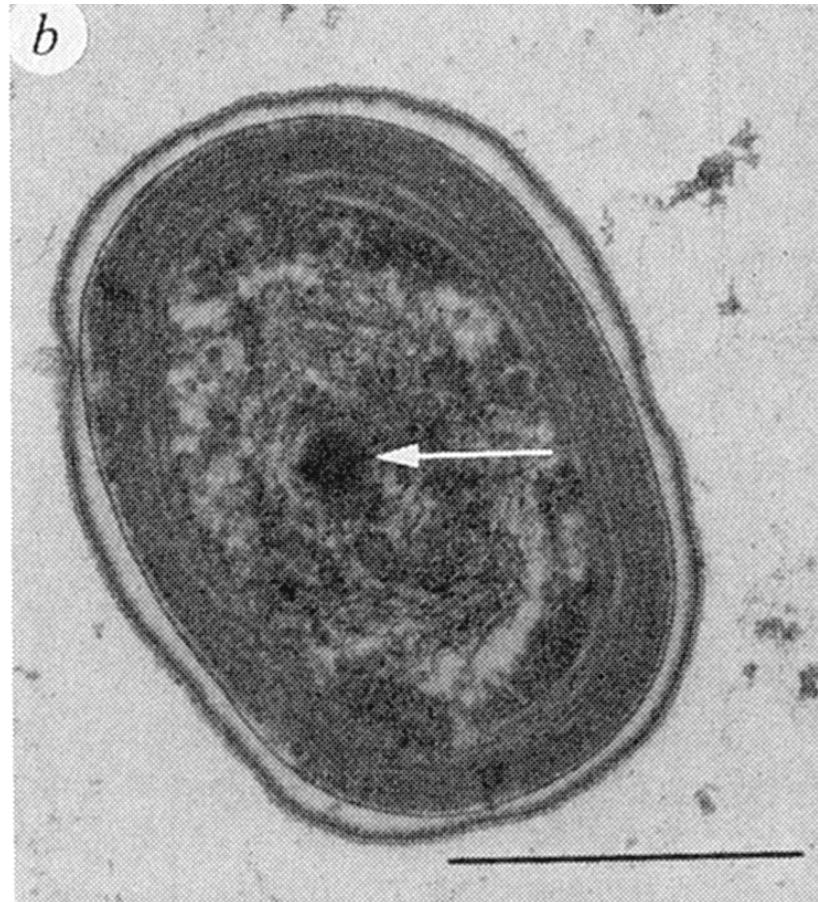
# Finally, an electron micrograph – It's a prokaryote!



*Synechococcus*

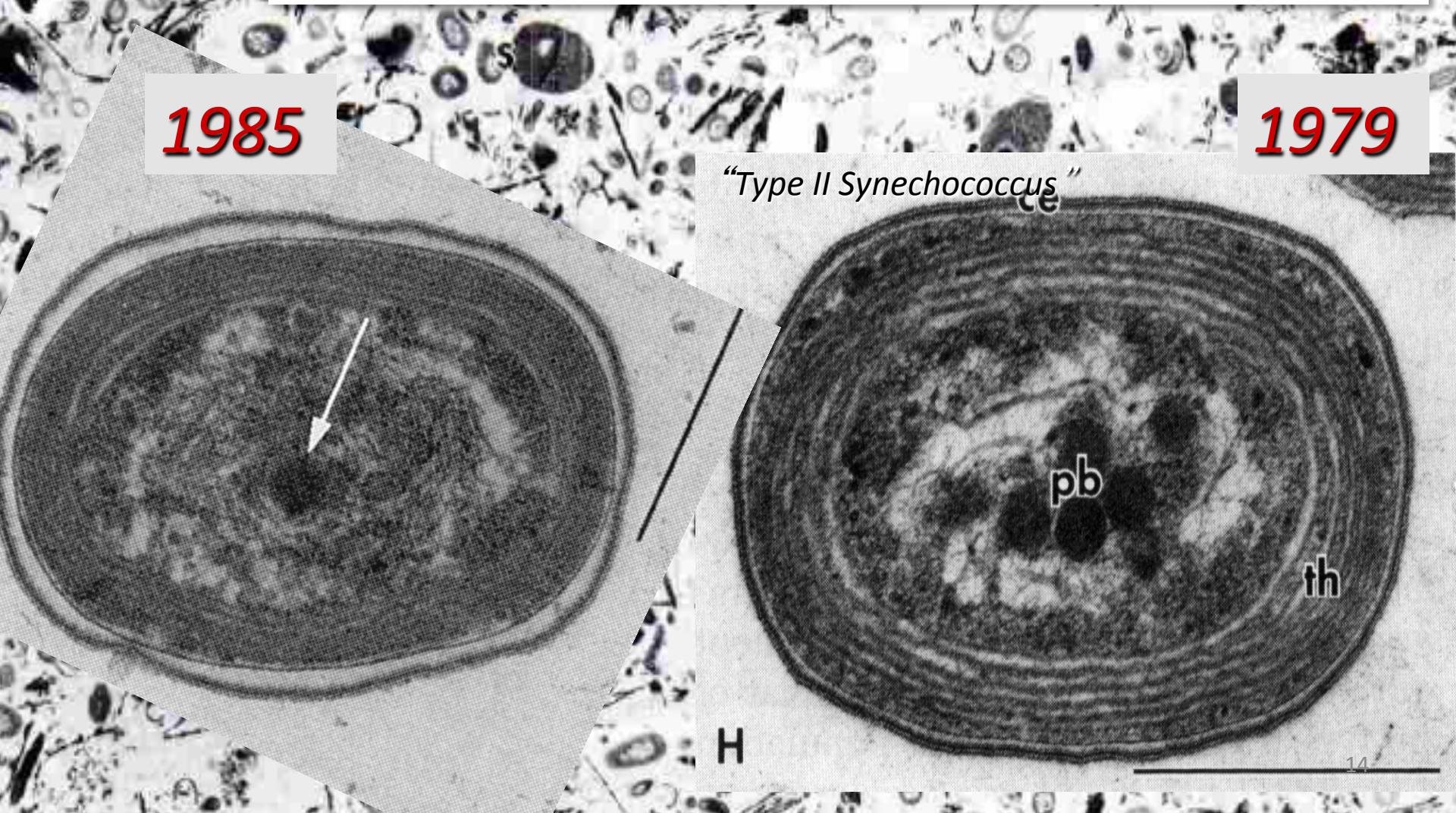


“Little Greens”



John Waterbury

# CLOSURE I: Johnson and Sieburth's “Type II Synechococcus” was the same as our cells



# What is their pigment composition?



Divinyl Chl

Ralph Goericke

HPLC pigment analysis of unidentified picoplankton

	0.8 $\mu\text{m}$ filtrate (>95% pure)	FCM sort
= divinyl chl a	chlorophyll a <sub>1</sub>	2.0 fg cell <sup>-1</sup> present
= divinyl chl b	chlorophyll b <sub>1</sub>	2.7 fg cell <sup>-1</sup> present
	zeaxanthin	0.6 fg cell <sup>-1</sup>
	$\alpha$ carotene	0.3 fg cell <sup>-1</sup>
	lutein	not detected
	chlorophyll a <sub>2</sub>	not detected
	$\beta$ carotene	not detected

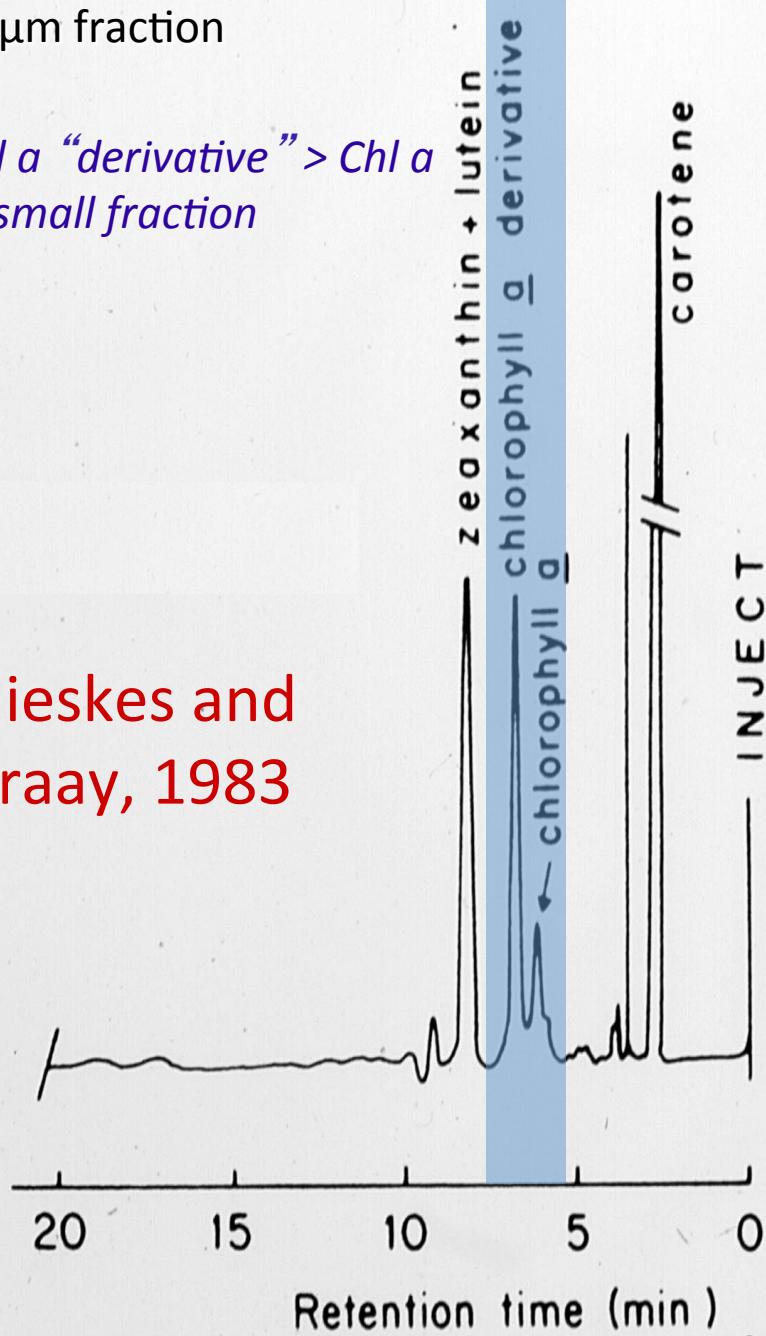
## CLOSURE II:

Gieskes and Kraay's  
“chlorophyll *a* derivative”  
was the divinyl chlorophyll *a*  
of the “Little Greens”

< 1 $\mu$ m fraction

*Chl a “derivative” > Chl a*  
*in small fraction*

Gieskes and  
Kraay, 1983



# But what do we call them?

# What else is prokaryotic and has Chlorophyll b?

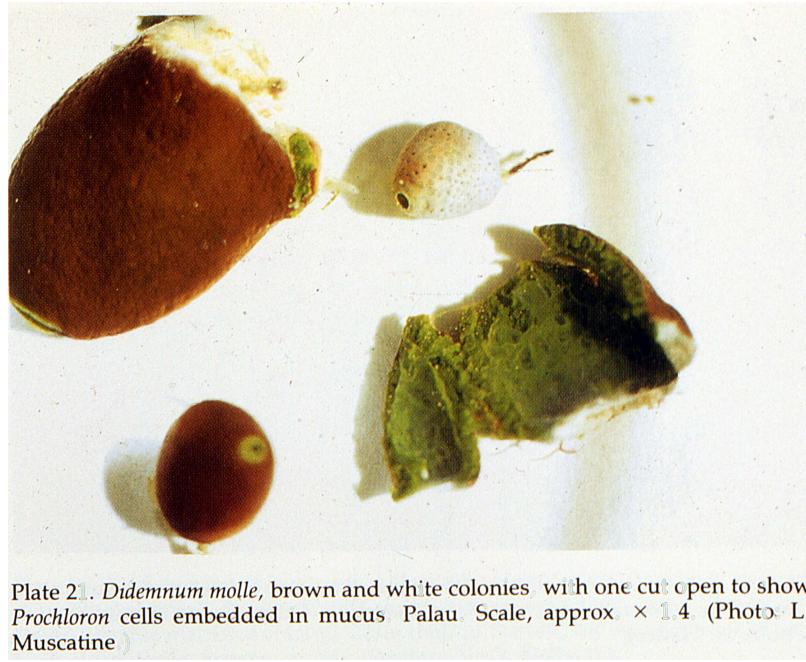
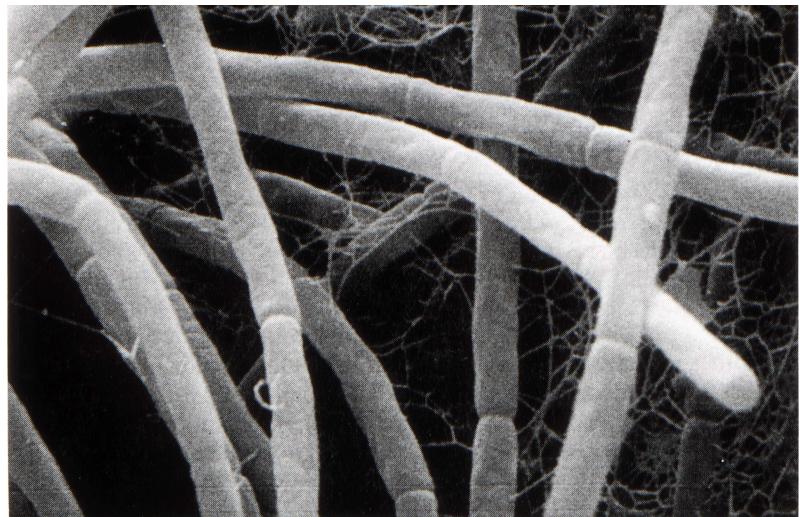


Plate 21. *Didemnum molle*, brown and white colonies with one cut open to show Prochloron cells embedded in mucus Palau Scale, approx  $\times 1.4$ . (Photo L Muscatine.)



*Prochloron*

*Prochlorothrix*

*So we called them “Prochlorophytes”*

# 1988: The Birth Announcement

340

LETTERS TO NATURE

NATURE VOL. 334 28 JULY 1988

## A novel free-living prochlorophyte abundant in the oceanic euphotic zone

Sallie W. Chisholm, Robert J. Olson\*, Erik R. Zettler\*,  
Ralf Goericke†, John B. Waterbury\*  
& Nicholas A. Welschmeyer†

48-425 Massachusetts Institute of Technology, Cambridge,  
Massachusetts 02139, USA

\* Woods Hole Oceanographic Institution, Woods Hole,  
Massachusetts 02543, USA

† Harvard University, Cambridge, Massachusetts 02138, USA

# 1992 Cultures



Brian Palenik

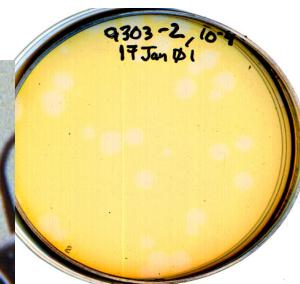


John Waterbury



Freddie Valois

Sometime later.....



Mak Saito

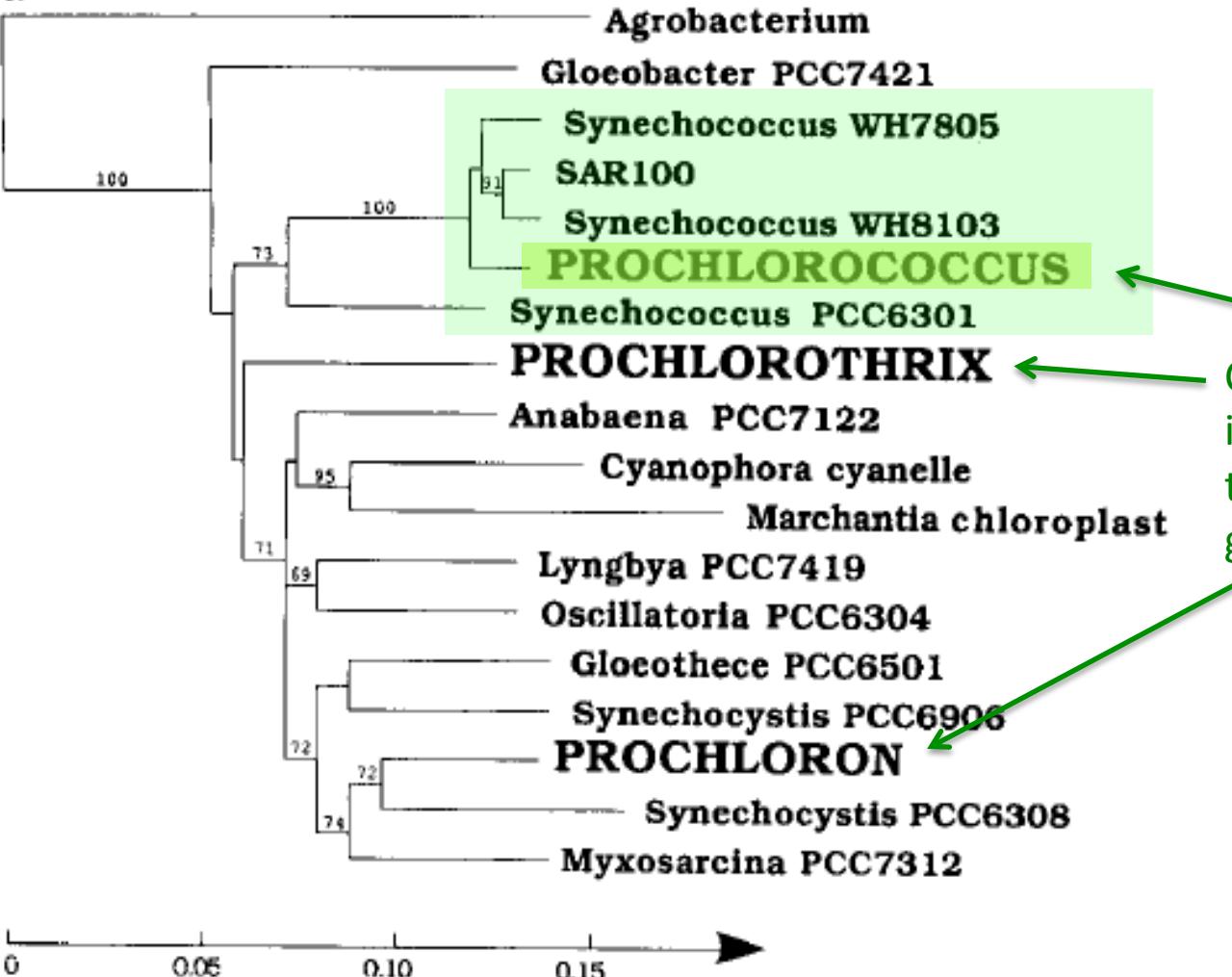
Lead to the name: *Prochlorococcus*

("coccoid prokaryote with chl b")

# Enter Molecular Phylogeny: We were wrong!

It *IS* closely related to *Synechococcus*, and not to *Prochloron* or *Prochlorothrix*

a



Chl b trait evolved independently in these three groups



Ena Urbach



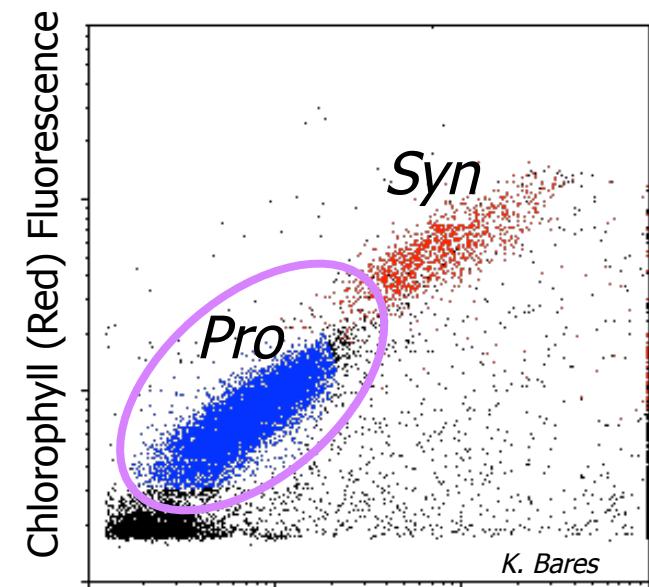
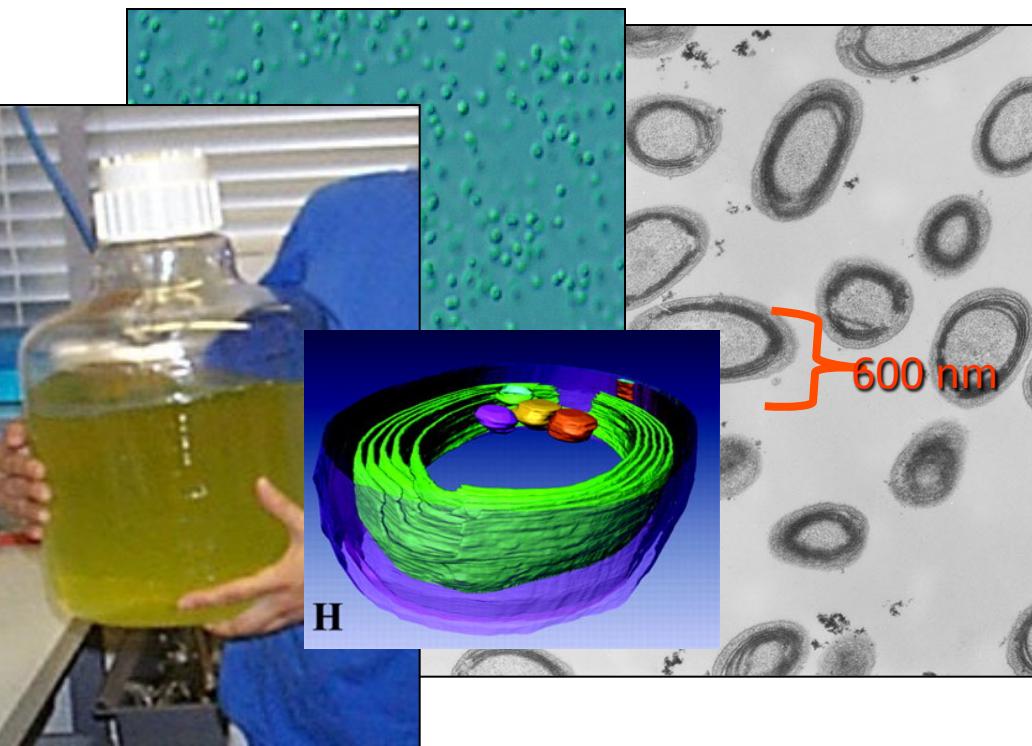
Mitch Sogin



Brian Palenik

# What exactly is *Prochlorococcus*?

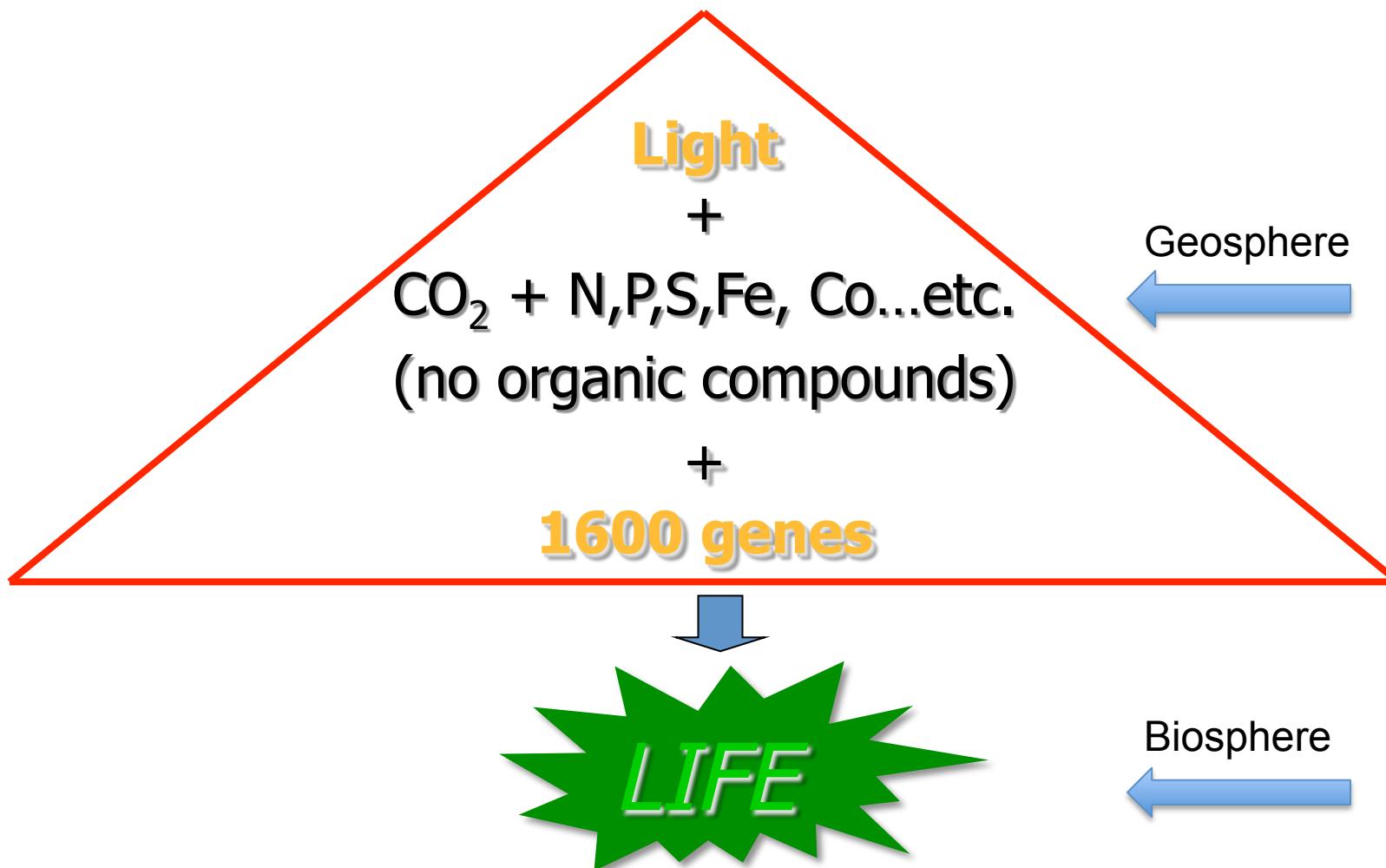
- Smallest cell in the oceans that fluoresces red
- Contains Divinyl Chl a and Chl b
- Oceanic cyanobacterium, 0.6 - 0.8  $\mu\text{m}$  diameter
- Smallest (size and genome), and most abundant photosynthetic cell on Earth



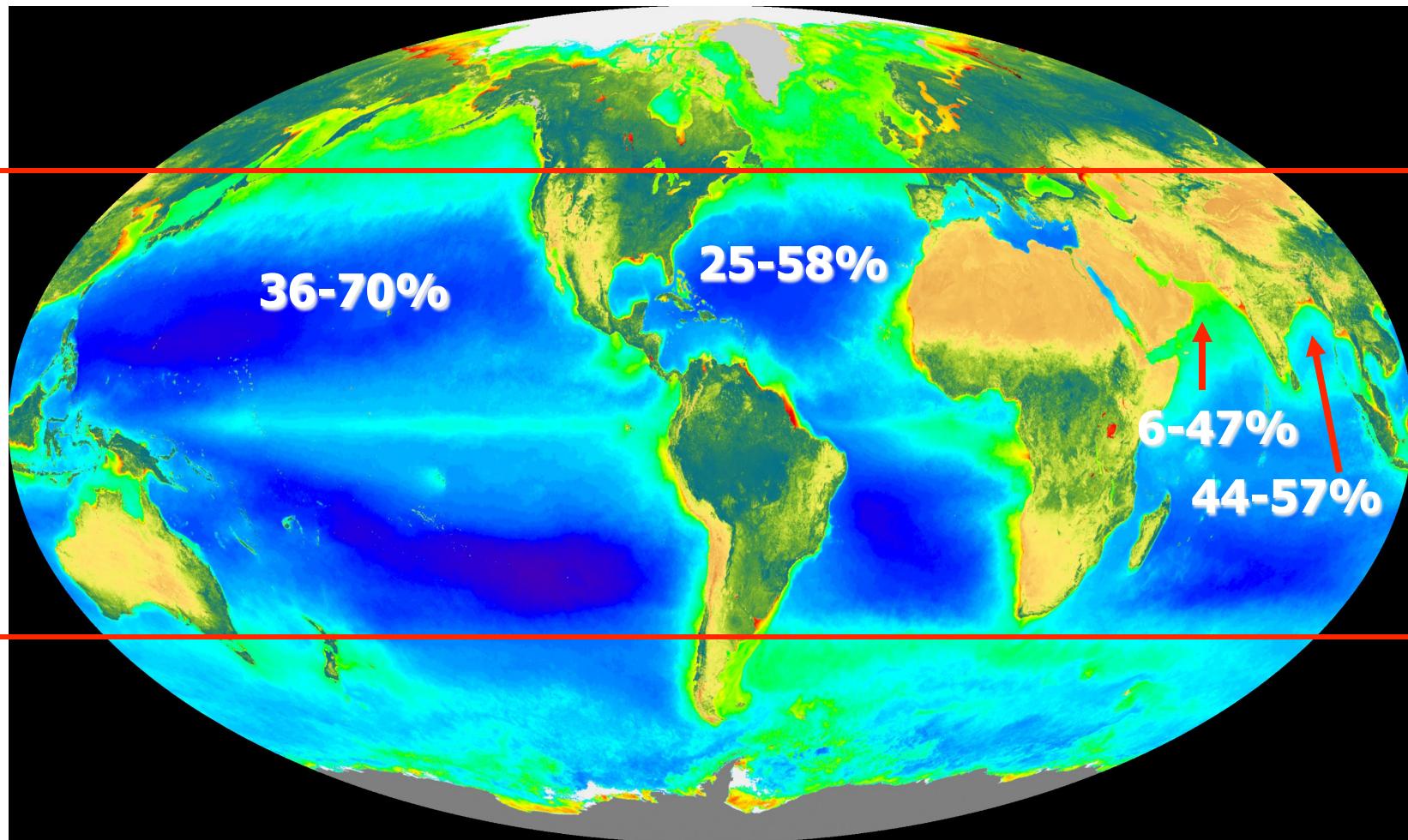
Forward Angle Light Scatter  
(Size)

# *Prochlorococcus* is the ‘minimal life’ form:

Smallest amount of information that can make life from scratch



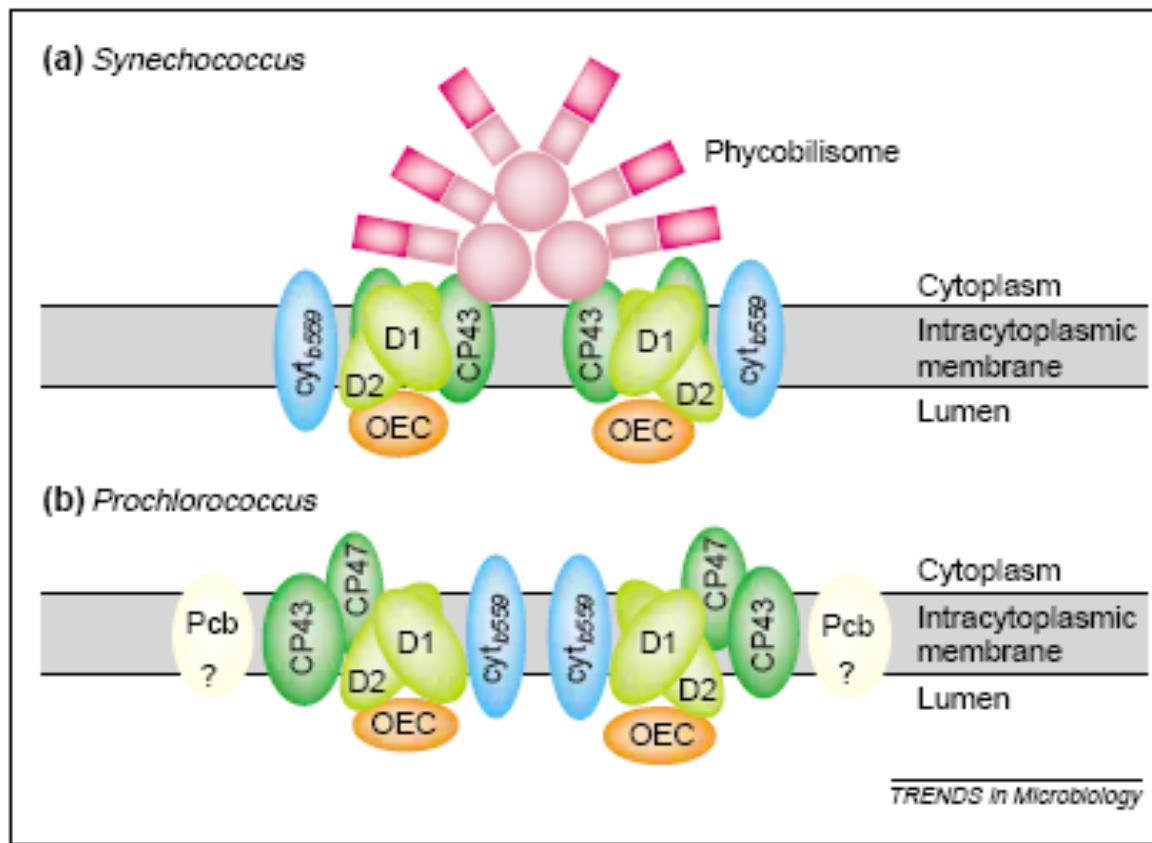
# Can be significant fraction of ocean chlorophyll



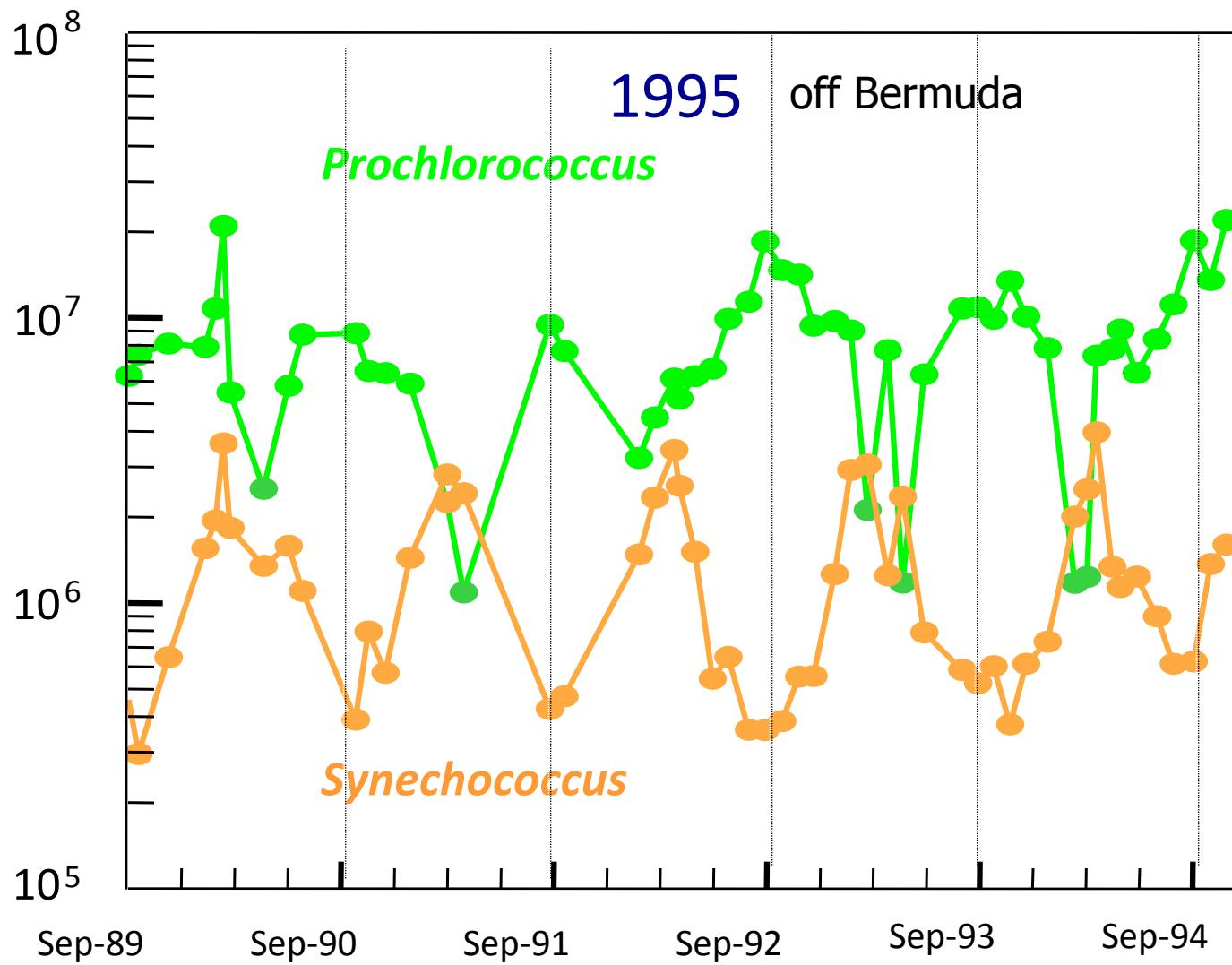
Record concentration:  
700,000 cells ml<sup>-1</sup>

Global  
photosynthesis ≈ 5 Gt C yr<sup>-1</sup> 23

# *Prochlorococcus* is basically a slightly smaller *Synechococcus*, with a different light harvesting system

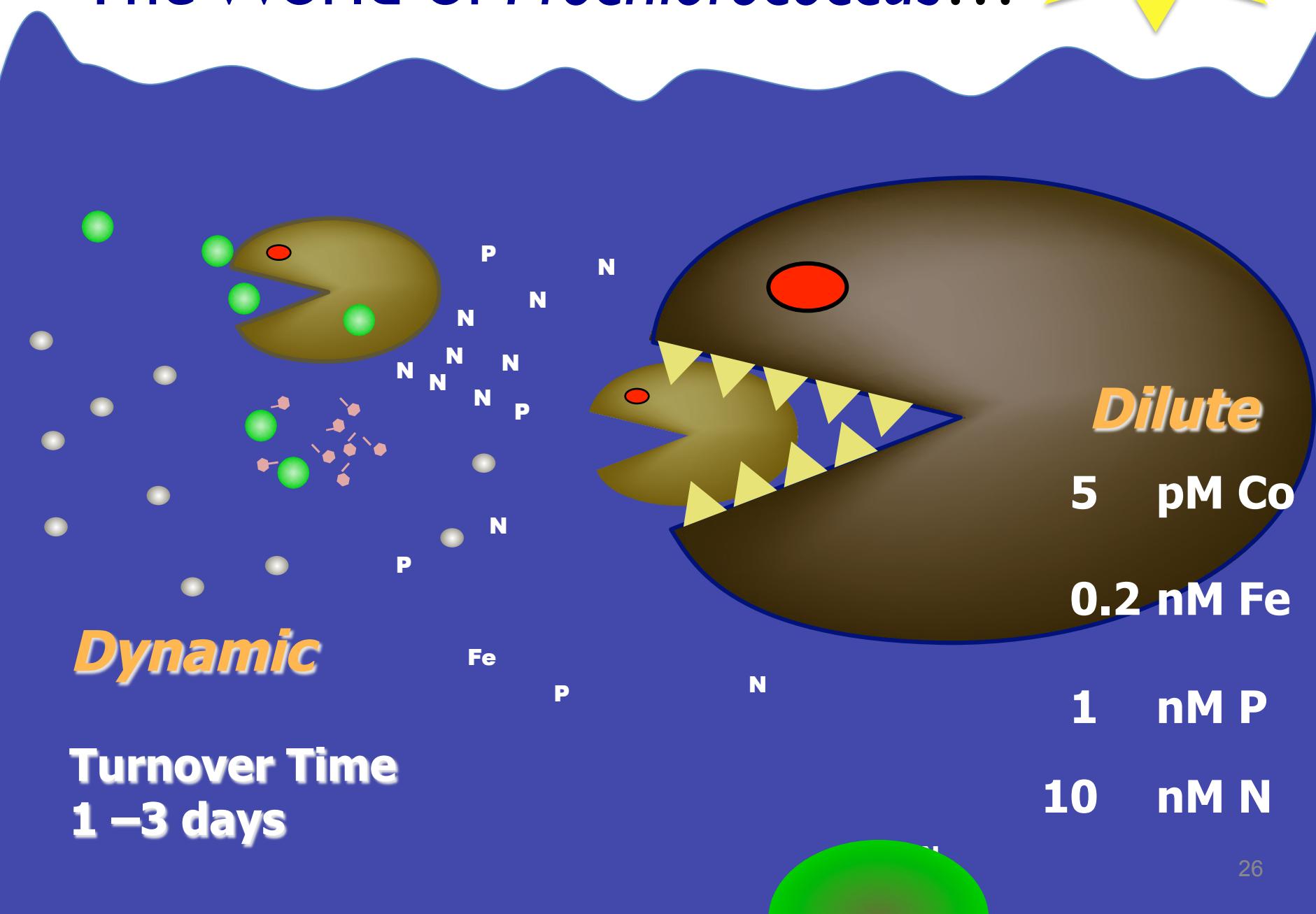


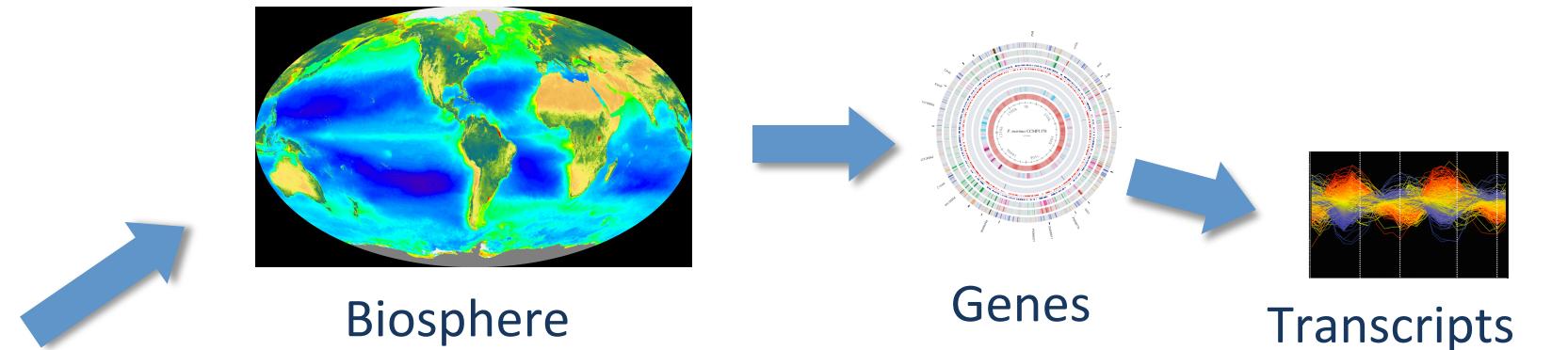
# They share (and partition) the “small size bin”



DuRand et al 2001

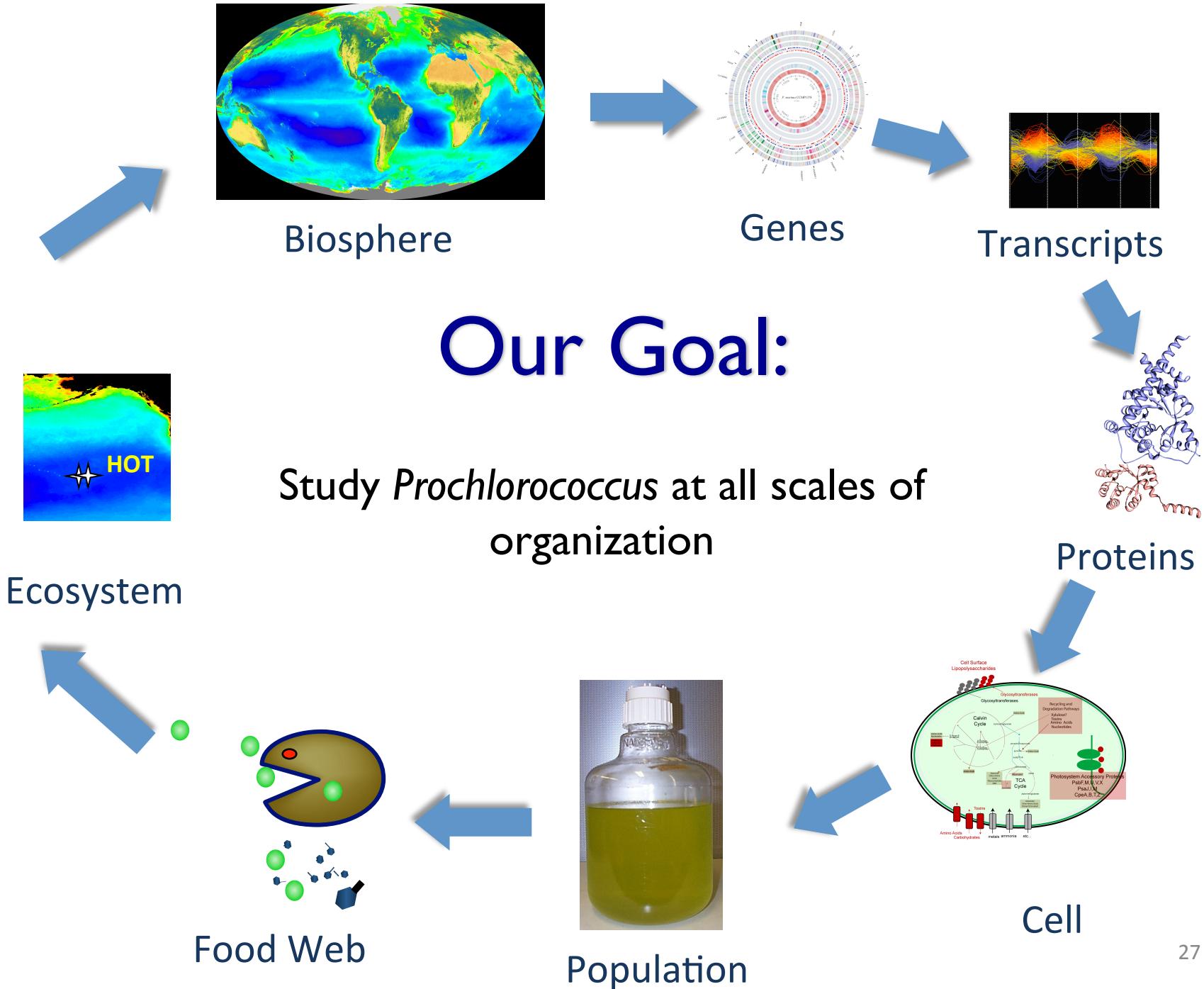
# The World of Prochlorococcus...





# Our Goal:

Study *Prochlorococcus* at all scales of organization



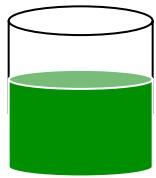
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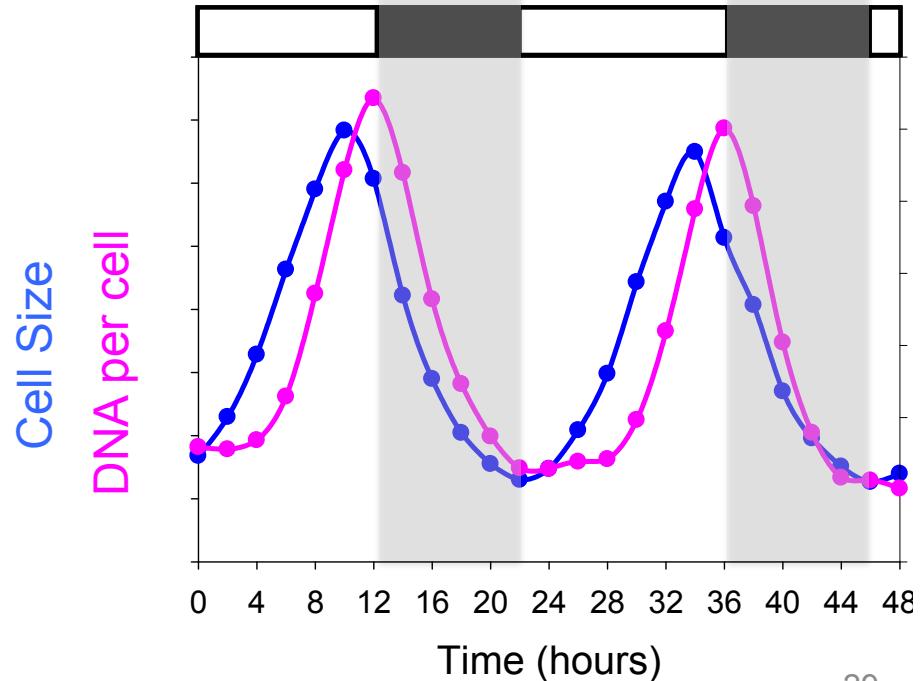
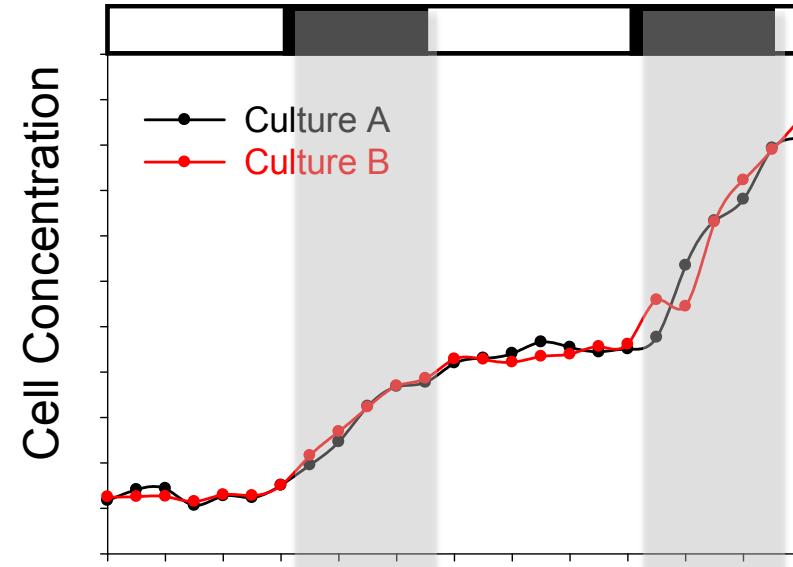
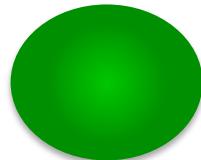
# Tight Cell Cycle Synchrony

Culture



==

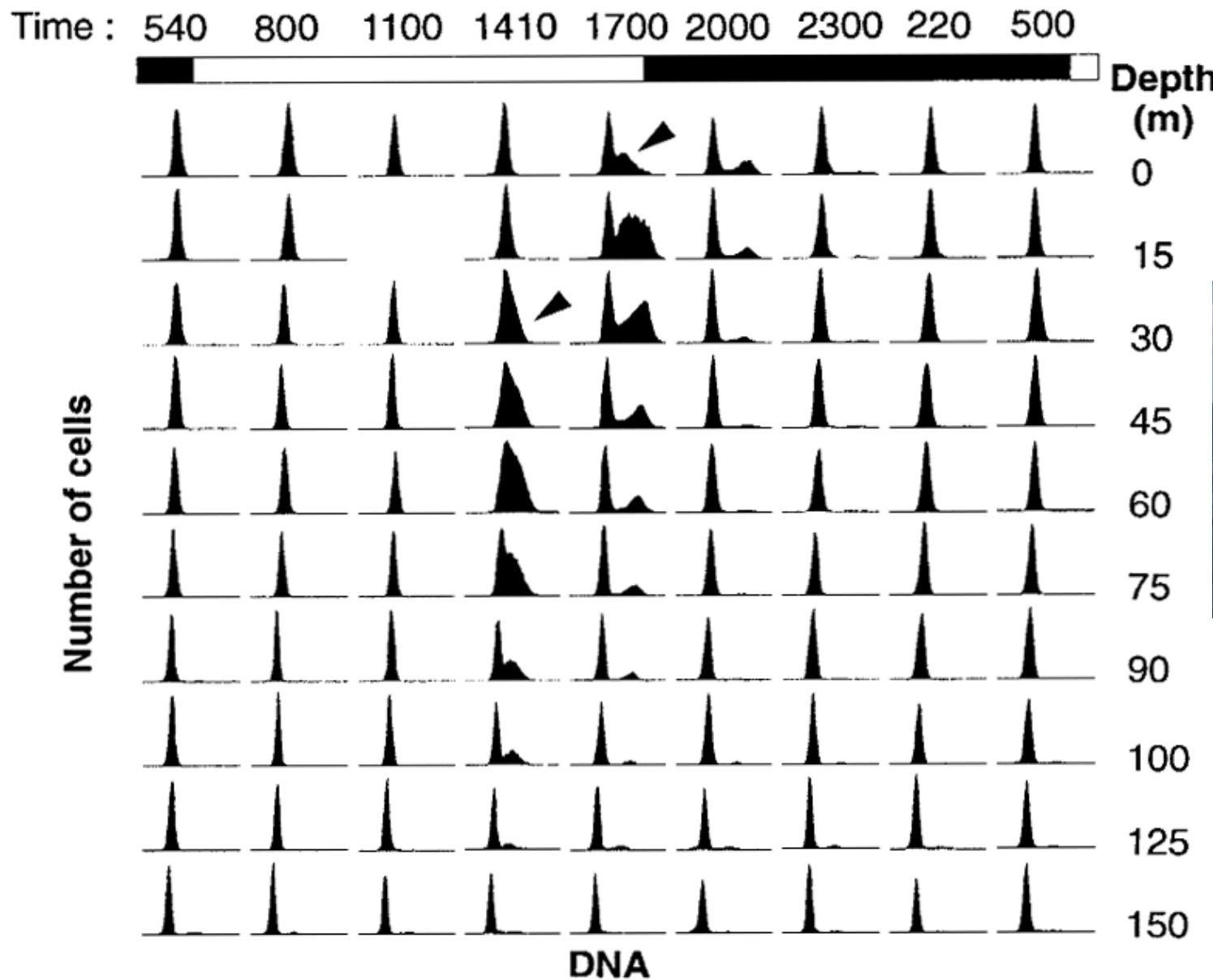
Cell



# Can see it growing in the wild

1 April 1992

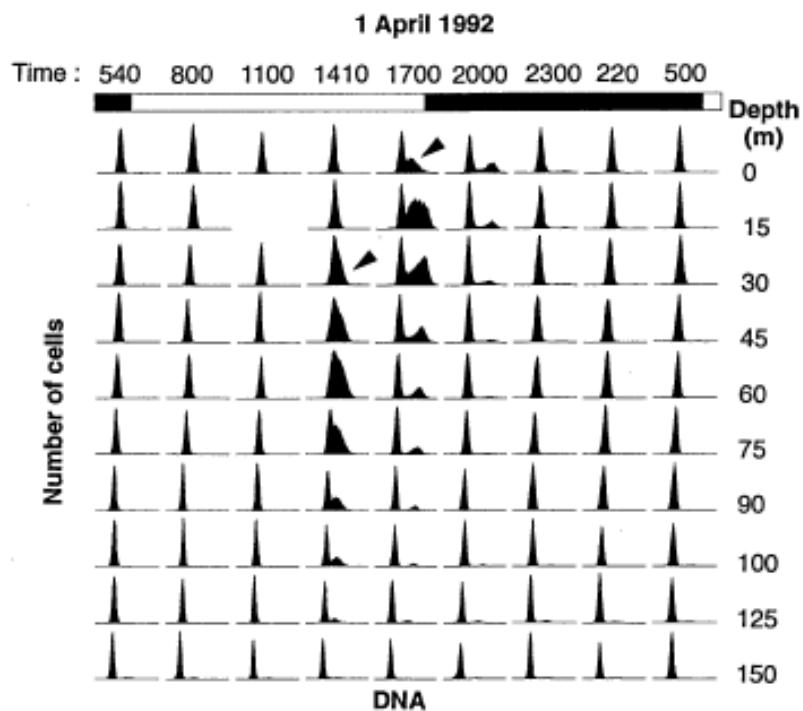
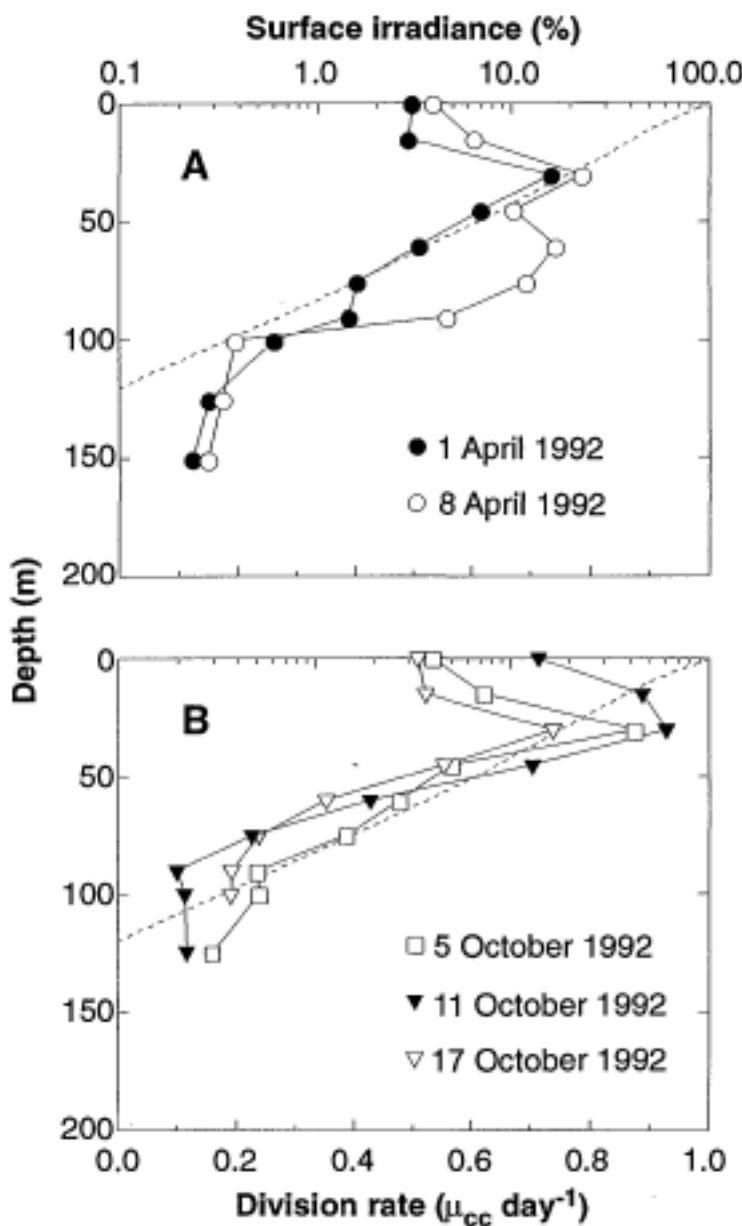
Equatorial  
Pacific



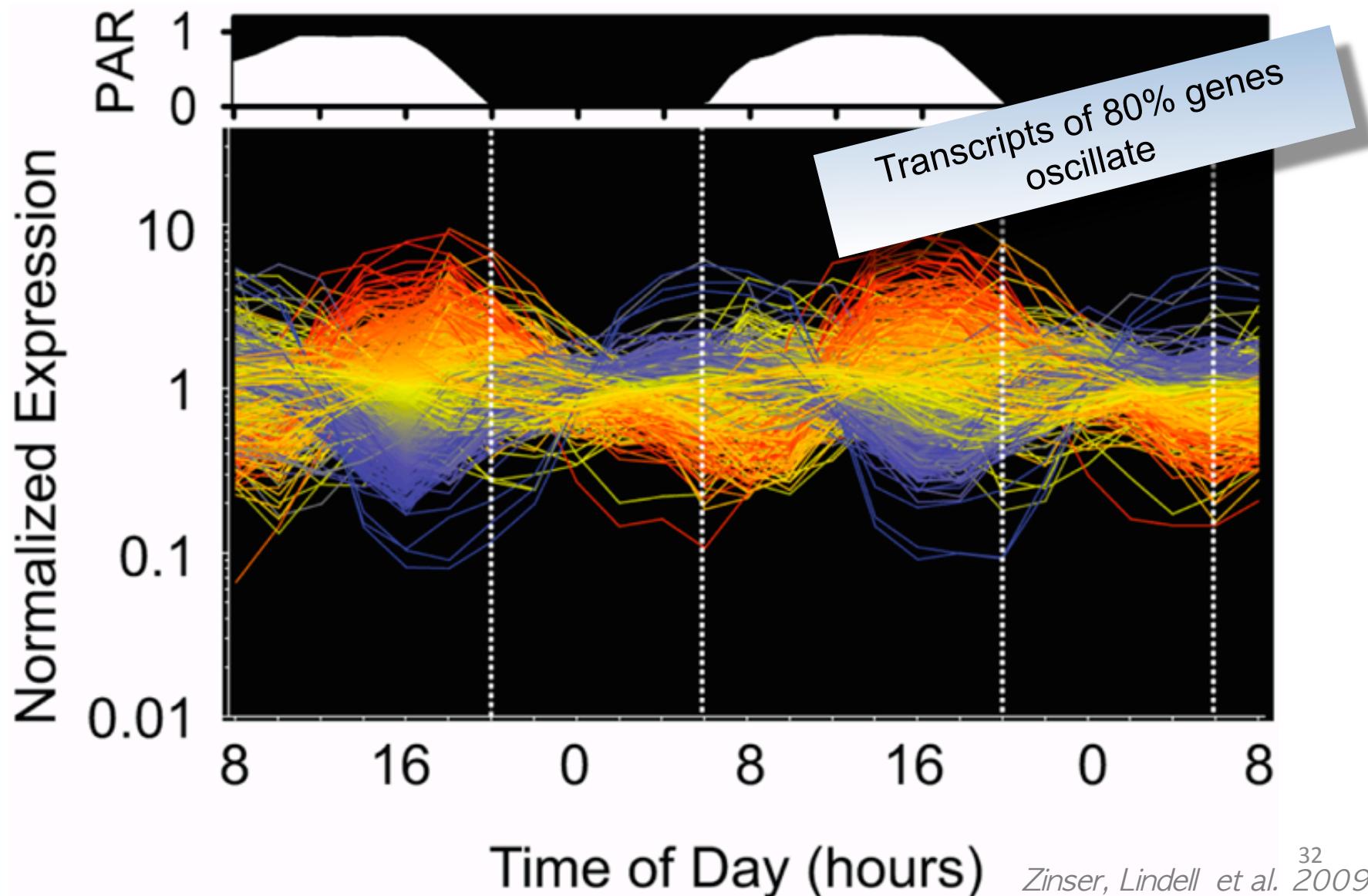
Daniel Vaulot

# *Prochlorococcus* growth rate as a function of depth

(measured by cell cycle progression)

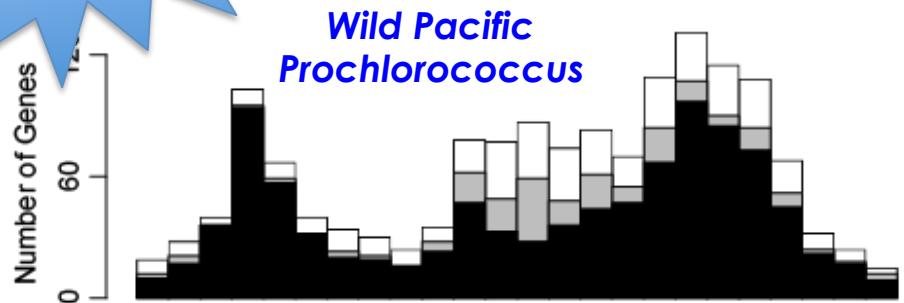


# Gene expression highly choreographed



News  
flash!

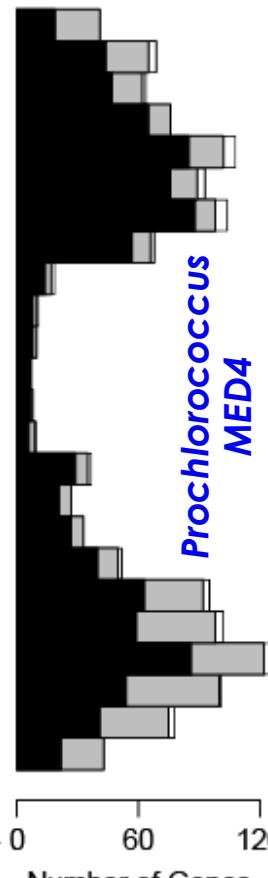
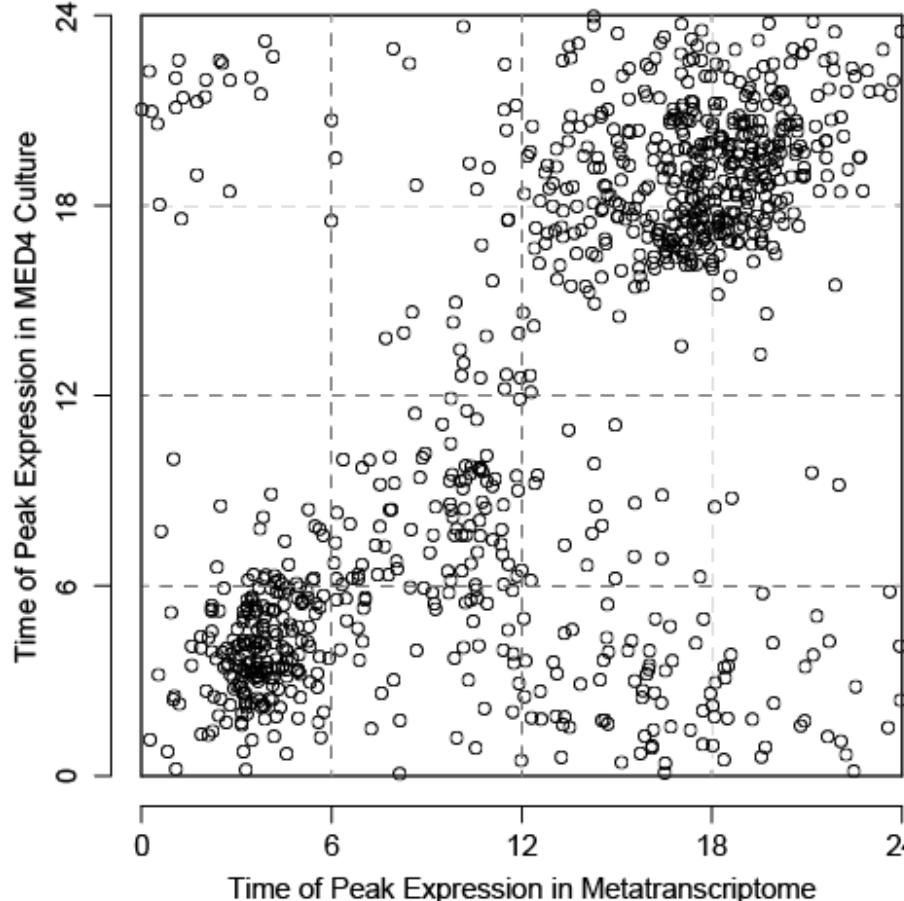
# ... in the wild too!



Ottesen et al & DeLong,  
*SCIENCE*, in press



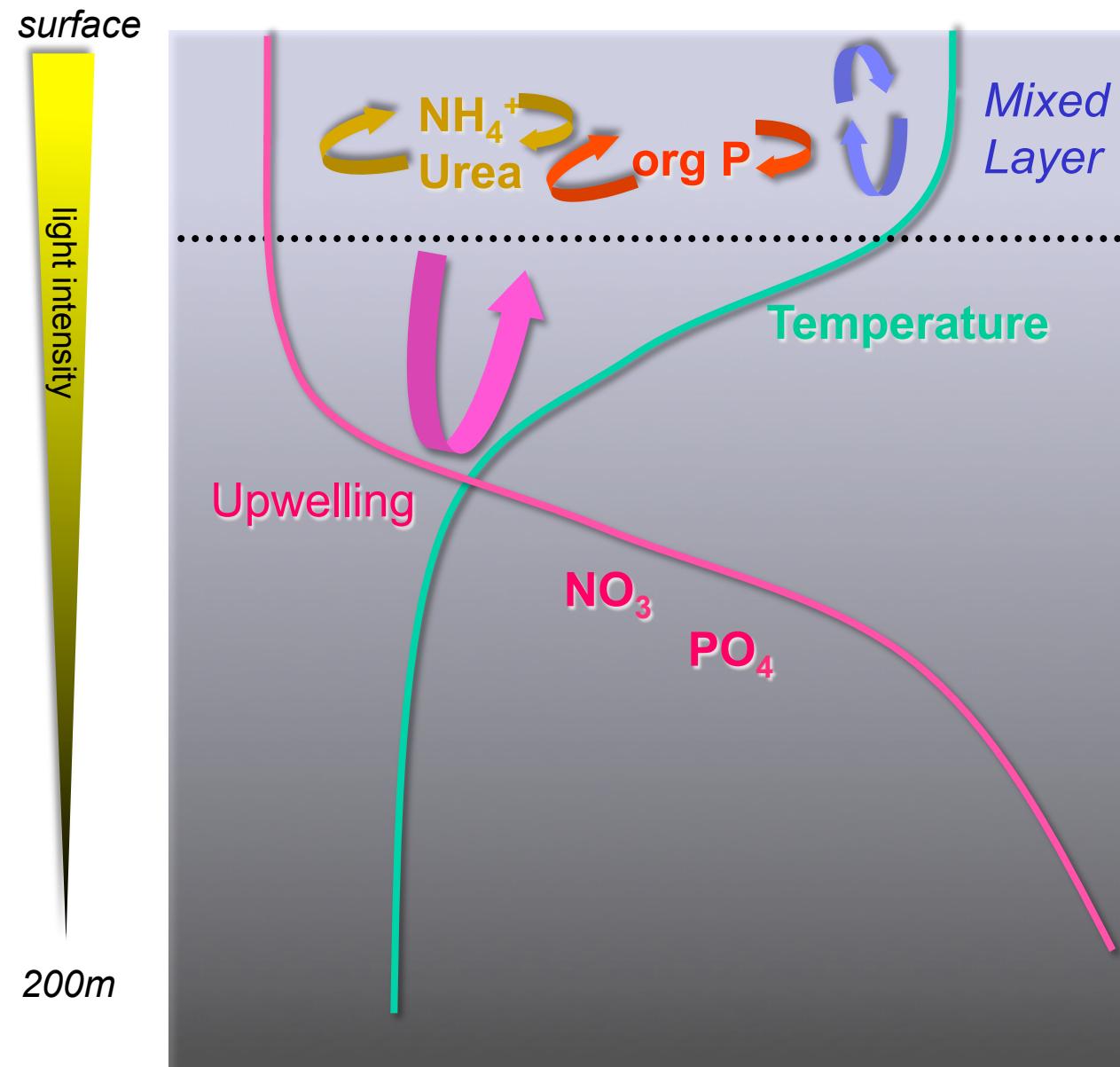
Multispecies diel transcriptional oscillations in open ocean heterotrophic bacterial assemblages



Zinser et al & Chisholm 2009

Choreography of the transcriptome, photophysiology and cell cycle of a minimal photoautotroph

# Reminder: Structure of the ocean habitat



...physical and  
chemical  
gradients =

**Niche  
dimensions!**

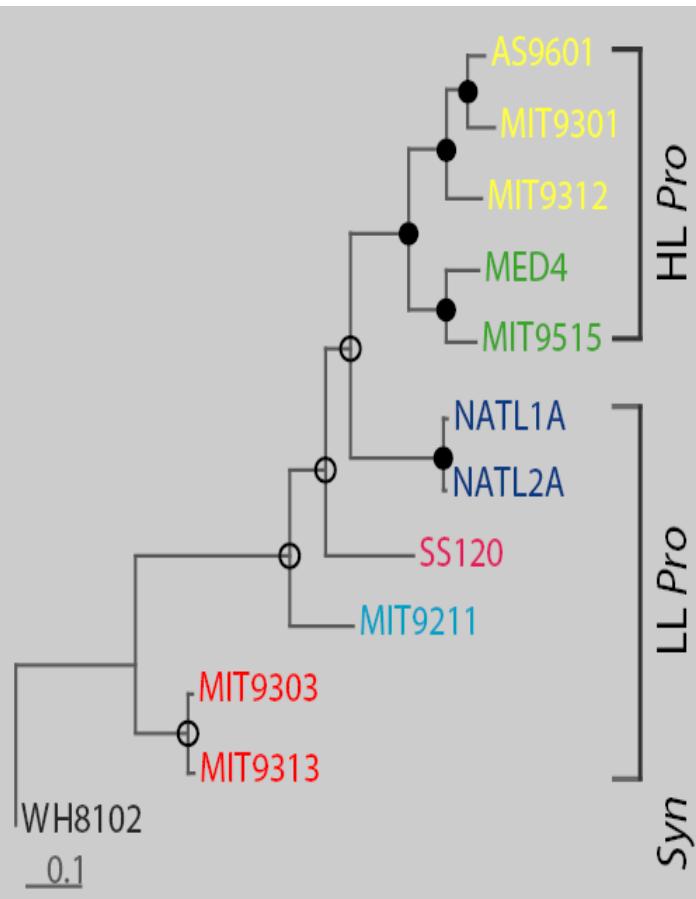
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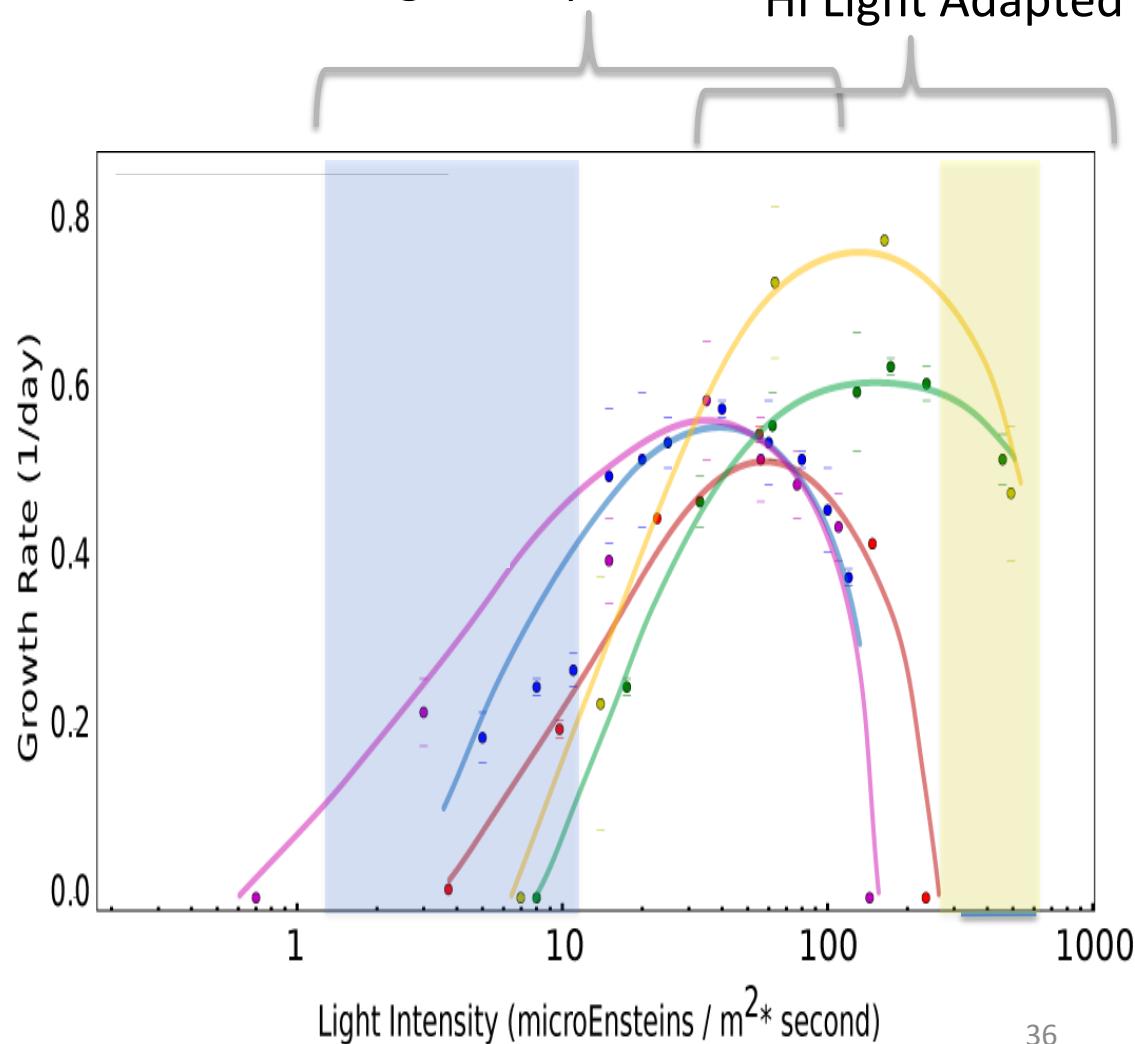
# Light adaptation defines ecotypes...

Whole Genome Phylogeny  
(matches rRNA ITS phylogeny)

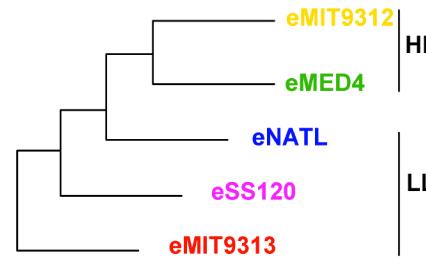
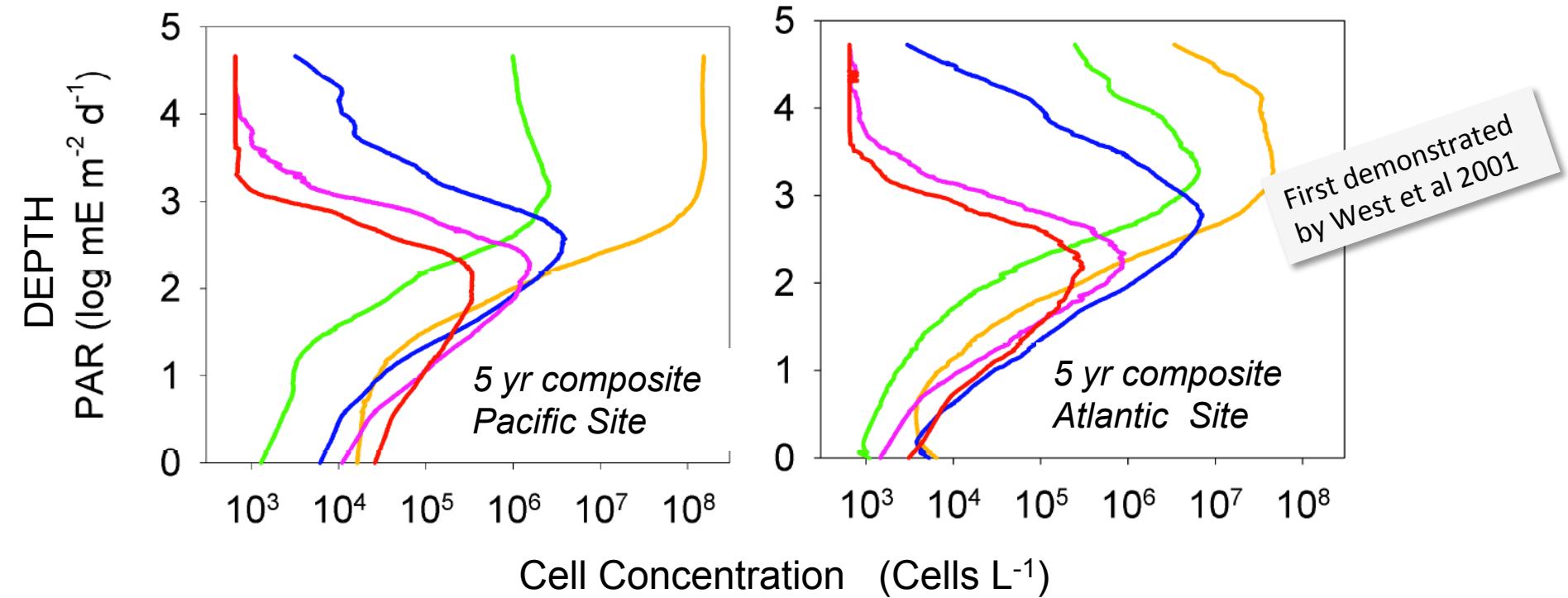


Low Light Adapted

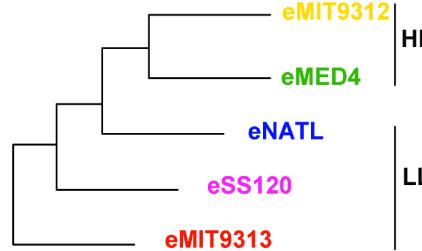
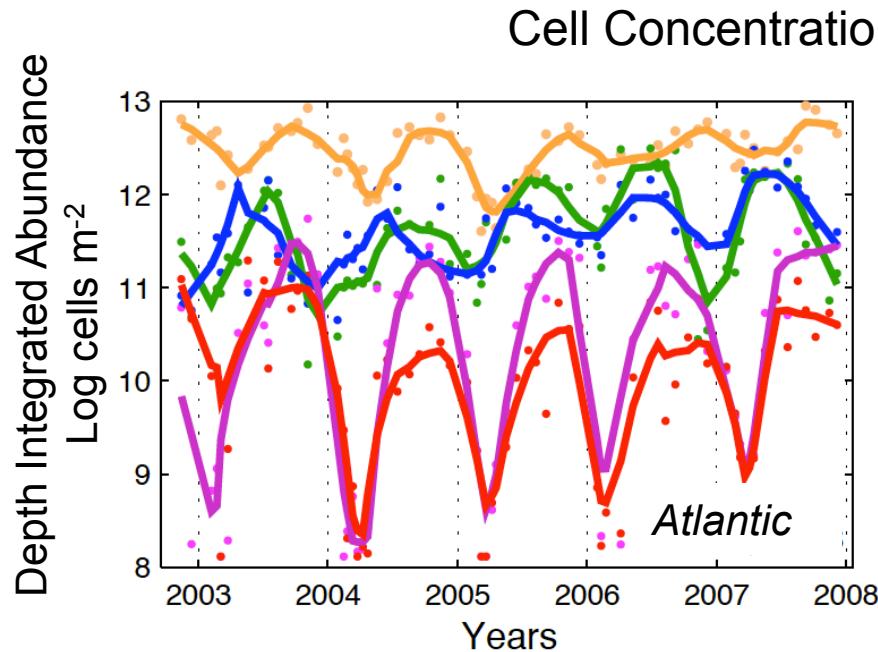
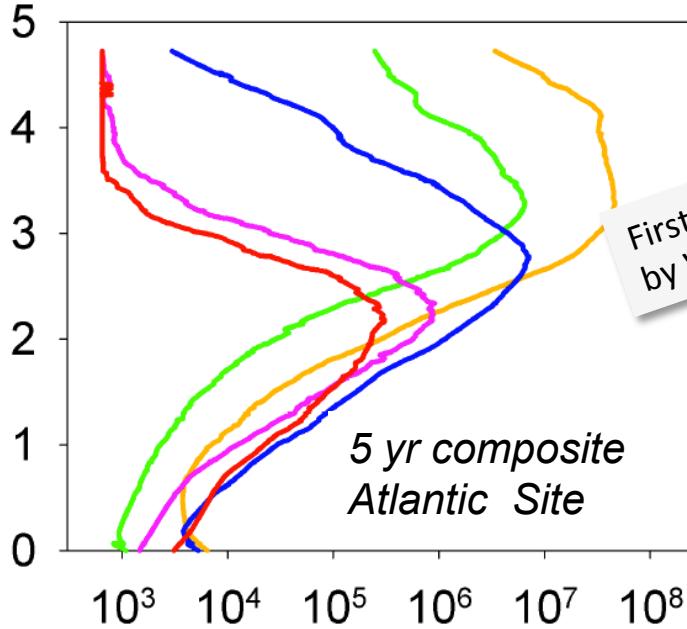
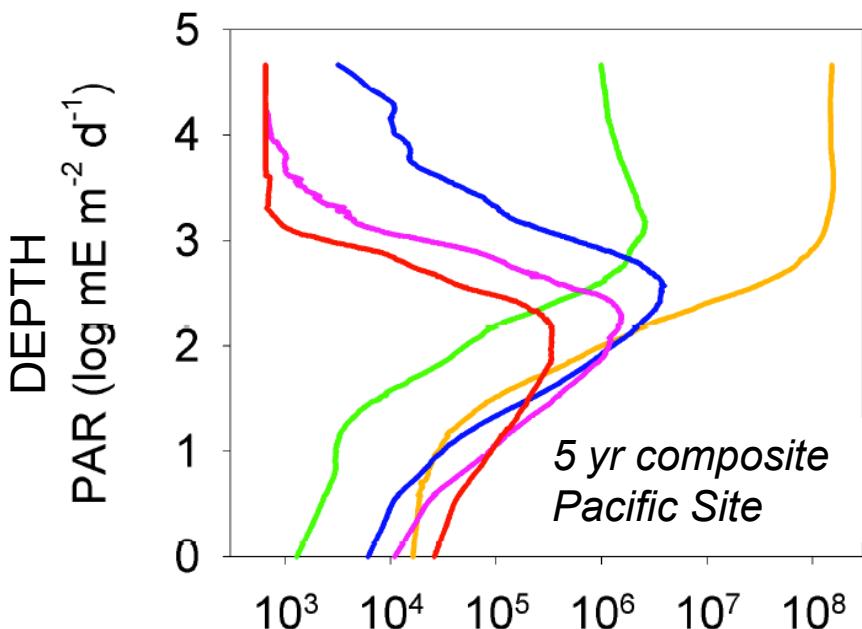
Hi Light Adapted



# ...leading to niche differentiation

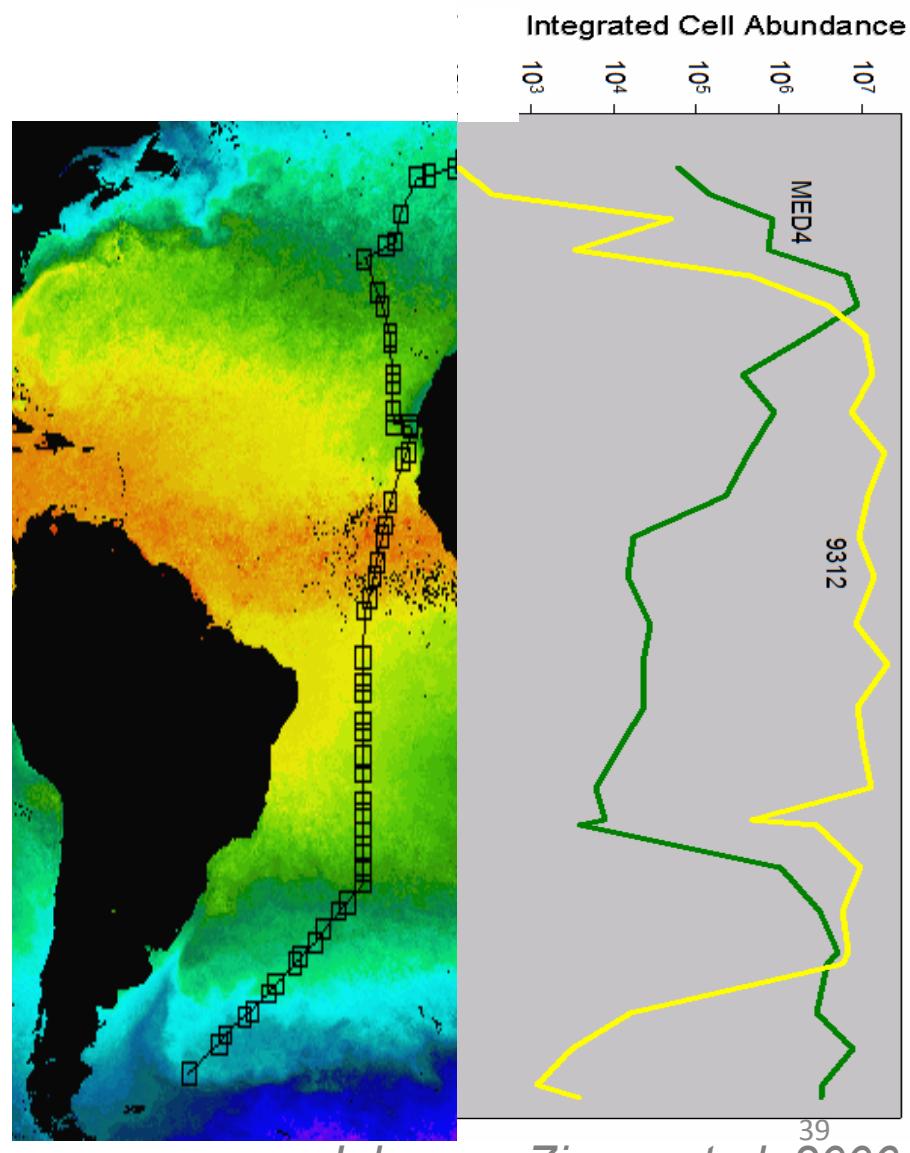
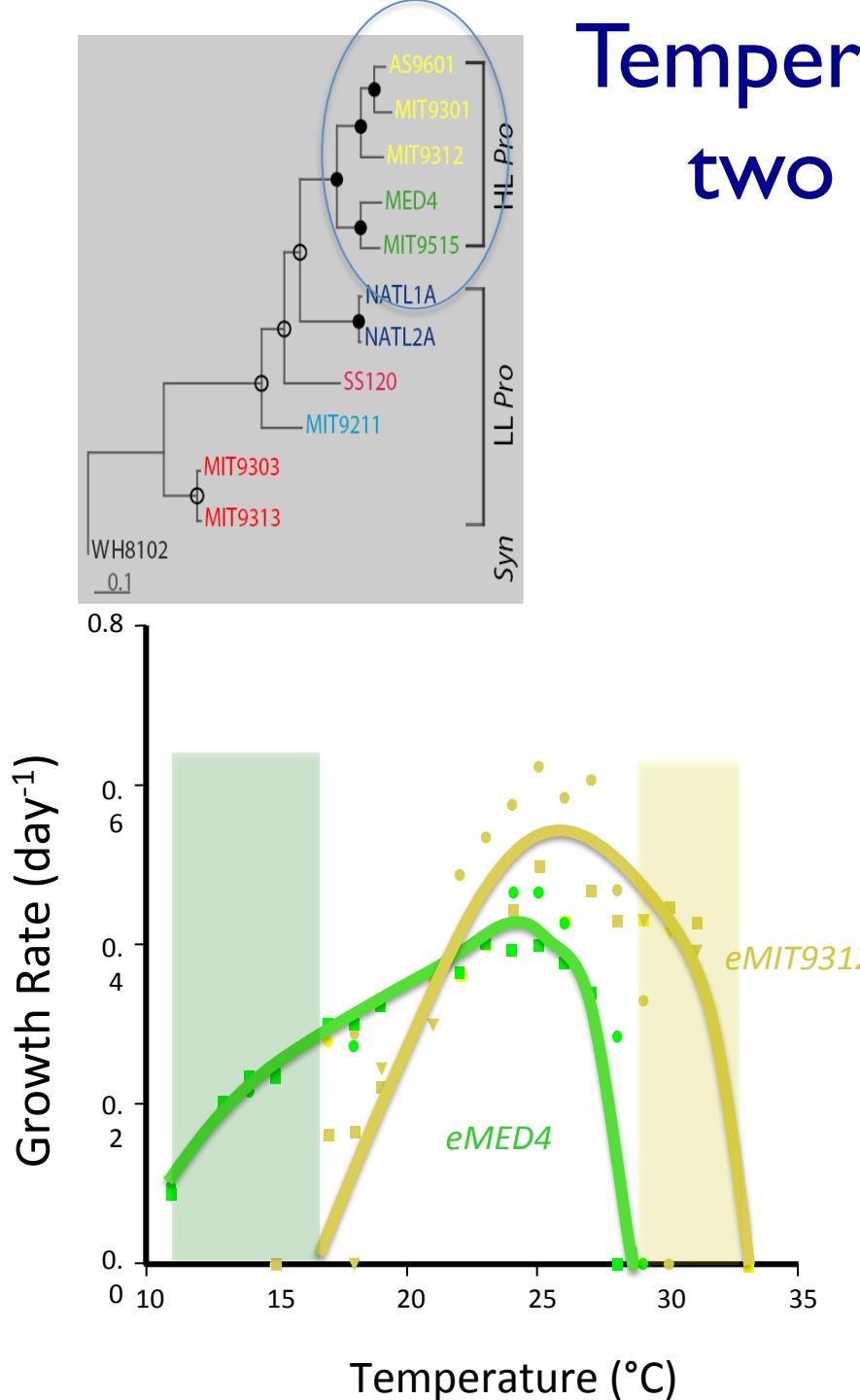


# ...leading to niche differentiation



Malmstrom et al 2010

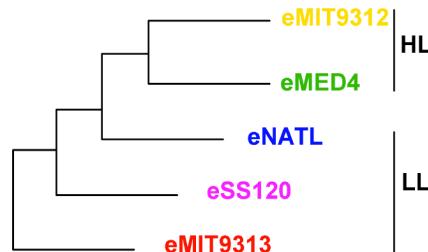
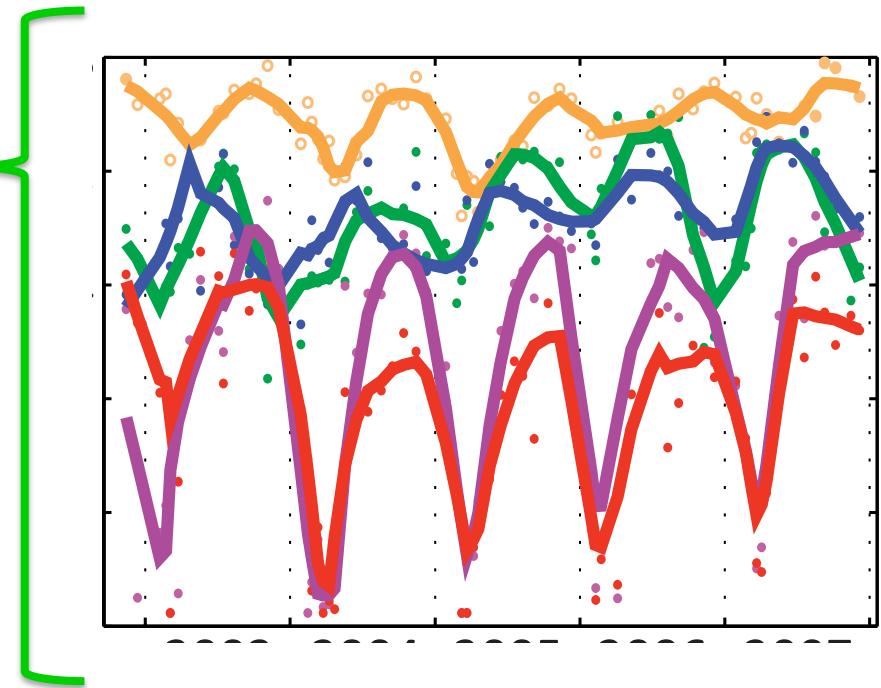
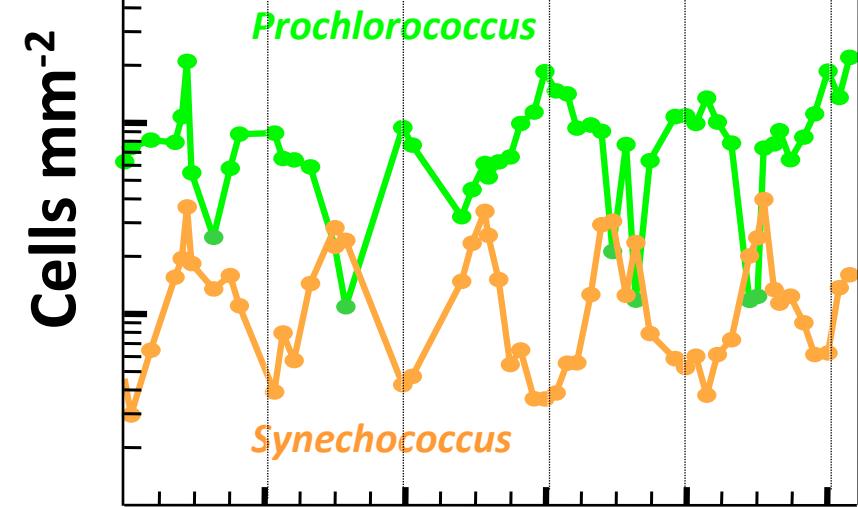
# Temperature differentiates the two High Light ecotypes



Johnson, Zinser et al, 2006

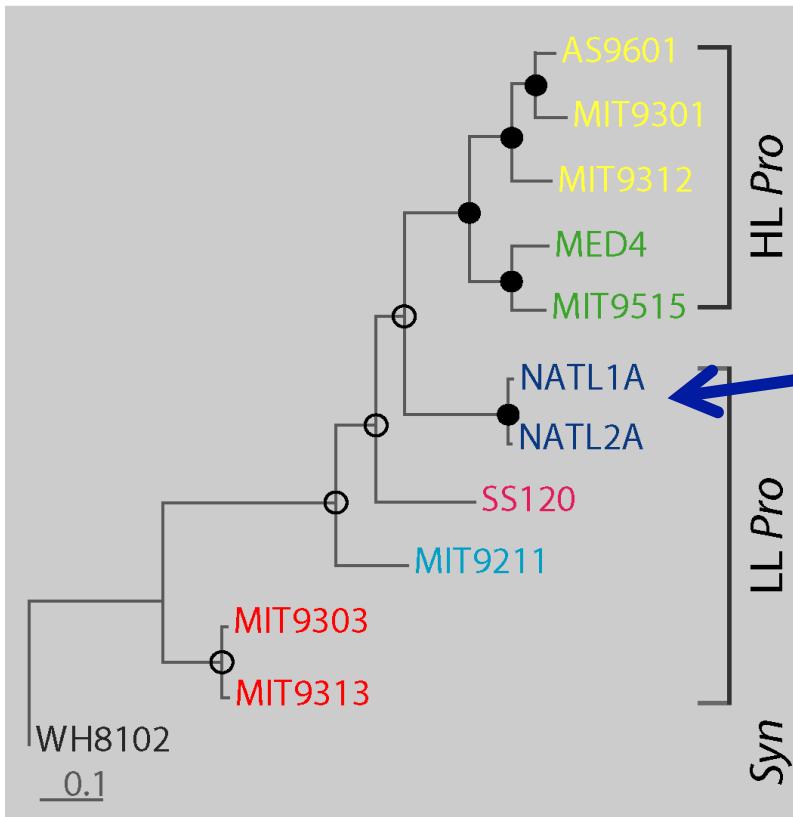
So there are meaningful layers, within layers, of diversity  
(leads to stability)

*Prochlorococcus* ecotypes

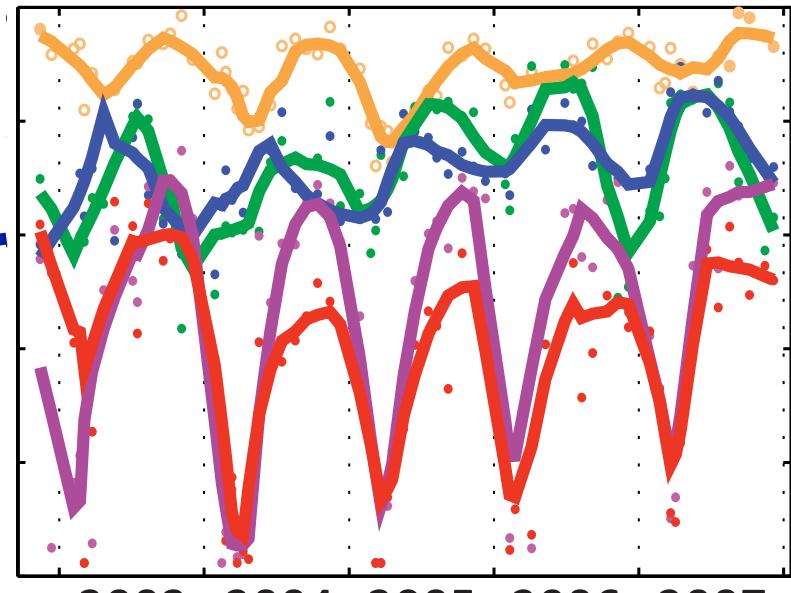


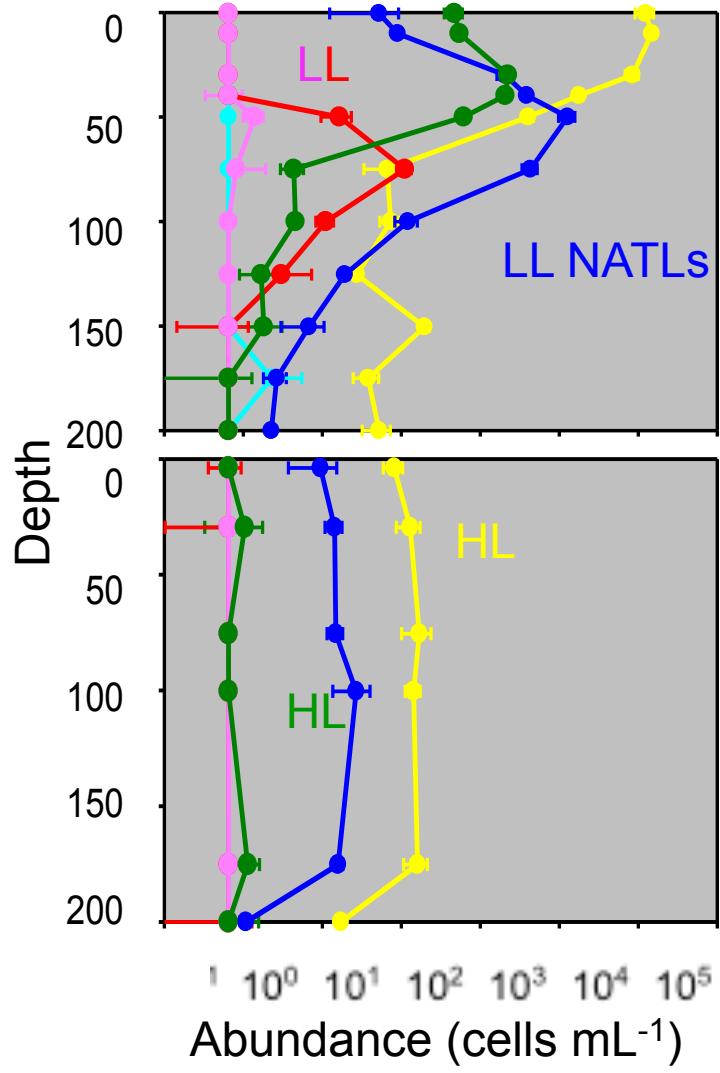
# “The NATLs” are very interesting

Not HL-Adapted, but intermediate taxonomic position



*Prochlorococcus* ecotypes

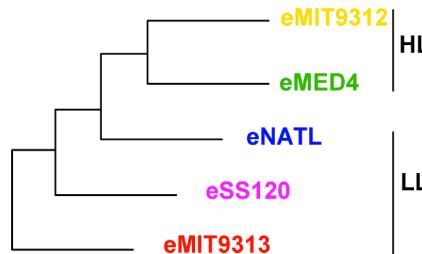




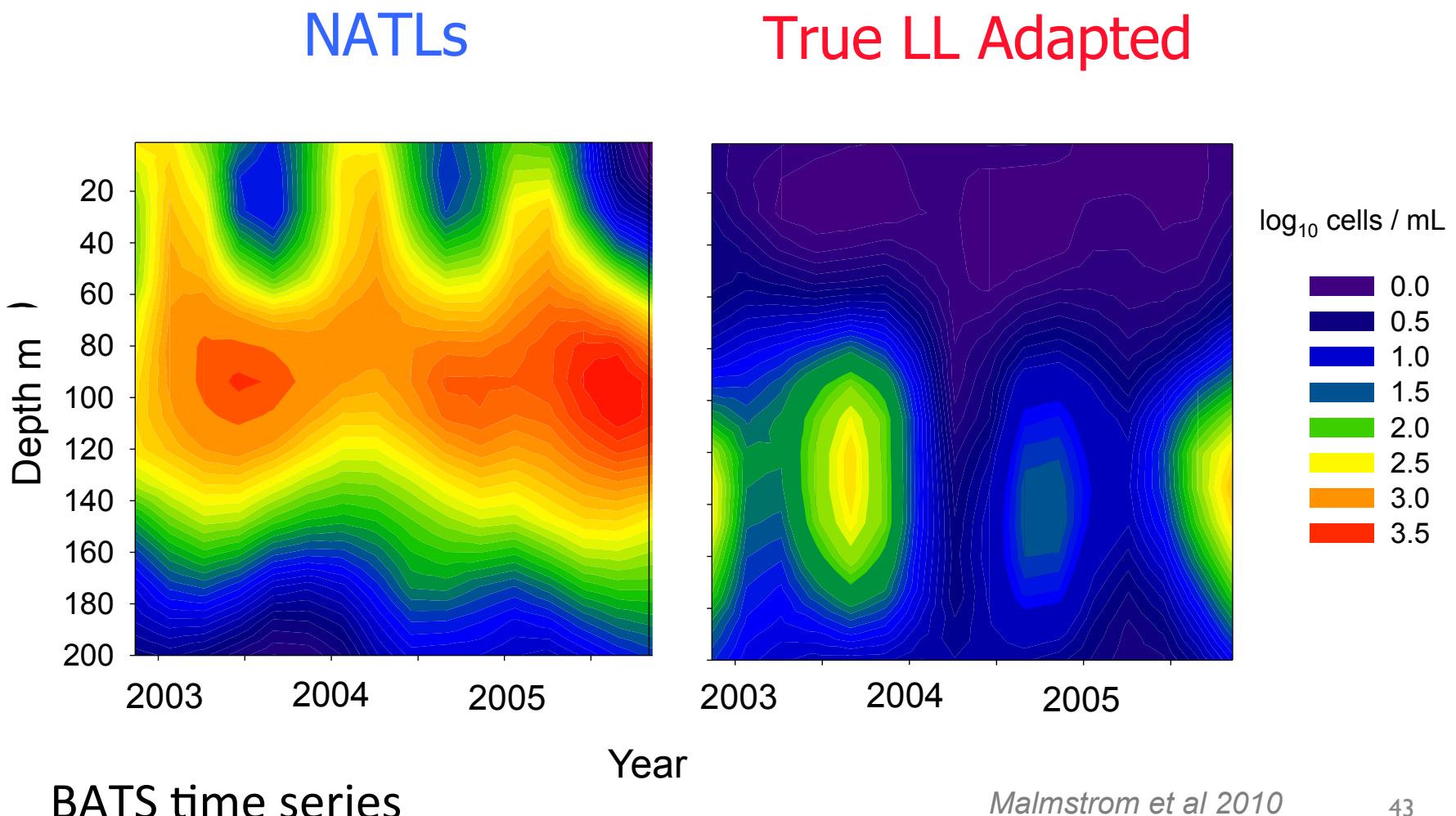
NATLs appear to:

handle high light better than other LL ecotypes...

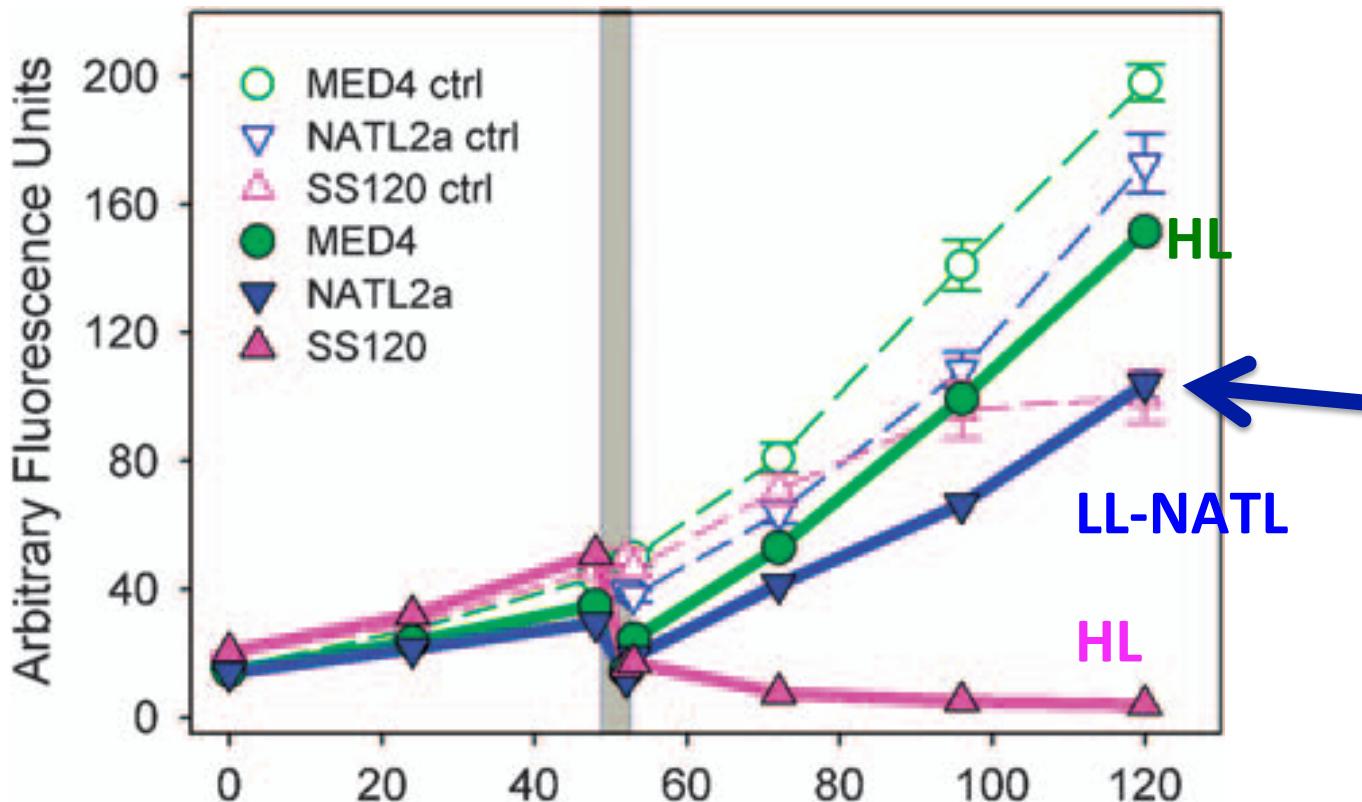
...and deep mixing better than some HL ecotypes



# HYPOTHESIS: NATLs better adapted to fluctuating light

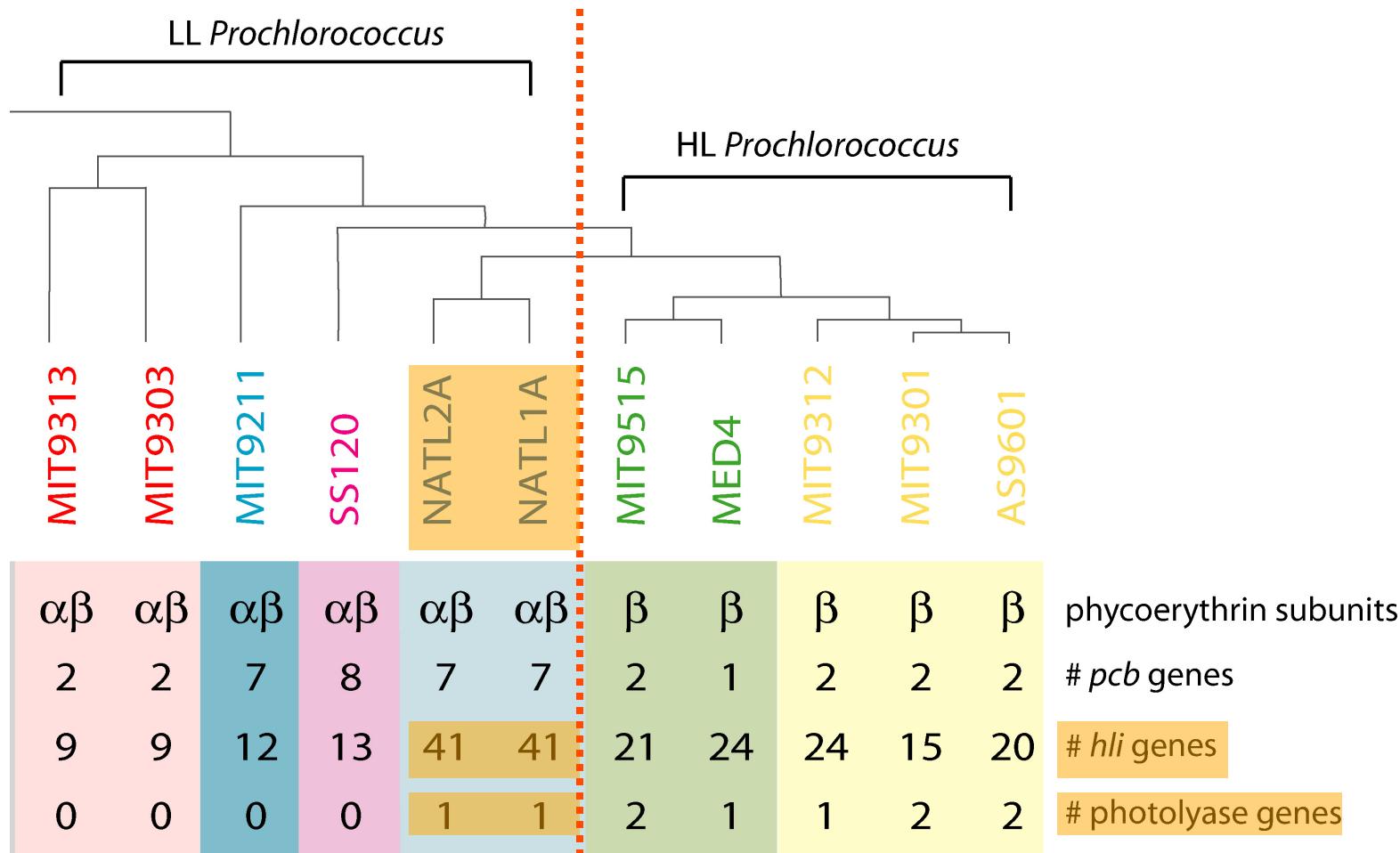


# Back to the lab...



**“The NATLs”**  
are more  
resistant to  
light shock  
than other LL  
strains

# Flashing forward: What do the genomes tell us?



Coleman and Chisholm 2007

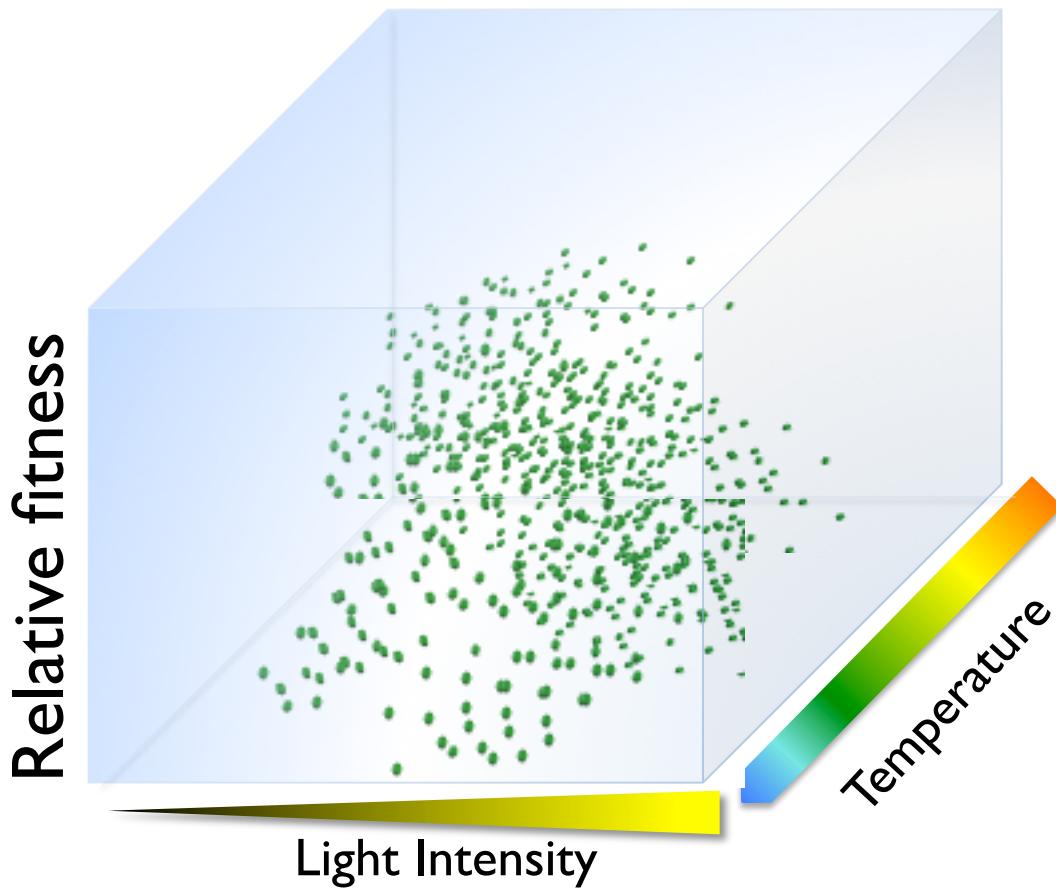
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# Beginning to understand *Prochlorococcus* niche differentiation in two dimensions

BUT Remember...



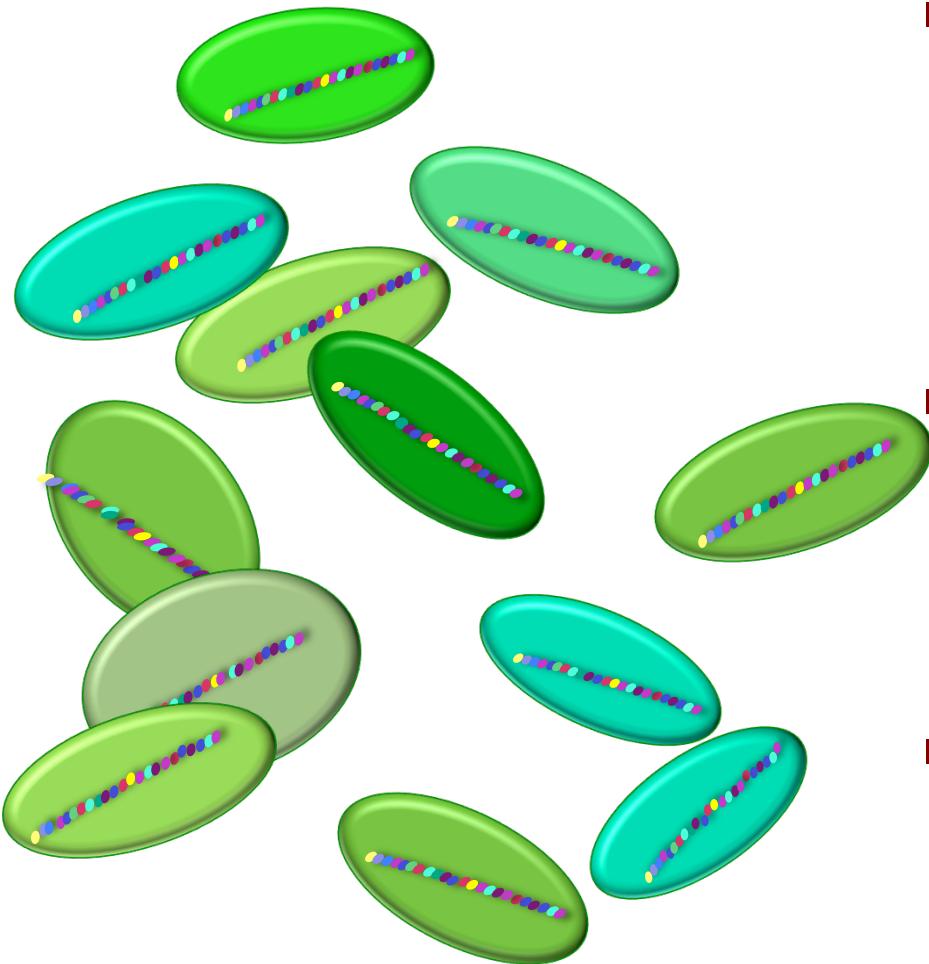
Ecological Niche:

*n*-dimensional hypervolume

*n*-2 dimensions to go!

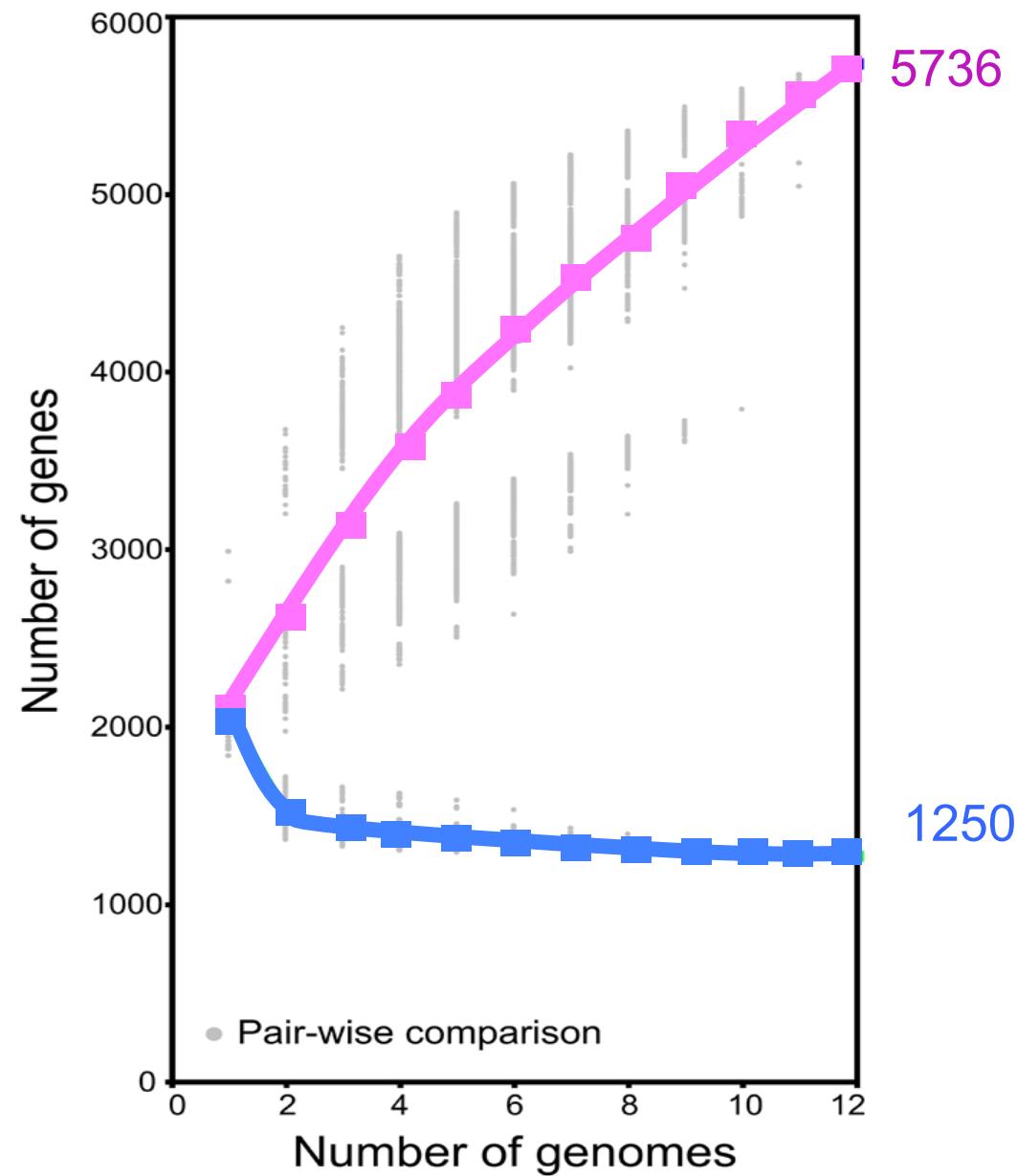
# Enter Genomics:

Genomes of 13 strains (now 45)



- What genes are “core” i.e. shared by all?
- How many unique genes in the gene pool of *ALL* *Prochlorococcus*?
- Their functions?

# The core and flexible genomes – 12 strains

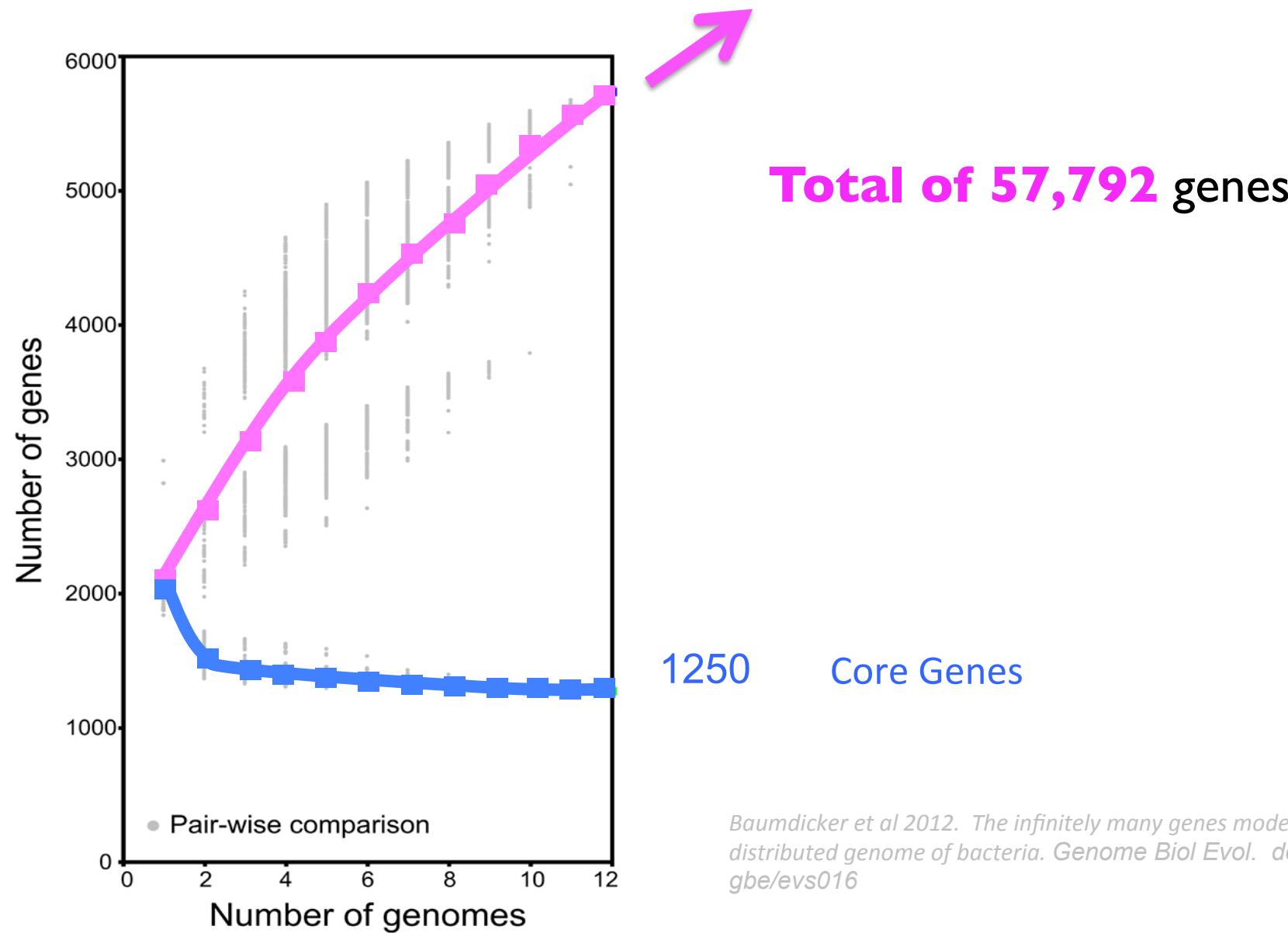


‘Flexible Genes’  
(w/ 45 genomes -12,000)

What is the global  
pan-genome?

1250 Shared Core Genes  
(now about 1000)

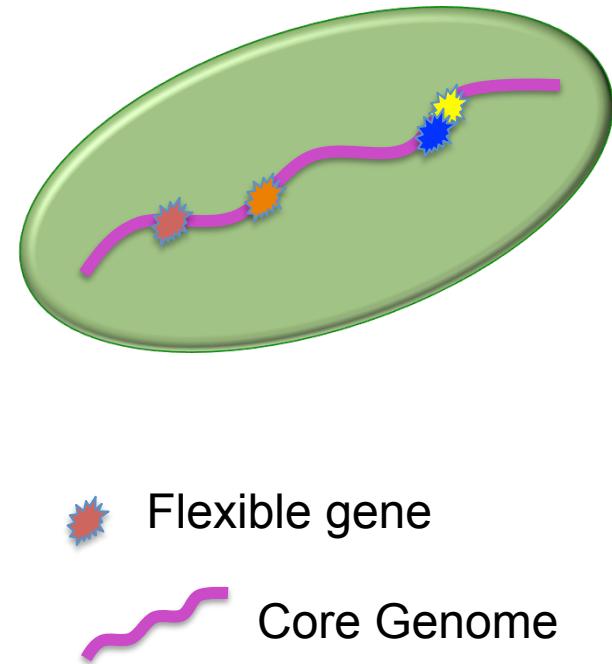
# Global Pan-genome is huge



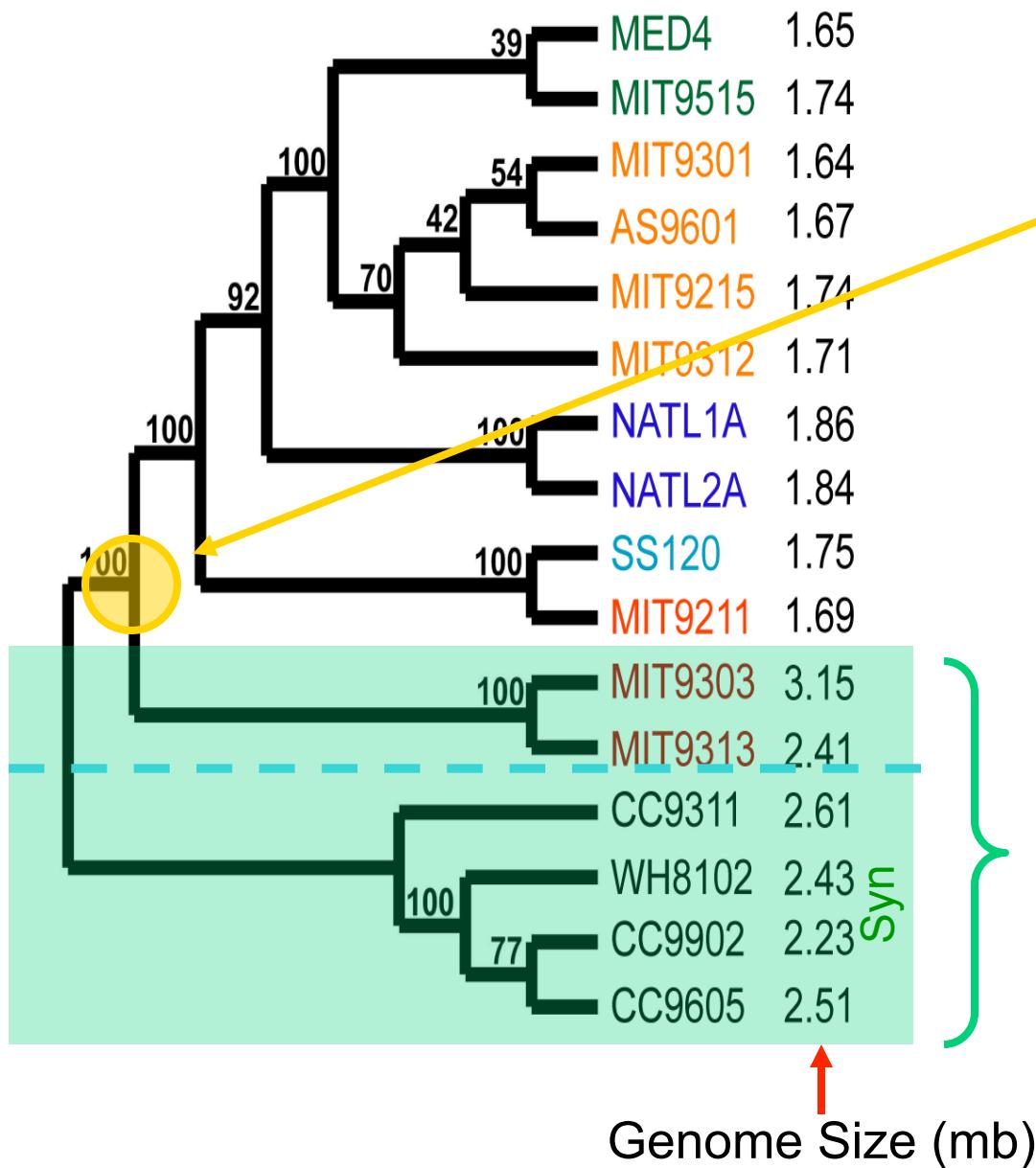
Baumdicker et al 2012. The infinitely many genes model for the distributed genome of bacteria. *Genome Biol Evol*. doi: 10.1093/gbe/evs016

# FLEXIBLE genes give us clues as to environmental pressures

- Viral defense
- Nutrient acquisition
- Stress responses
- 80 % from distant phyla
  - a lot of gene swapping going on out there!



# Genome size in *Prochlorococcus* is variable



Systematic Genome Reduction? NO

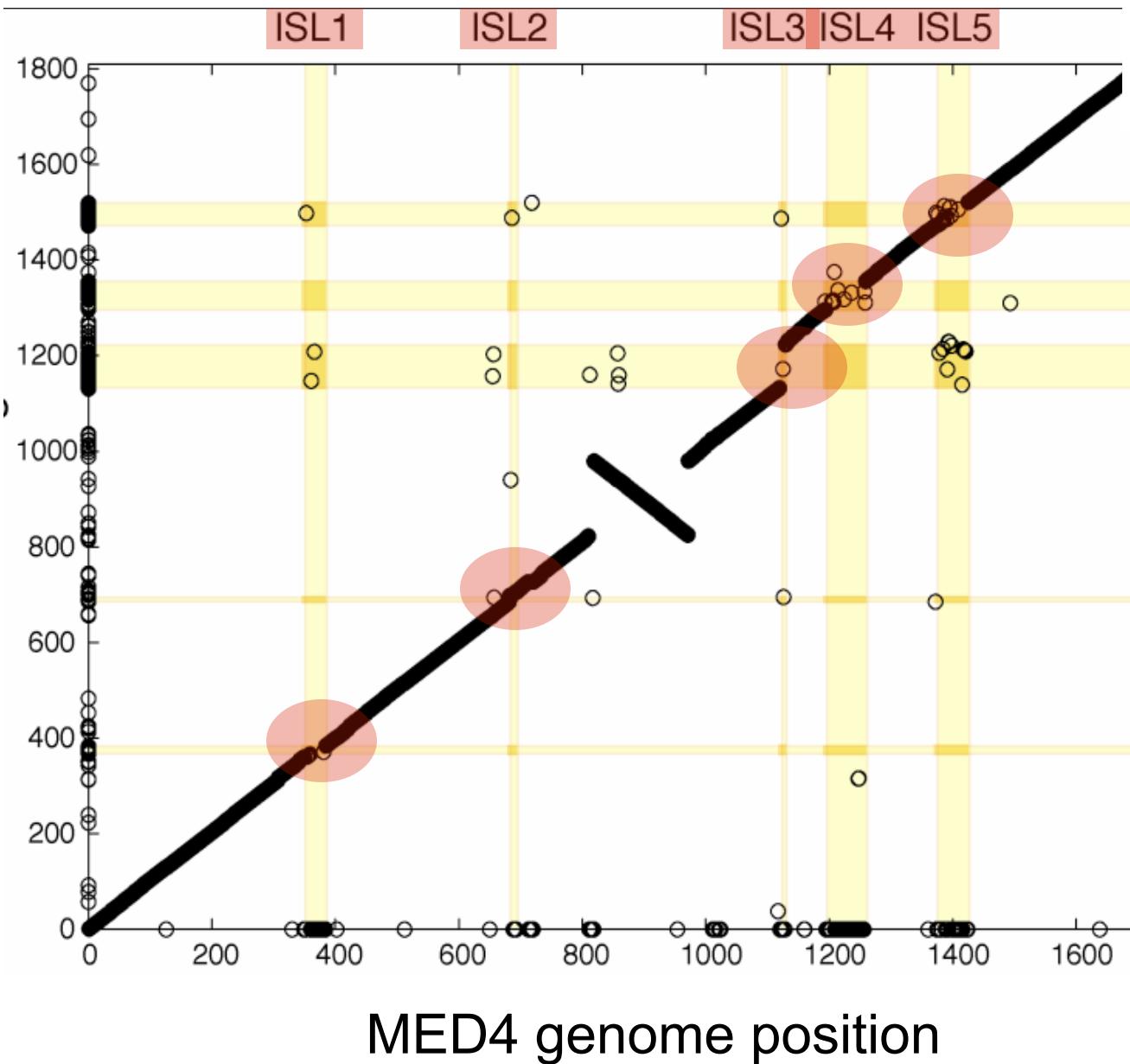
small genomes

Genes are gained and lost – with most of the action in the “leaves of the tree”

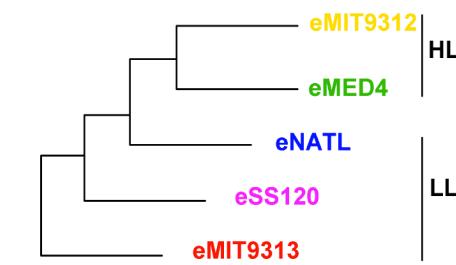
large genomes

# Variability focused in genomic islands

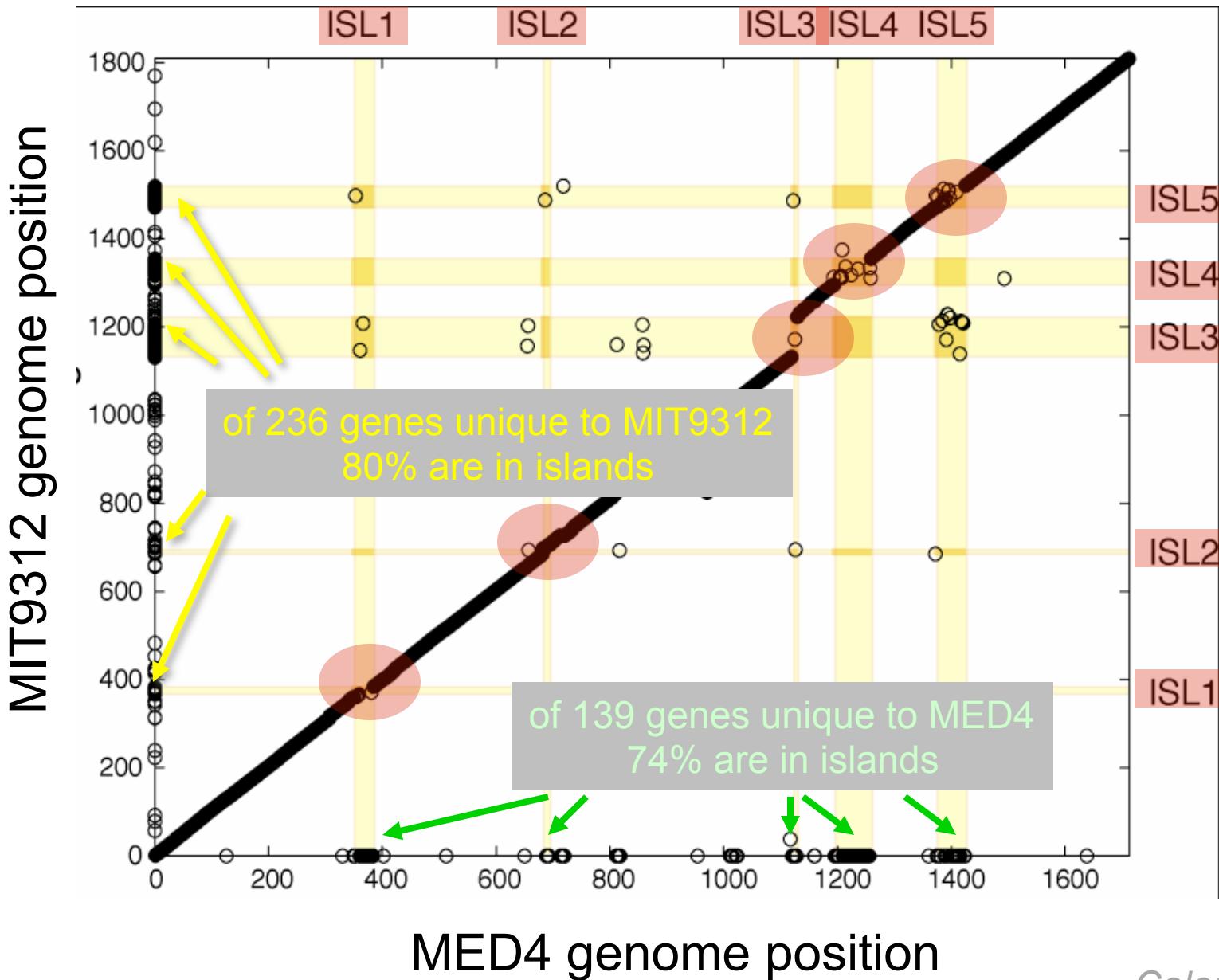
MIT9312 genome position



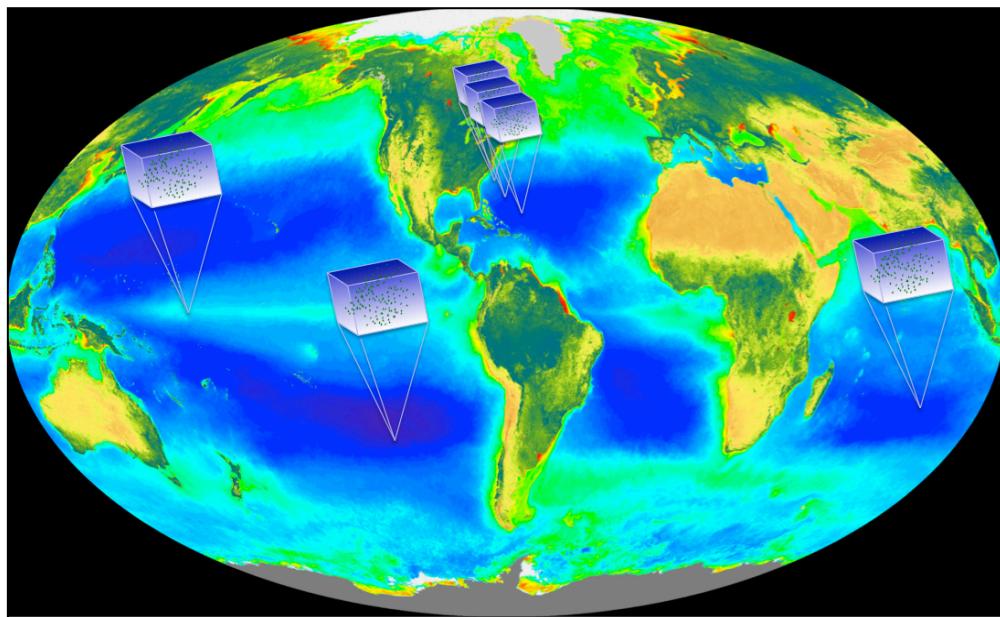
MED4 genome position



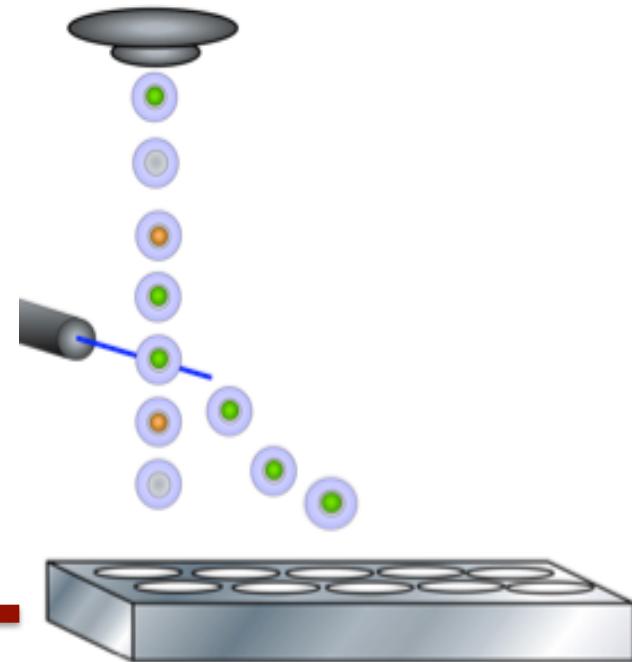
# Variability focused in genomic islands



# Exploring the full diversity through single cell genomics of wild *Prochlorococcus*



Flow Sort individual cells

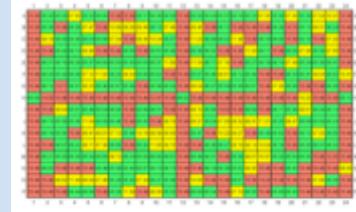


Select cells for sequencing



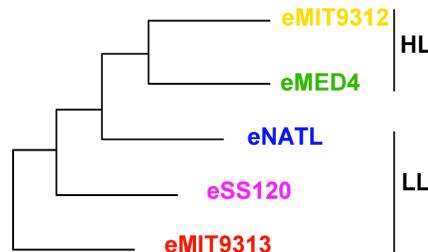
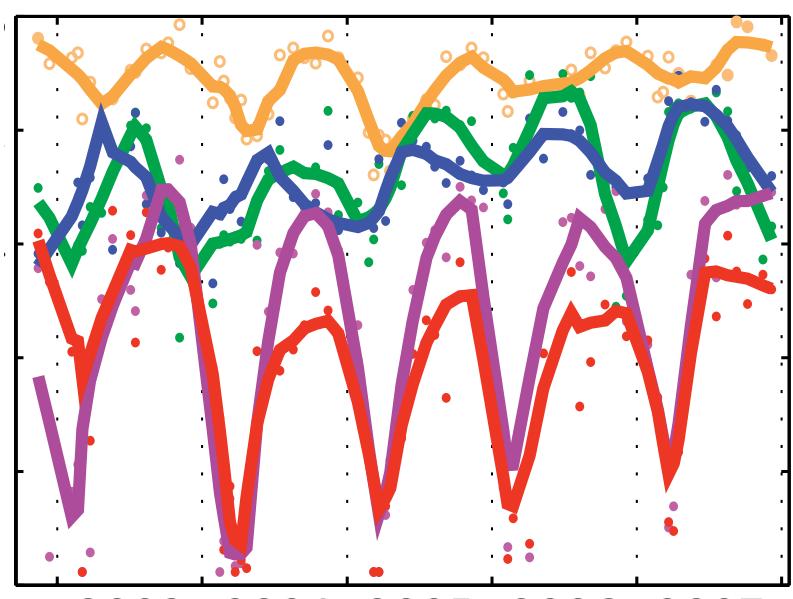
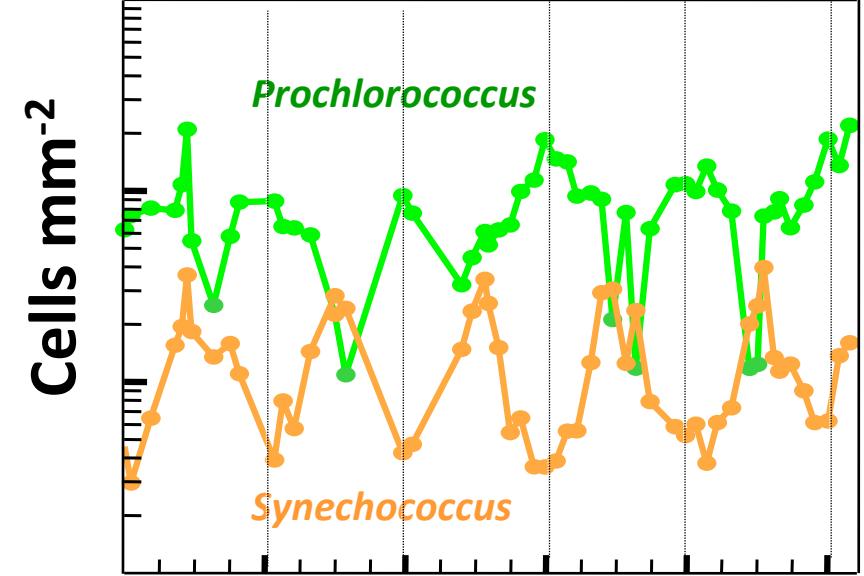
Sequence Genomes

DNA Amplification

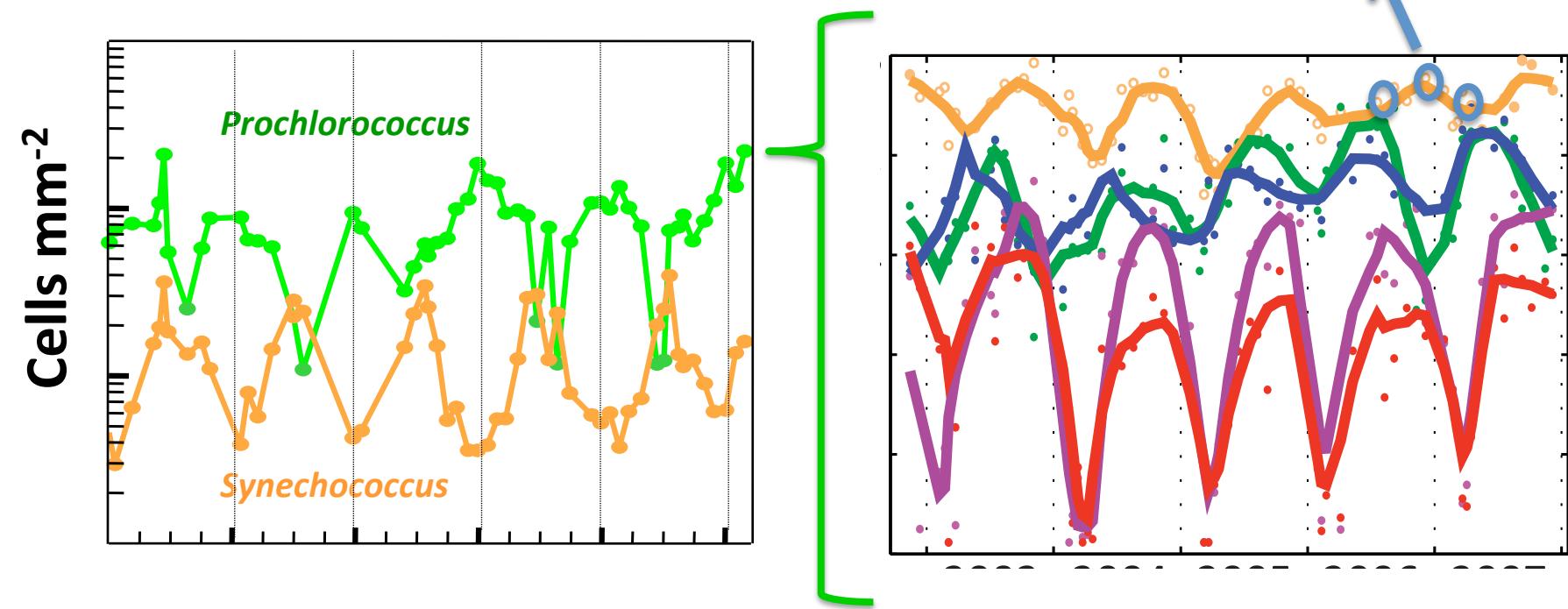
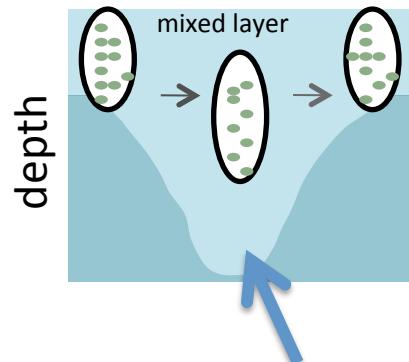


# Remember this?

*Prochlorococcus* ecotypes

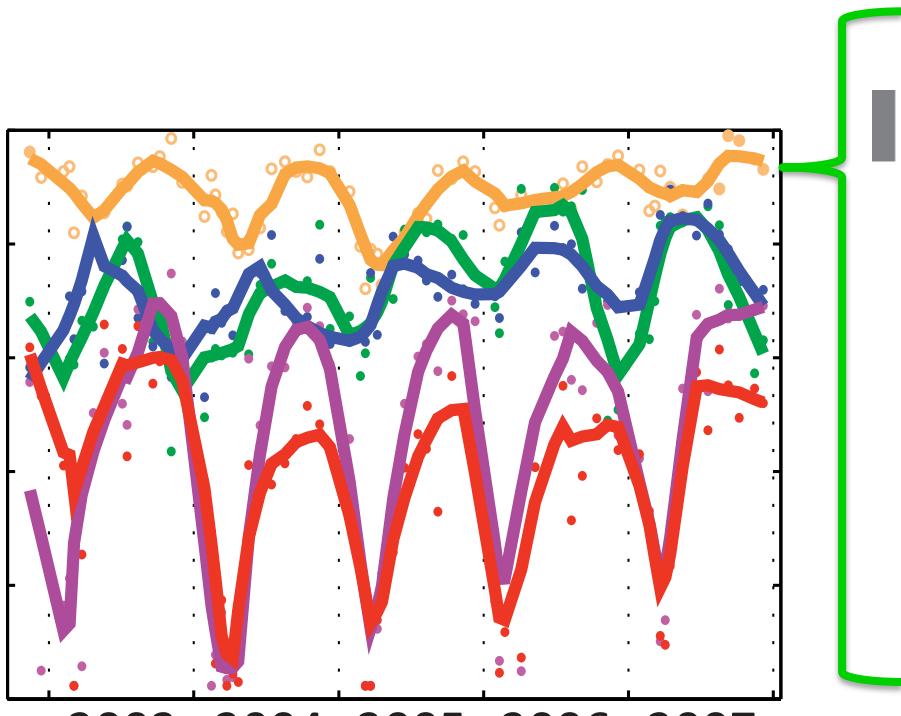


Thousands of single cell rRNA ITS sequence, and hundreds of (partial) genomes



# Astounding Diversity!

(layer upon layer...leading to stability)



REPORTS

## Single-Cell Genomics Reveals Hundreds of Coexisting Subpopulations in Wild *Prochlorococcus*

Nadav Kashtan,<sup>1\*</sup> Sara E. Roggensack,<sup>1</sup> Sébastien Rodrigue,<sup>1,2</sup> Jessie W. Thompson,<sup>1</sup> Steven J. Biller,<sup>1</sup> Allison Coe,<sup>1</sup> Huiming Ding,<sup>1,3</sup> Pekka Marttinen,<sup>4</sup> Rex R. Malmstrom,<sup>5</sup> Roman Stocker,<sup>1</sup> Michael J. Follows,<sup>6</sup> Ramunas Stepanauskas,<sup>7</sup> Sallie W. Chisholm<sup>1,3\*</sup>

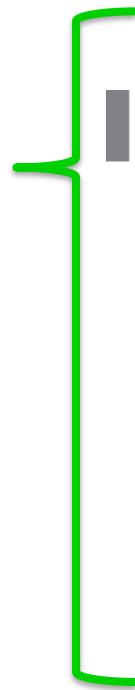
- Roughly 5000 new genes added to pan-genome
- ‘Genomic backbones’ (core allele variation + associated flexible genes)



# Astounding Diversity!

(layer upon layer...leading to stability)

- Populations well-mixed  
10 km<sup>2</sup> × 3 m (1 week)
- Each backbone sub-population > 10<sup>13</sup> cells
- ≈ Effective Population size



REPORTS

## Single-Cell Genomics Reveals Hundreds of Coexisting Subpopulations in Wild *Prochlorococcus*

Nadav Kashtan,<sup>1\*</sup> Sara E. Roggensack,<sup>1</sup> Sébastien Rodrigue,<sup>1,2</sup> Jessie W. Thompson,<sup>1</sup> Steven J. Biller,<sup>1</sup> Allison Coe,<sup>1</sup> Huiming Ding,<sup>1,3</sup> Pekka Marttinen,<sup>4</sup> Rex R. Malmstrom,<sup>5</sup> Roman Stocker,<sup>1</sup> Michael J. Follows,<sup>6</sup> Ramunas Stepanauskas,<sup>7</sup> Sallie W. Chisholm<sup>1,3\*</sup>



# The assembly of diversity

A

## PRIMARY BUILDING BLOCKS OF *PROCHLOROCOCCUS* DIVERSITY

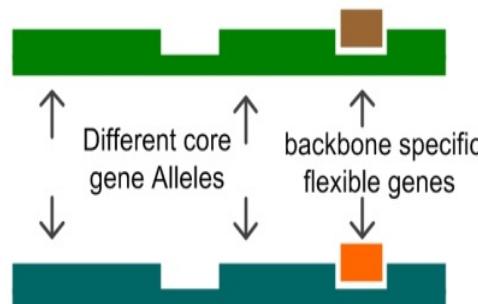
Distinct variants of core gene alleles

Flexible gene pool



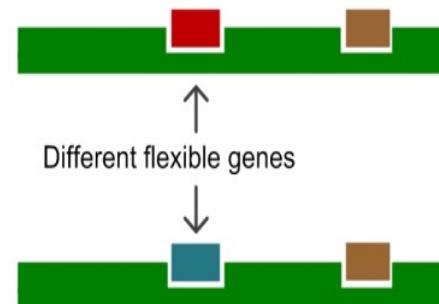
B

Each backbone consists of a distinct variant of core gene alleles and a small distinct set of flexible genes



C

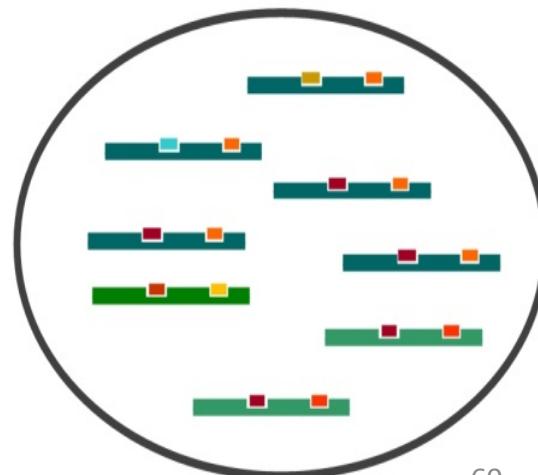
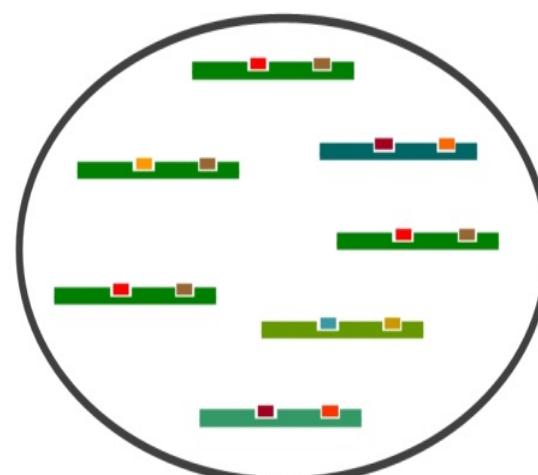
Cells within the same backbone may differ in some flexible genes in island regions



D

Population selected by Environment 1

Population selected by Environment 2



# Overview

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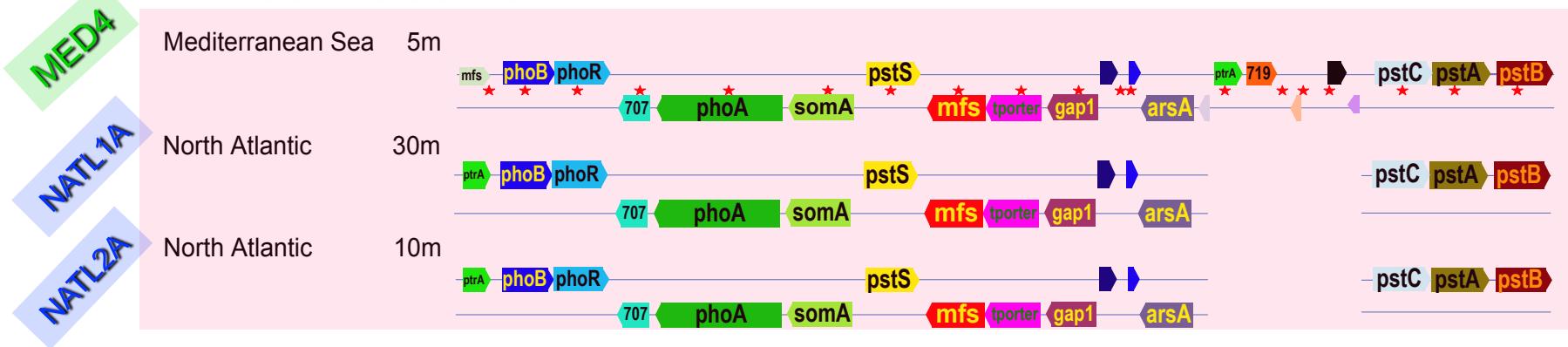
- ◆ History
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- ◆ Niche Dimensions of *Prochlorococcus*
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  - Genomics and Niche Dimensions:
    - Phosphorus
    - Nitrogen
    - Iron
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- ◆ Marine Vesicles
- ◆ Integrative Systems Biology

# *Prochlorococcus* is P–challenged

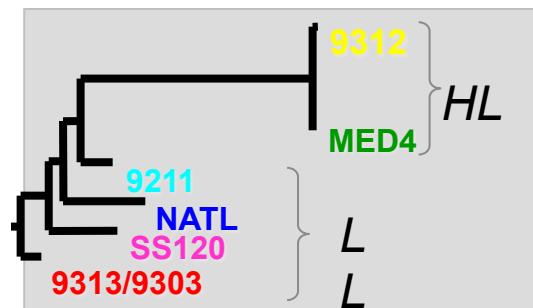
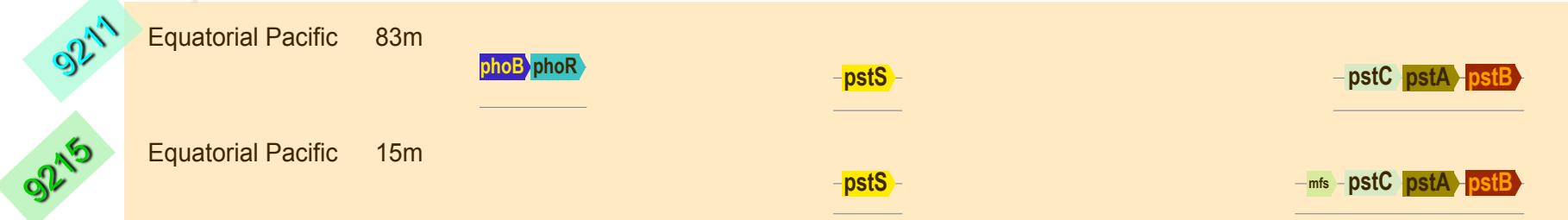
- 13 amole P per cell
- N:P Ratio higher than Redfieldian (21-62)  
(Bertilsson et al 2003)
- Sulfo- instead of Phospho- lipids  
(Van Mooy et al 2006)
- 96% P is in nucleic acids  
(Waldbauer 2010)

# *phoBR* genomic region (phosphorus acquisition) in different strains is dictated by ocean of origin and not ribotype

## P-limited oceans



## P-replete ocean

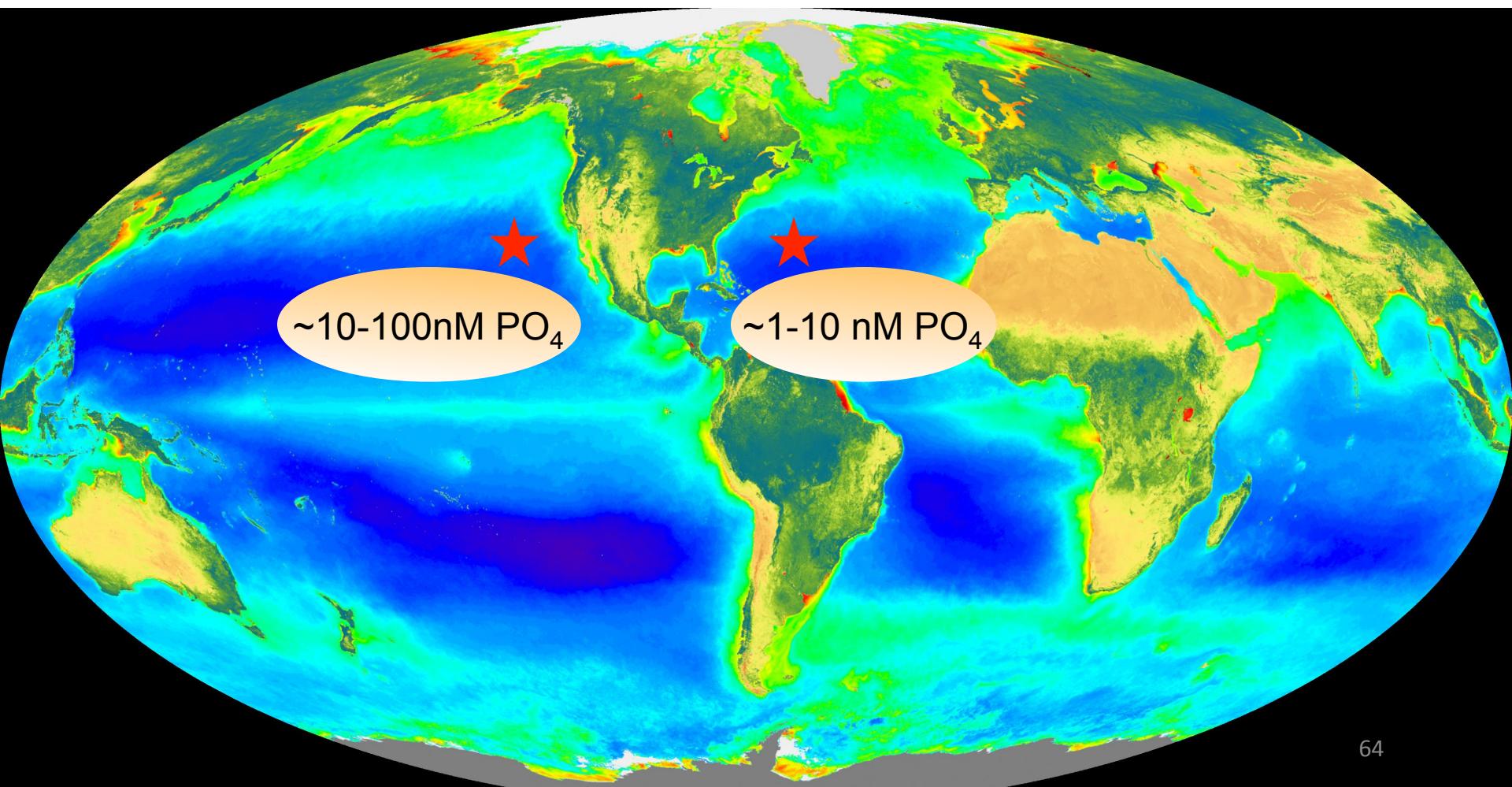


Martiny, Coleman et al 2006

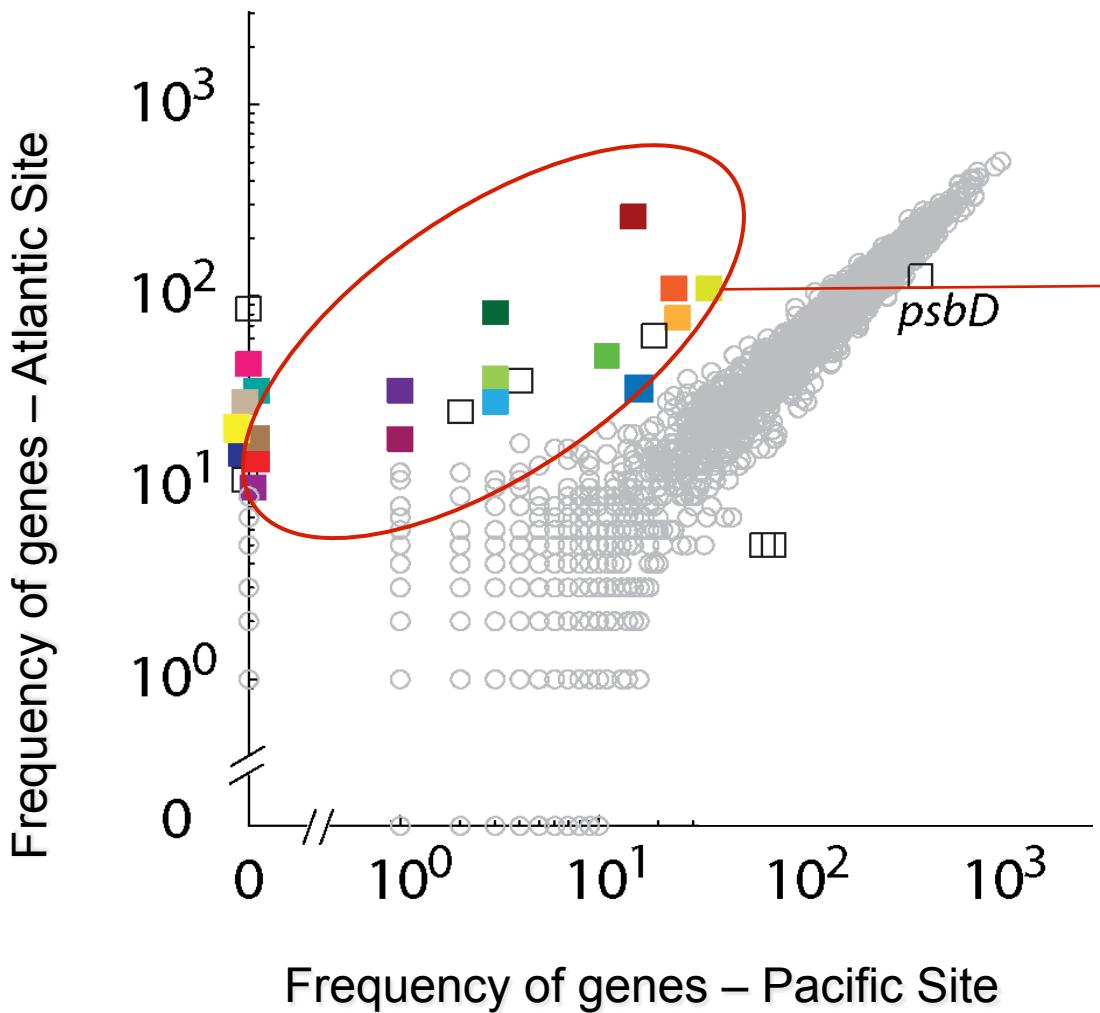
Martiny, et al 2009

# Back to the oceans...

## Comparing the *Prochlorococcus* populations in the Atlantic and Pacific



For *Prochlorococcus*, the only major difference between the two ecosystems is ...



P- acquisition  
genes

+

Arsenate  
reductase and  
arsenite efflux  
genes

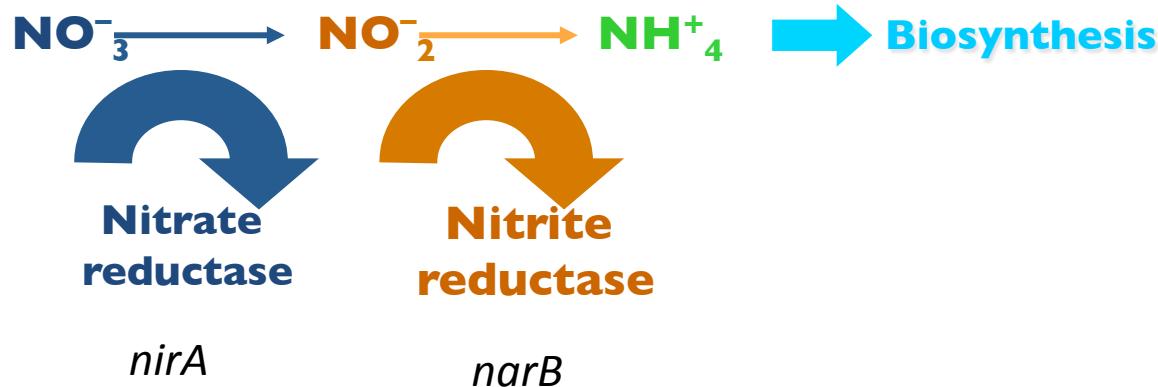
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- ◆ Marine Vesicles!
- ◆ Integrative Systems Biology

# *Prochlorococcus* and N sources:

*Synechococcus* (like “the ancestor”)

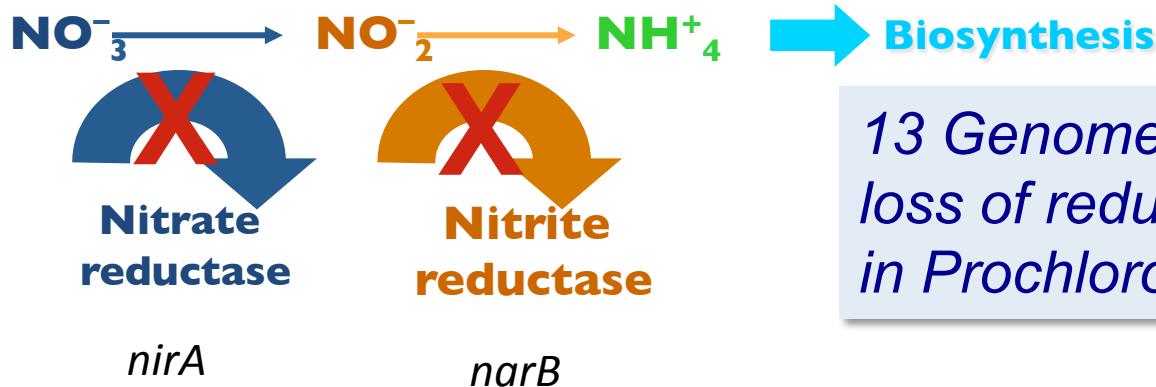


The *Prochlorococcus* dogma (based on cultured isolates):

- ❖ All *Prochlorococcus* can use  $\text{NH}_4^+$
- ❖ Some can use  $\text{NO}_2^-$
- ❖ None can use  $\text{NO}_3^-$  (odd...)

# Prochlorococcus and N sources:

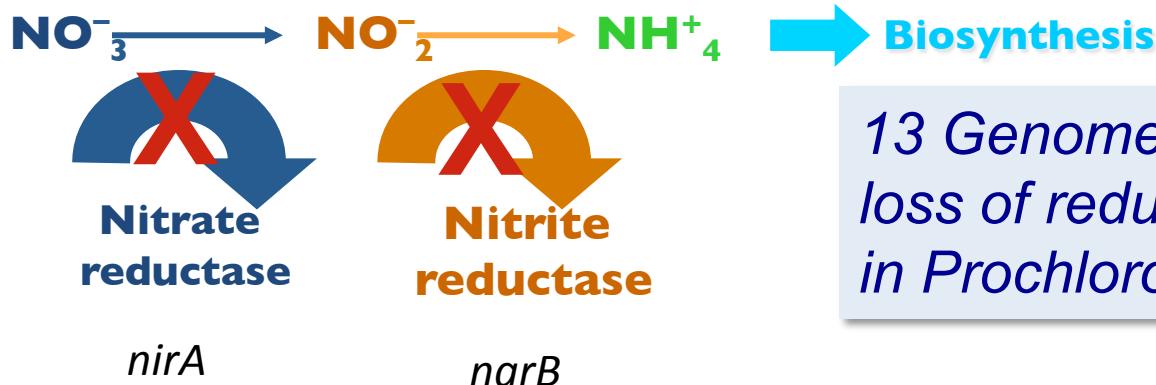
*Synechococcus* (like “the ancestor”)



13 Genomes confirmed  
loss of reductase genes  
in *Prochlorococcus*

# *Prochlorococcus* and N sources:

*Synechococcus* (like “the ancestor”)



13 Genomes confirmed  
loss of reductase genes  
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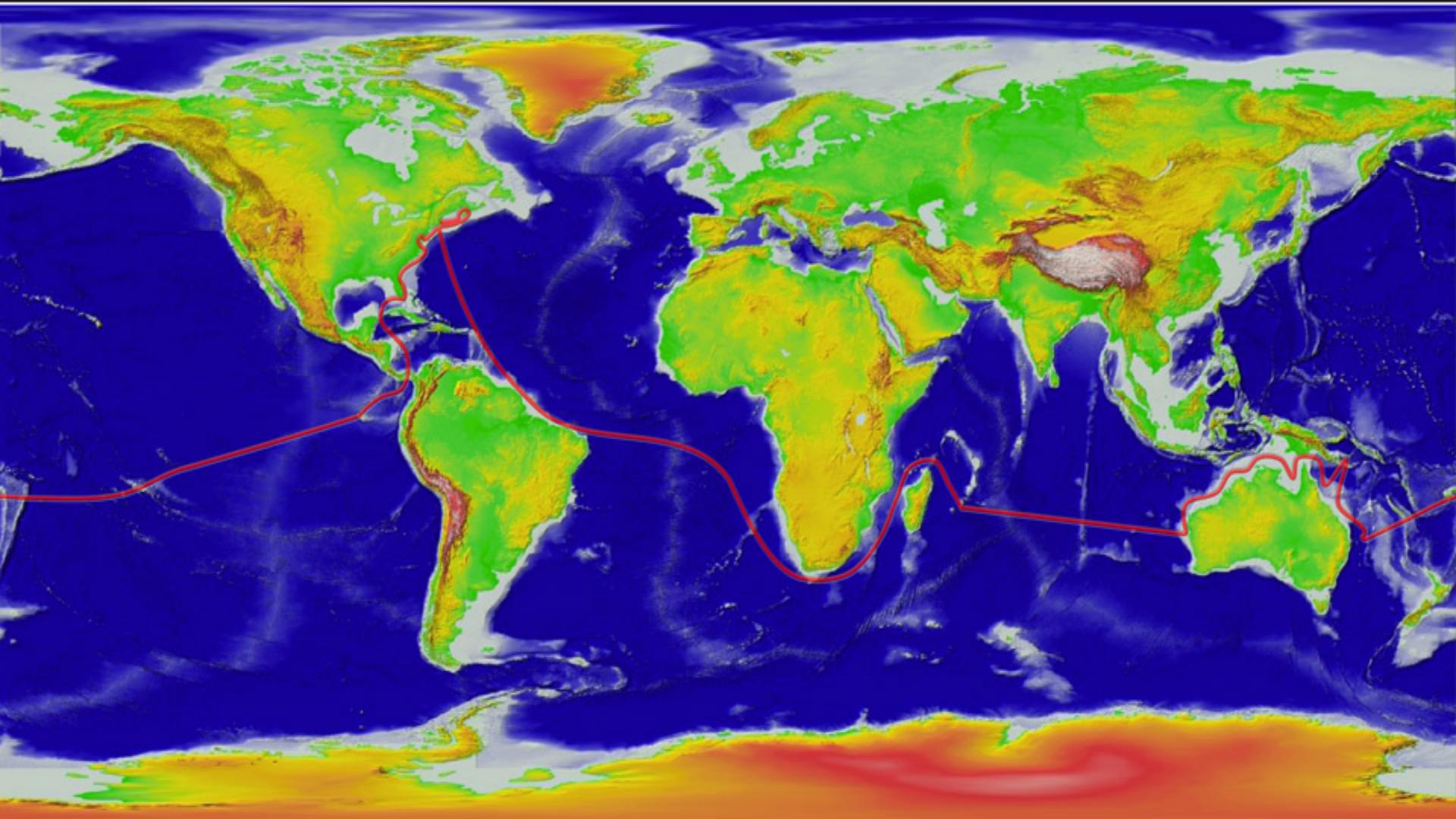
BUT... There was some evidence for  $\text{NO}_3^-$  utilization

CULTURES Williams EZ, Campbell L, DiTullio G. (1999). ASLO Aquatic Sciences Meeting – reported on  $\text{NO}_3^-$  utilizing *Prochlorococcus*

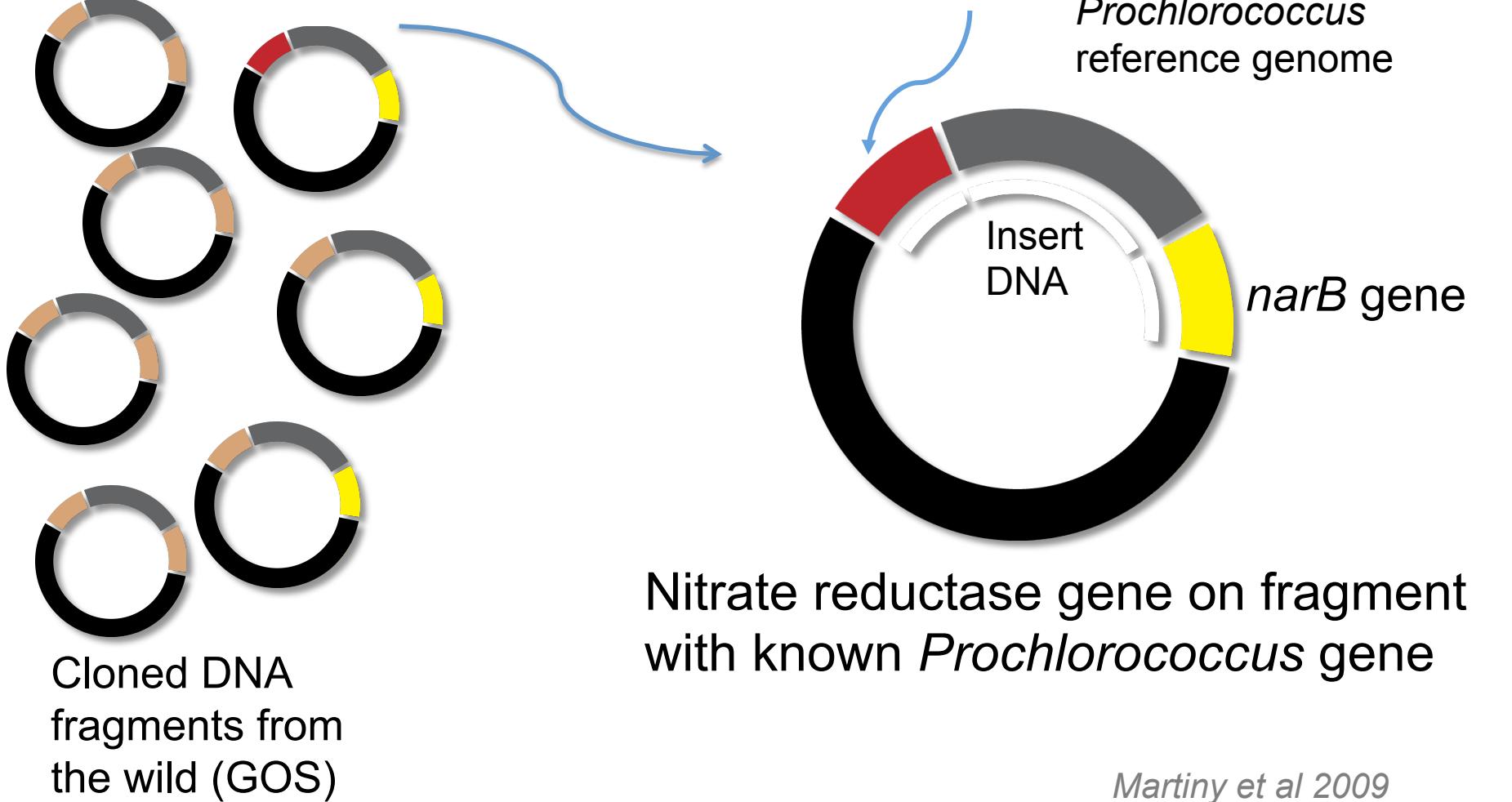
WILD Casey JR, Lomas MW, Mandecki J, Walker DE. (2007). *Prochlorococcus* contributes to new production in the Sargasso Sea deep chlorophyll maximum. Geophysical Research Letters 34: L10604.

# Turning to GOS again...

*Adam Martiny and Paul Berube*

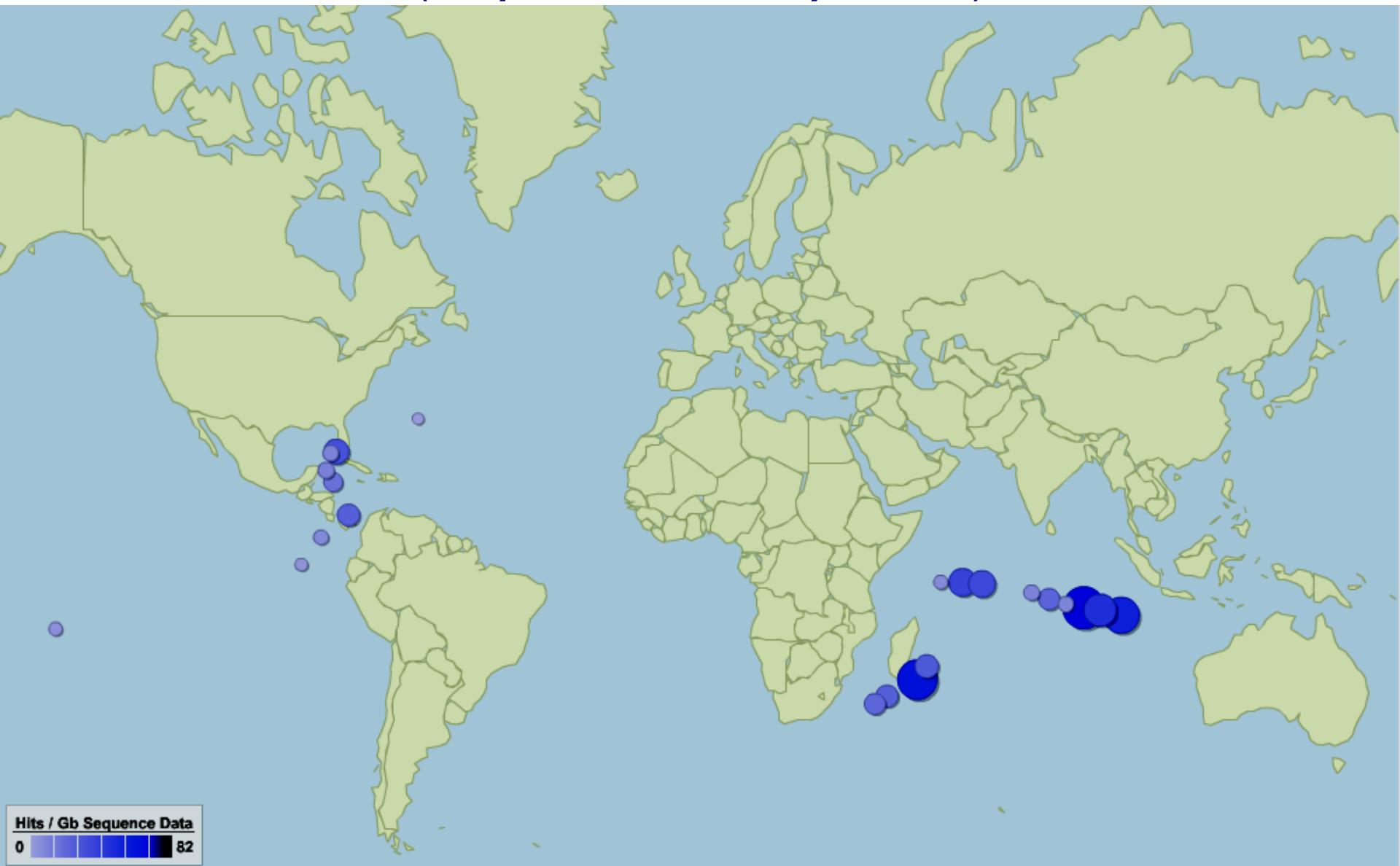


- Recruit all fragments with *narB* gene
- Find those containing known *Prochlorococcus* gene



*Martiny et al 2009*

# Abundance distribution of *Prochlorococcus narB* genes in GOS (they are not everywhere)

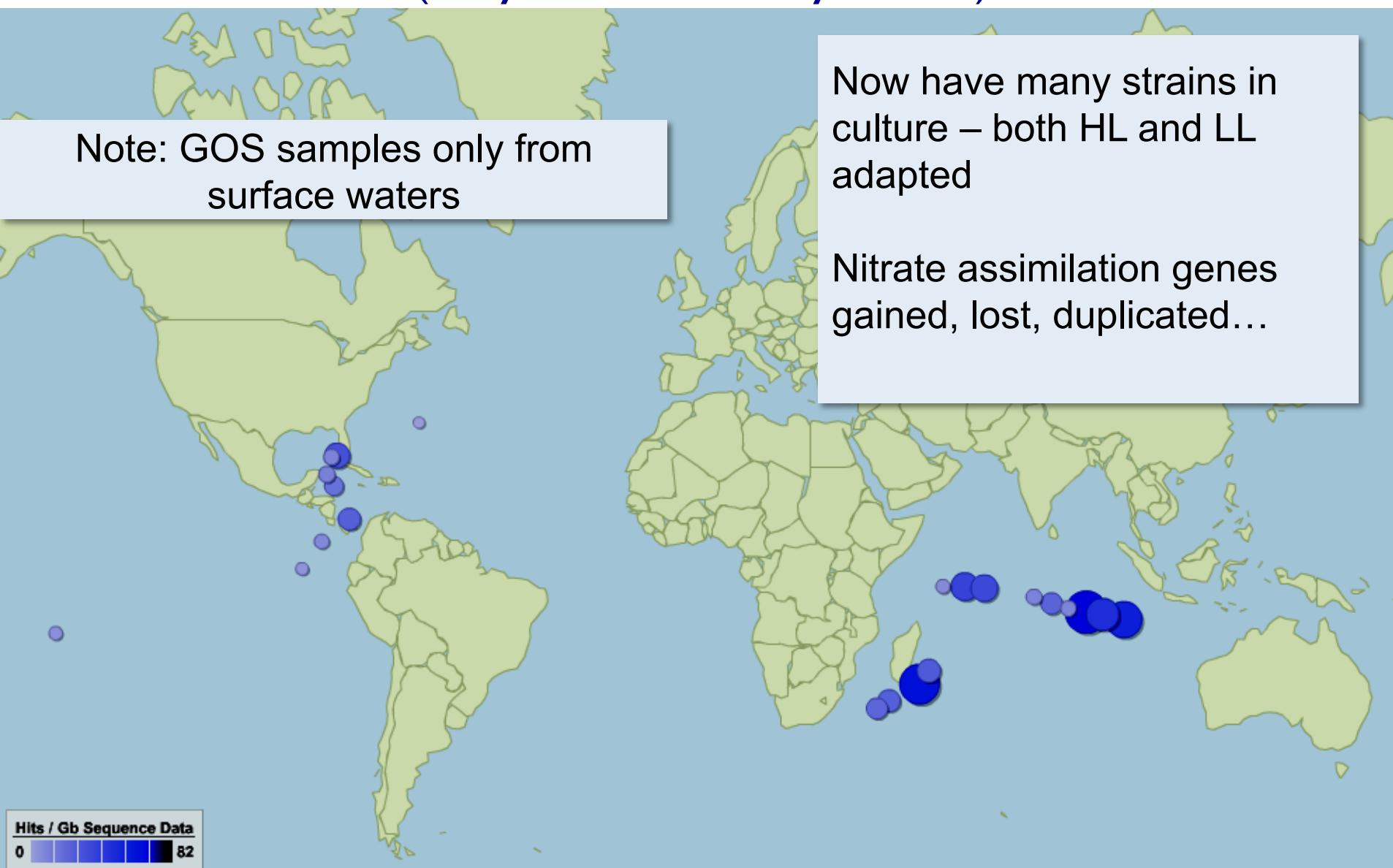


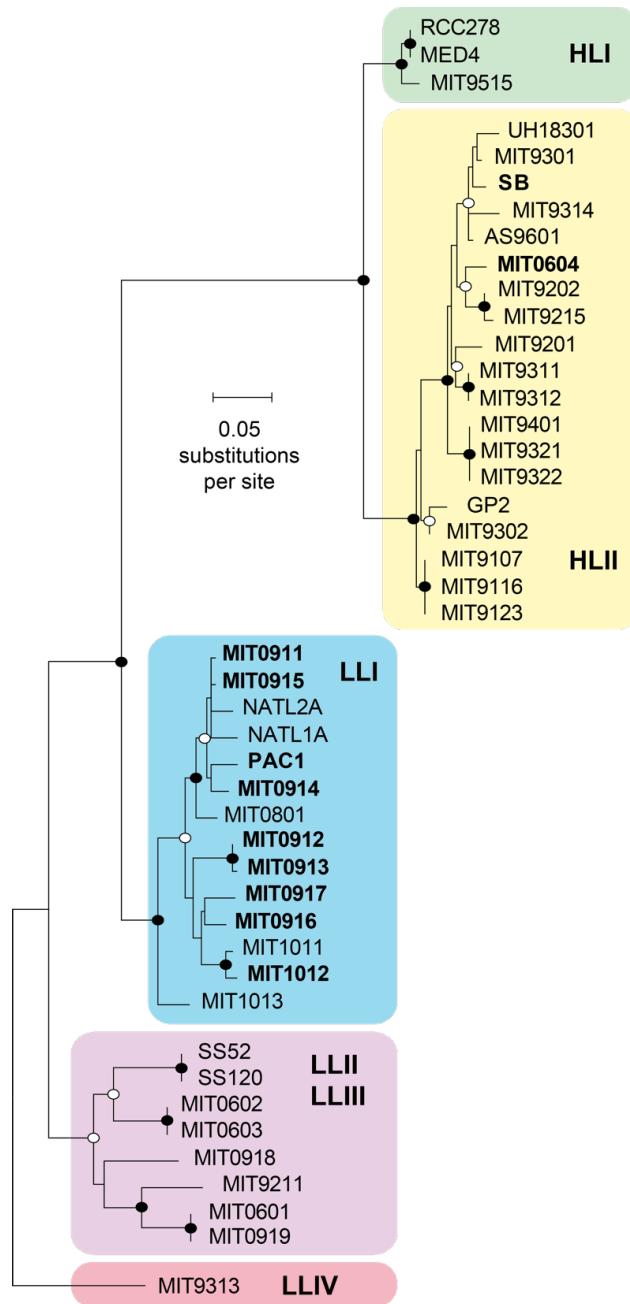
# Abundance distribution of *Prochlorococcus narB* genes in GOS (they are not everywhere)

Note: GOS samples only from surface waters

Now have many strains in culture – both HL and LL adapted

Nitrate assimilation genes gained, lost, duplicated...

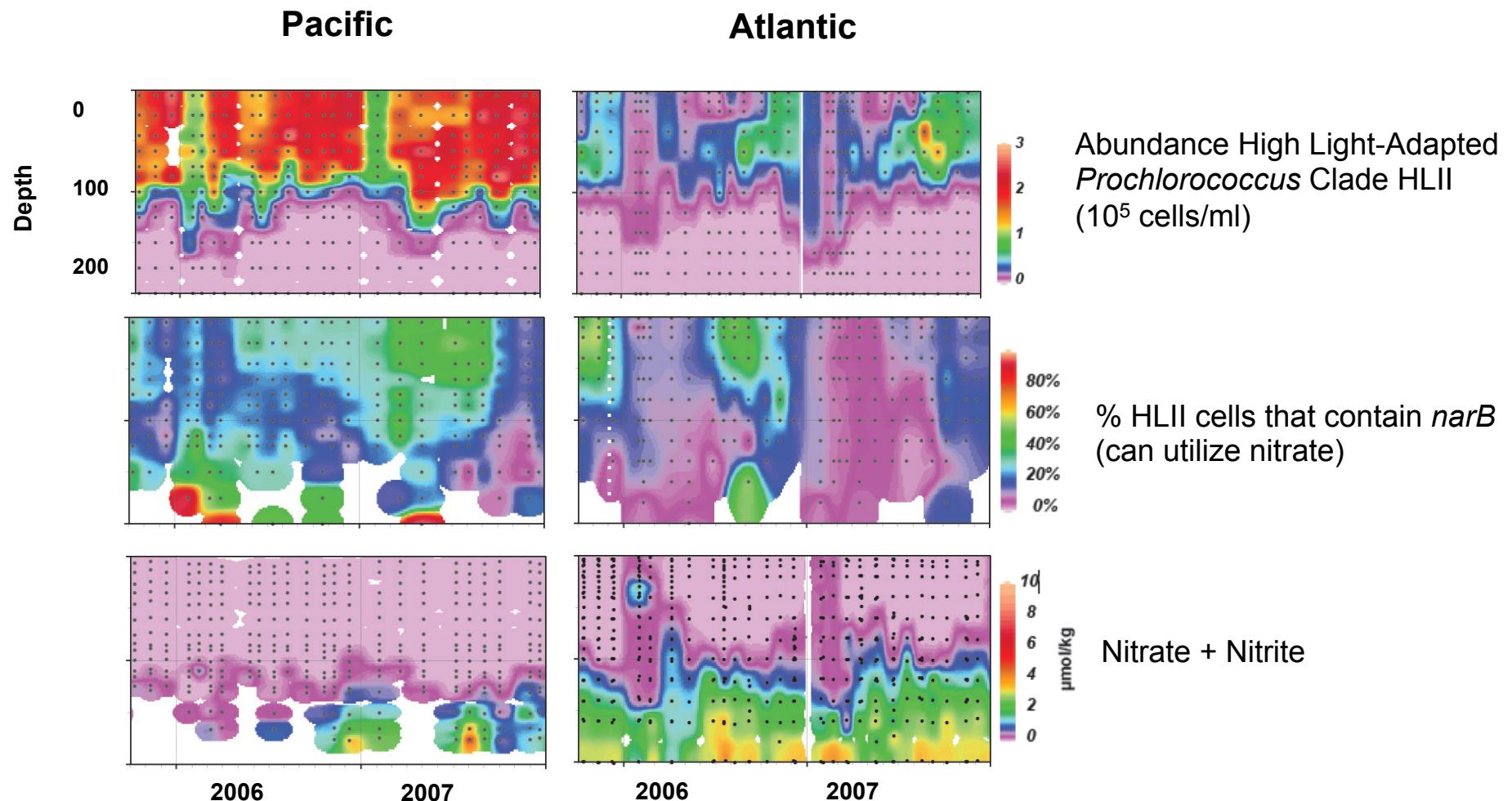




*Prochlorococcus* isolates (**bold**)  
that can utilize nitrate – i.e.  
contain narB  
(a number are axenic)

- We suspect that all clades will ultimately have representatives
- This trait, like P-acquisition, does not follow taxonomy

# Dynamics of *narB* - containing *Prochlorococcus* in the wild (they are a fraction of the population)

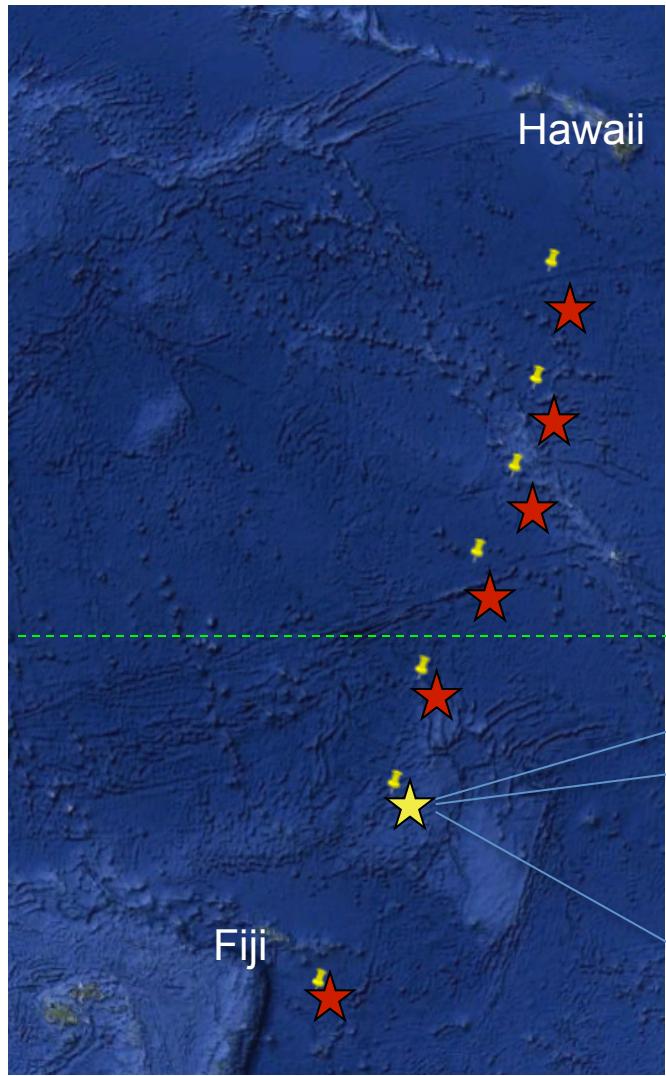


# Overview

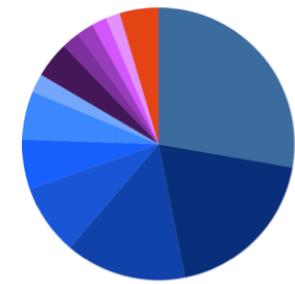
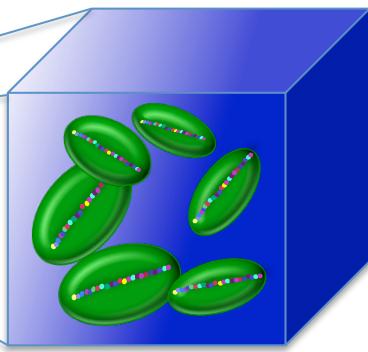
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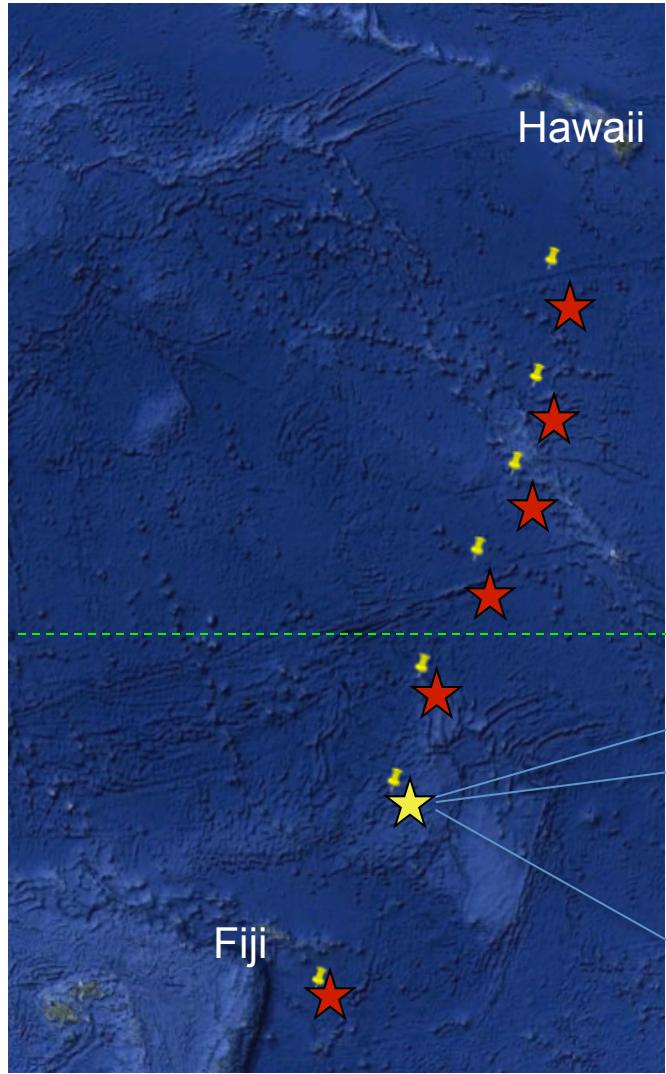
# Genomes of 5 Wild *Prochlorococcus* Cells



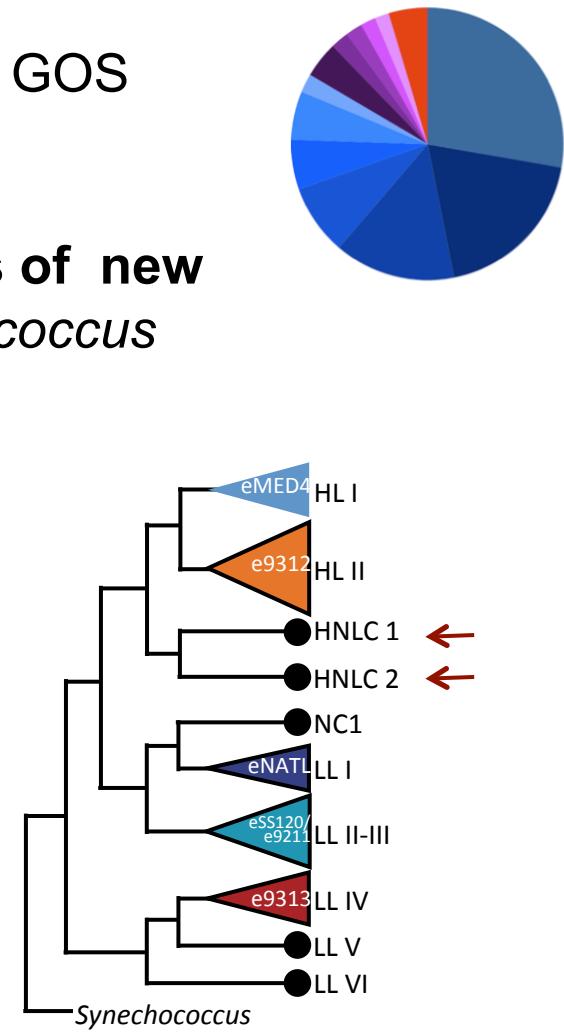
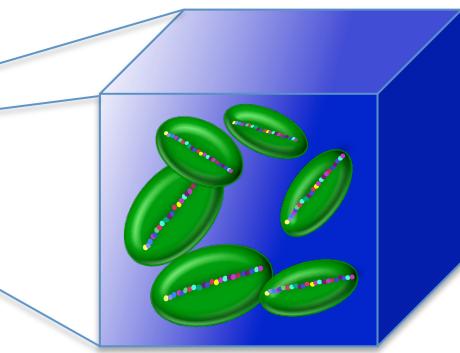
- ❑ Expanded recruited GOS reads by 15%
- ❑ Added to **hundreds of new genes** to *Prochlorococcus* pan-genome
- ❑ New functions



# Genomes of 5 Wild *Prochlorococcus* Cells



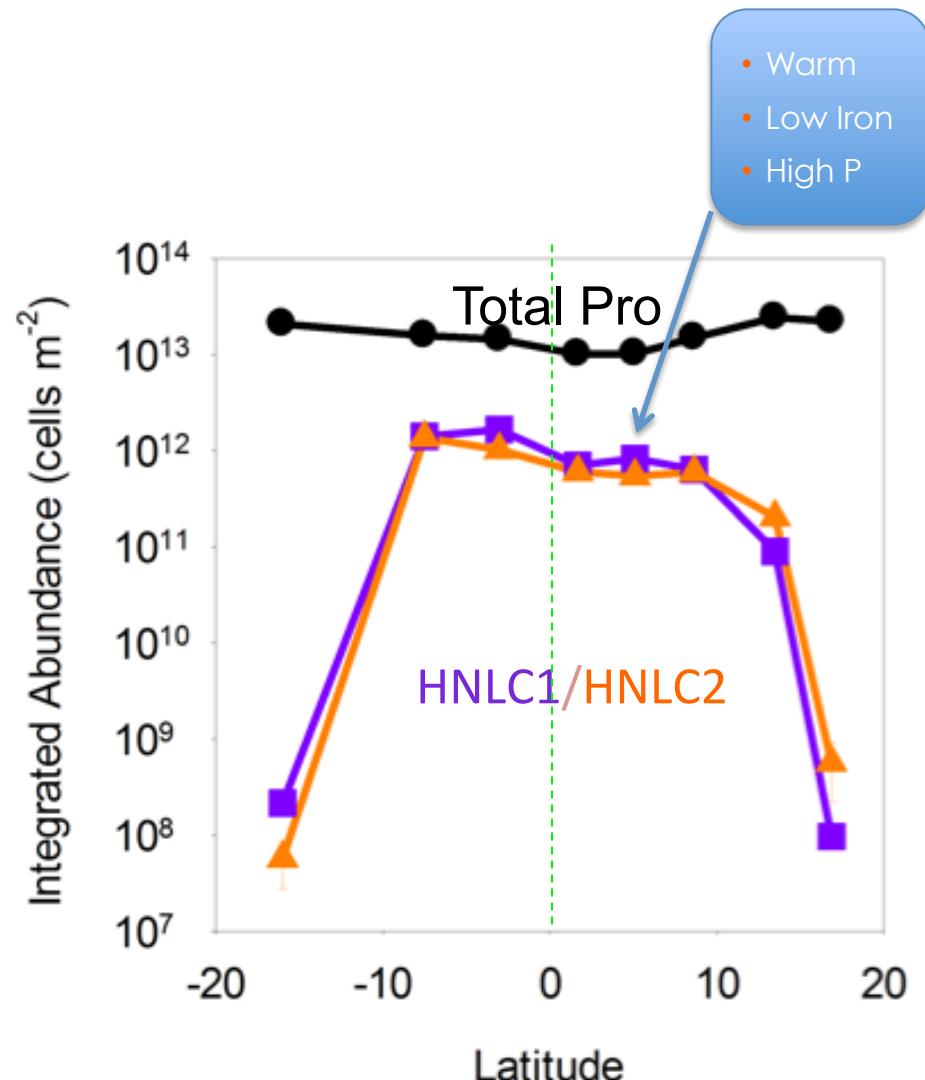
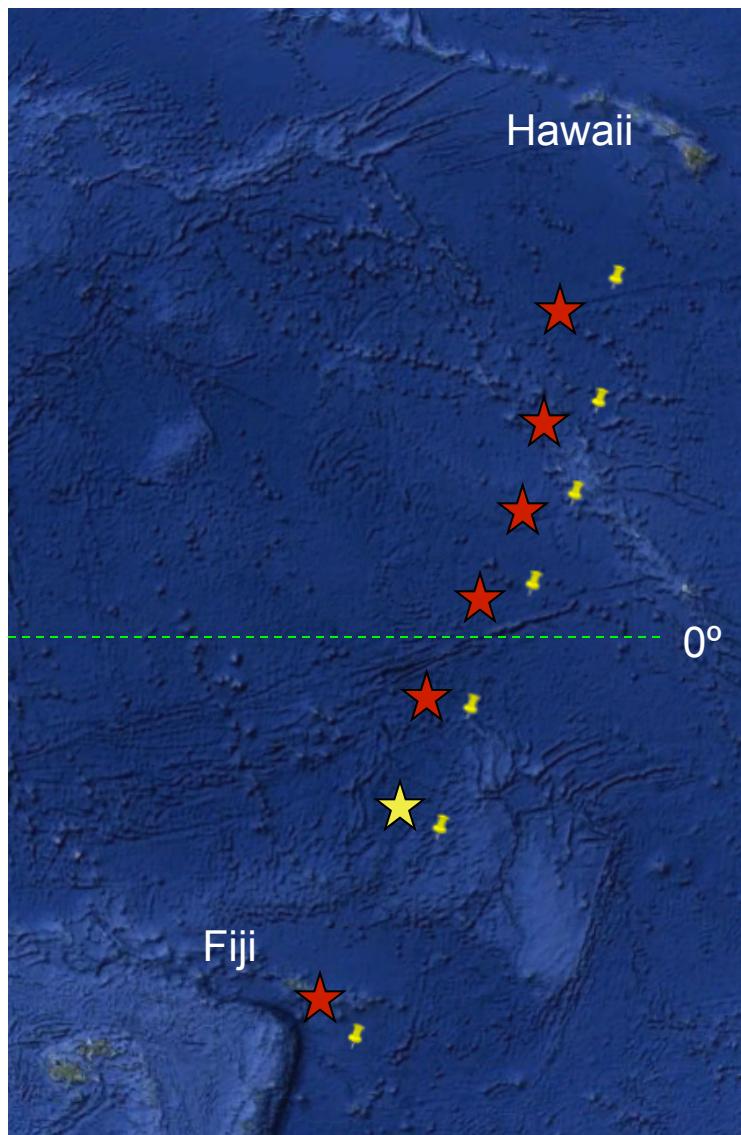
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- ❑ New functions



Malmstrom, Rodrigue, 2012

Rusch et al 2010,  
West et al 2011

# New HL clades restricted to equatorial waters



# Siderophore transport genes found in wild cells, and in one of our cultures

Culture - MIT9292

*Prochlorococcus* MIT9202



Wild single cell

*Prochlorococcus* W12



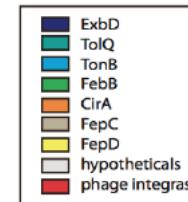
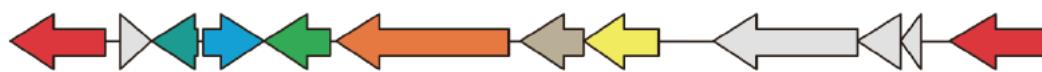
Wild single cell

*Prochlorococcus* W4

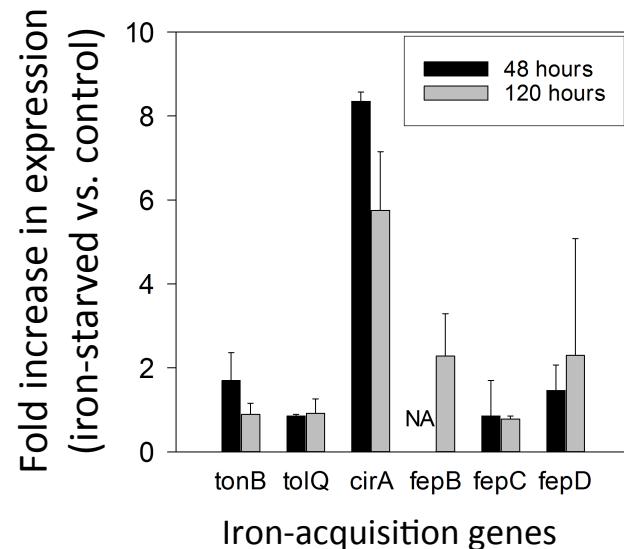


Wild single cell

*Prochlorococcus* W2

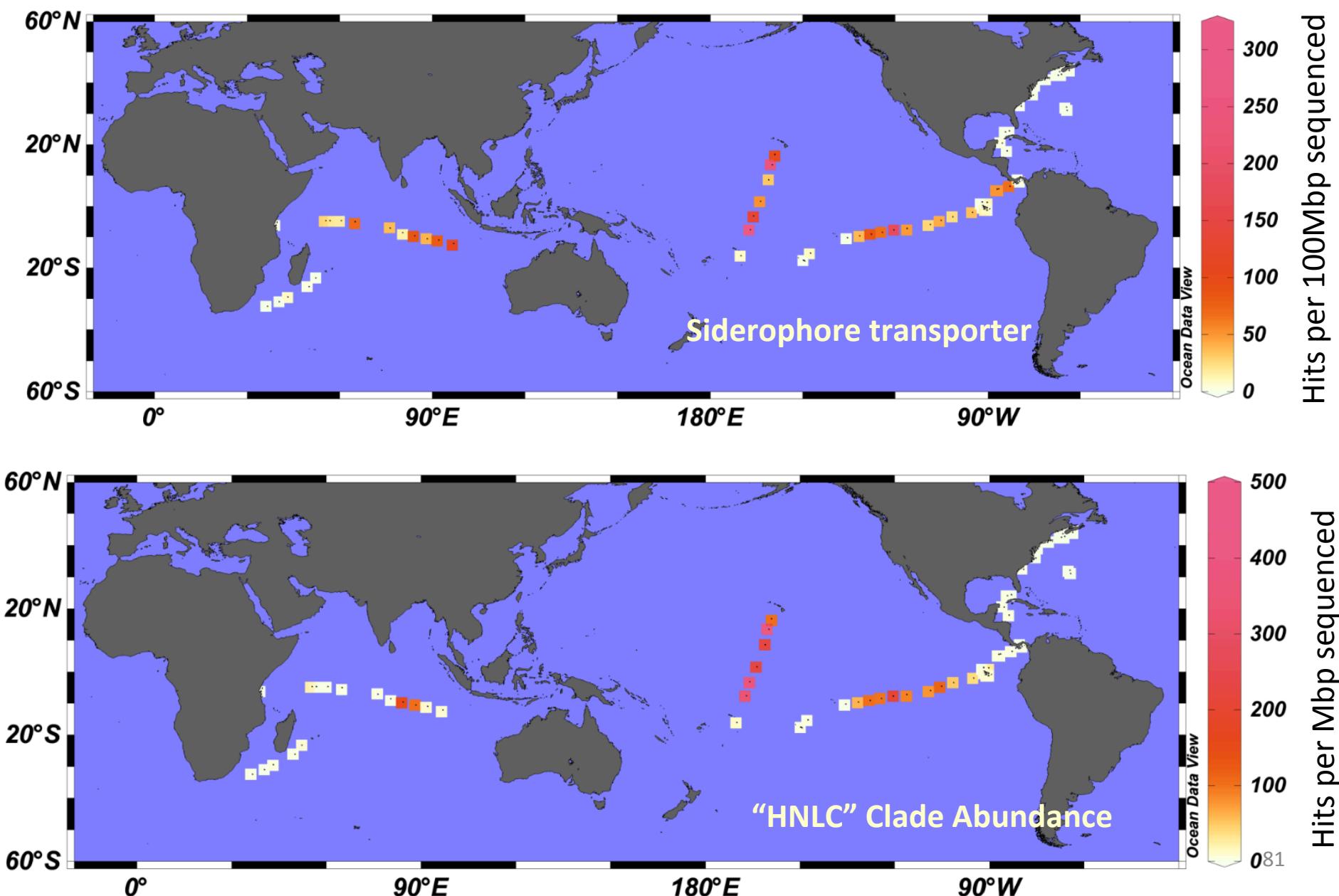


Phage integrase

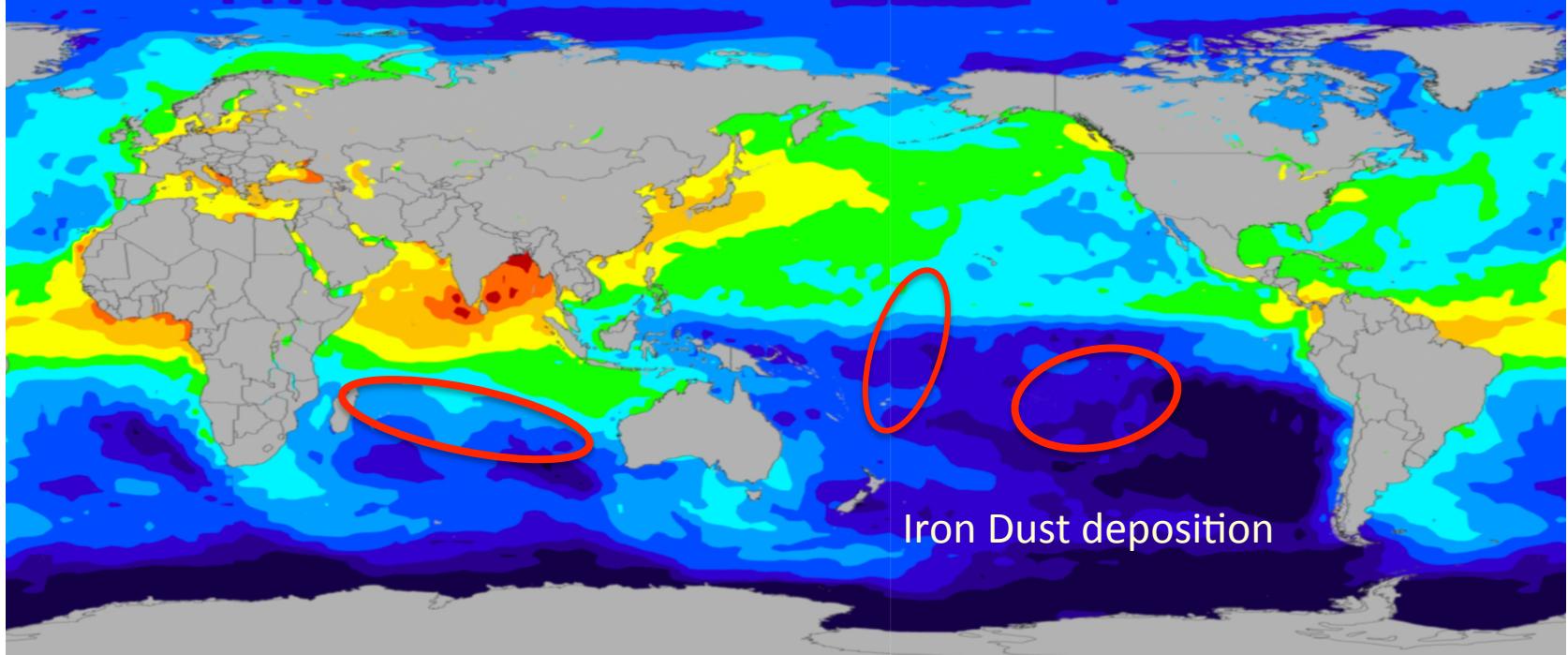
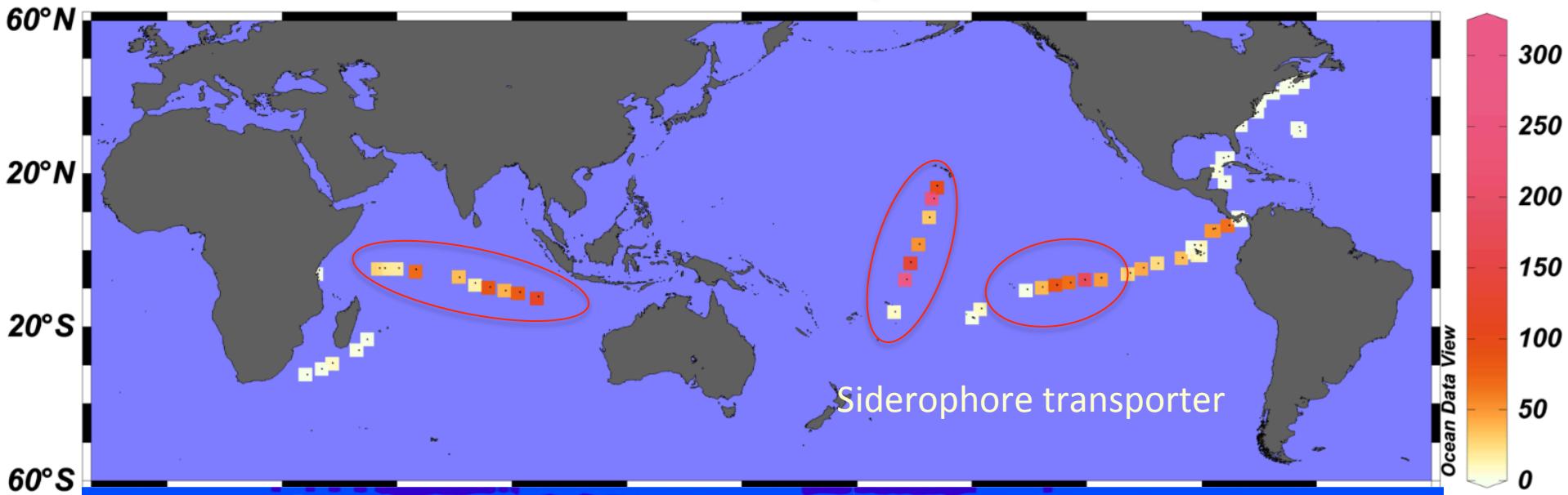


Genes are expressed under conditions of iron starvation in cultures

# Global distributions through GOS metagenomics



# Makes sense, so far...

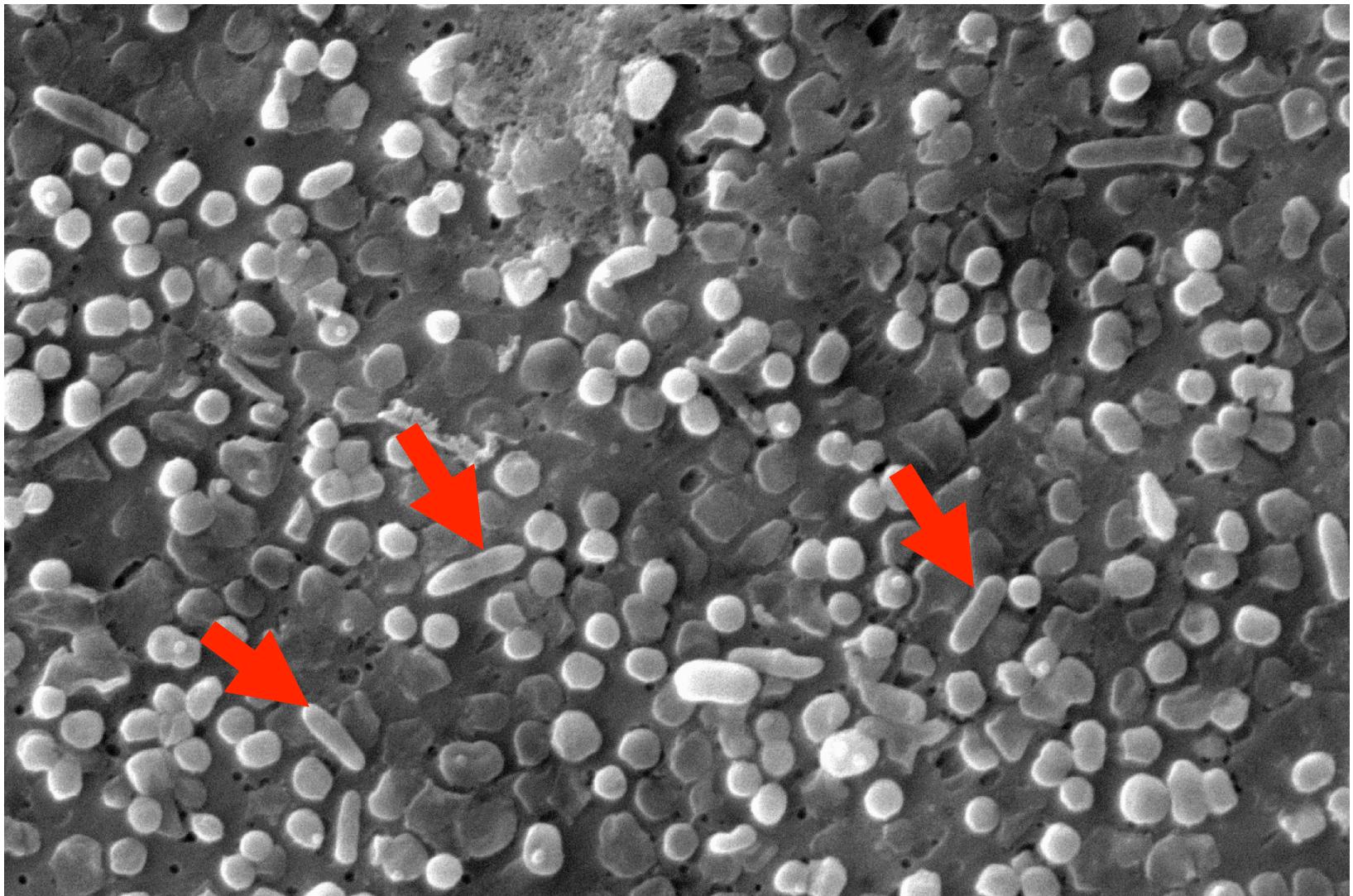


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# *Prochlorococcus* loves its heterotrophs

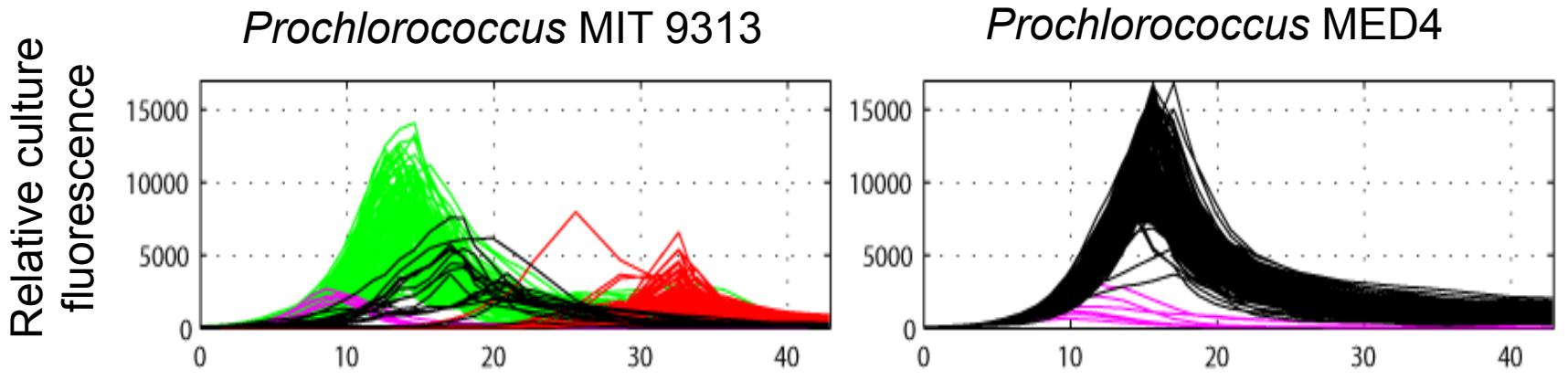


1  $\mu\text{m}$

Photo by Anne Thompson

# *...but not ALL heterotrophs*

- No effect of co-culture
- enhanced
- inhibited



*...and different strains respond differently to the same suite of heterotrophs  
(a lifetime of PhD theses!)*



# The Black Queen Hypothesis

Jeffrey Morris, Richard Lenski and Erik Zinser

OPINION/HYPOTHESIS



## The Black Queen Hypothesis: Evolution of Dependencies through Adaptive Gene Loss

J. Jeffrey Morris,<sup>a,b</sup> Richard E. Lenski,<sup>a,b</sup> and Erik R. Zinser<sup>c</sup>

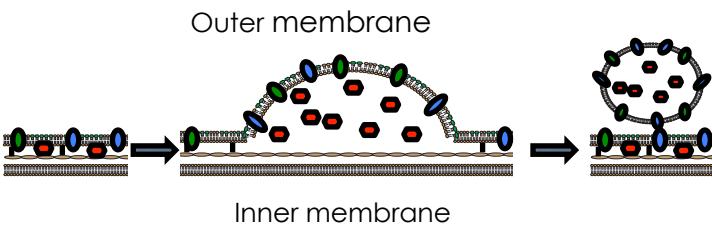
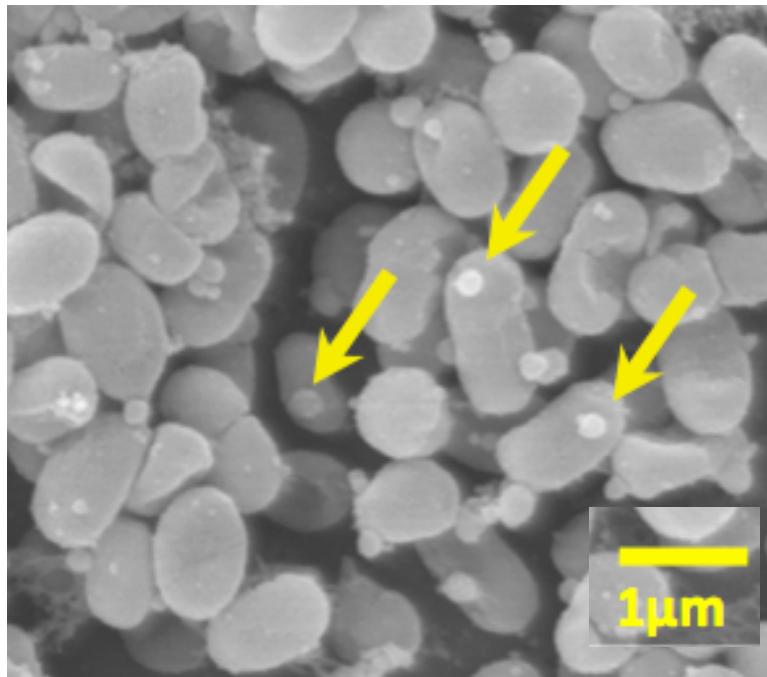
Michigan State University, East Lansing, Michigan, USA<sup>a</sup>; BEACON Center for the Study of Evolution In Action, East Lansing, Michigan, USA<sup>b</sup>; and University of Tennessee, Knoxville, Tennessee, USA<sup>c</sup>

**Evolution generates “beneficiaries” of reduced genomic content  
[*Prochlorococcus*] dependent on leaky “helpers,” [heterotrophs that leak catalase-peroxidase] perhaps explaining the observed non-universality of phototrophy, stress resistance, and other cellular functions...**

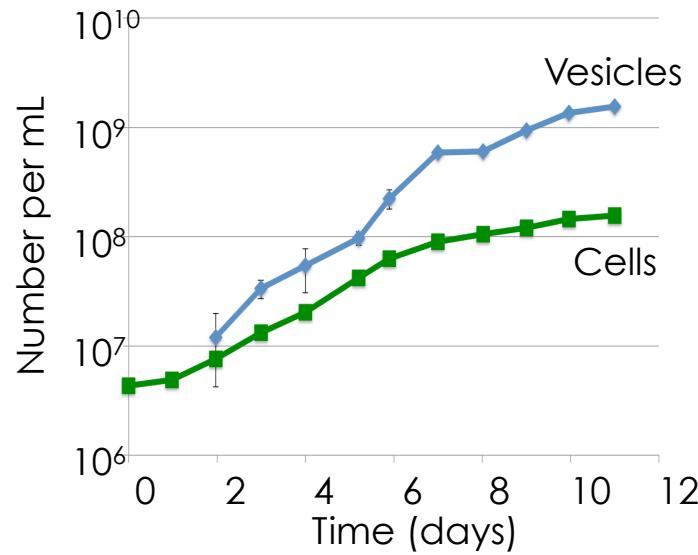
It's a metabolic marketplace...  
...a meta-metabolic web

J. Jeffrey Morris, Richard E. Lenski and Erik R. Zinser. 2012.  
mBio 3(2) doi:10.1128/mBio.00036-12.

# *Prochlorococcus* reveals another new ocean feature



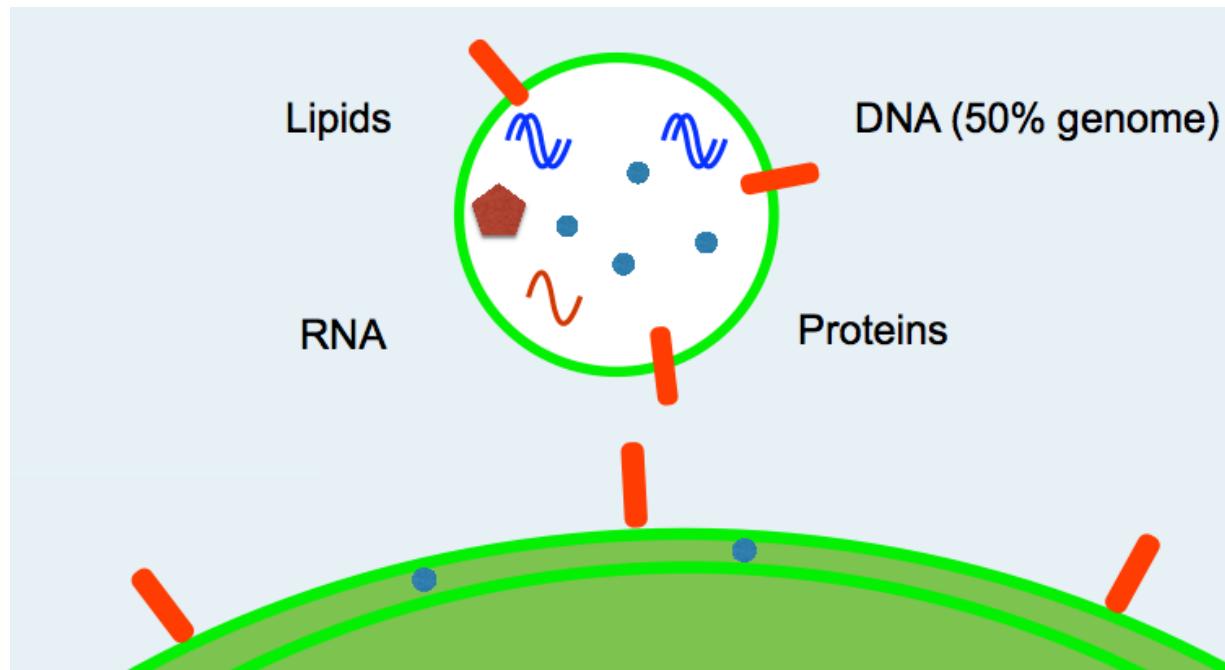
After Ellis and Kuehn (2010) MMBR, Schertzer and Whiteley (2012) mBio



- 2-5 vesicles per cell per generation
- Stable for weeks



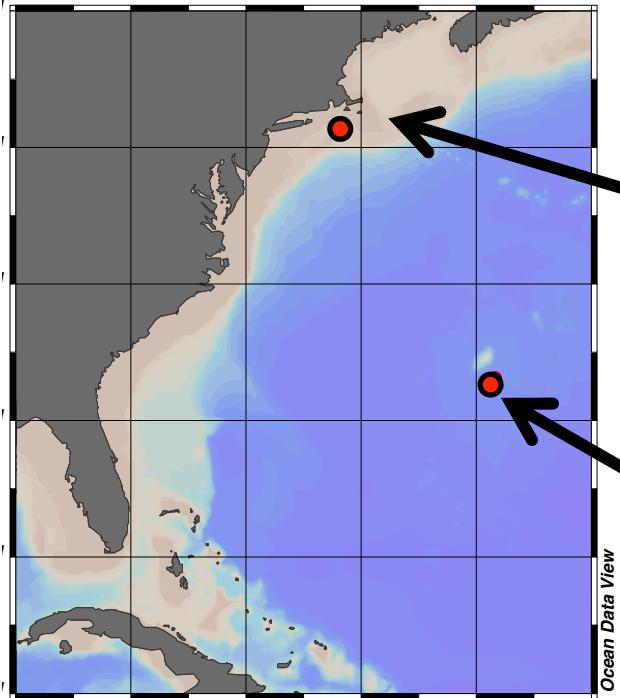
- Global production  $\sim 10^{27}$ - $10^{28}$  vesicles/day (0.1 – 1 megatonnes carbon)
- Vesicles contain interesting cargo



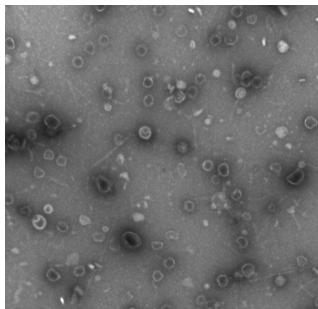


# Many ocean microbes produce vesicles

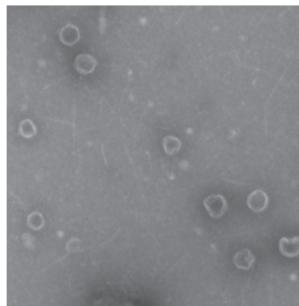
- A new dimension of ocean biogeochemistry



Vesicles abound  
 $10^5 - 10^6 \text{ mL}^{-1}$



Coastal



Open ocean

Vesicles contain DNA from...

34 phyla across all prokaryotes

# What is the function of vesicle release?

## Why should *Prochlorococcus* release valuable resources?

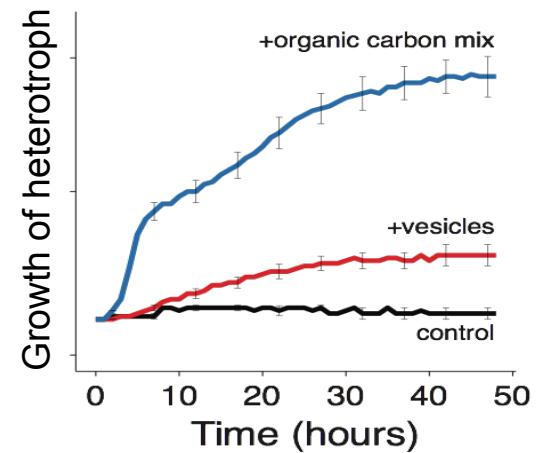
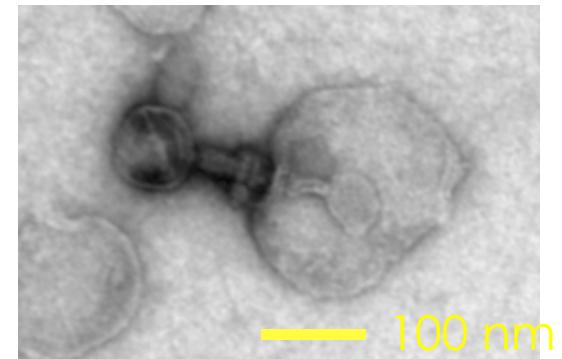
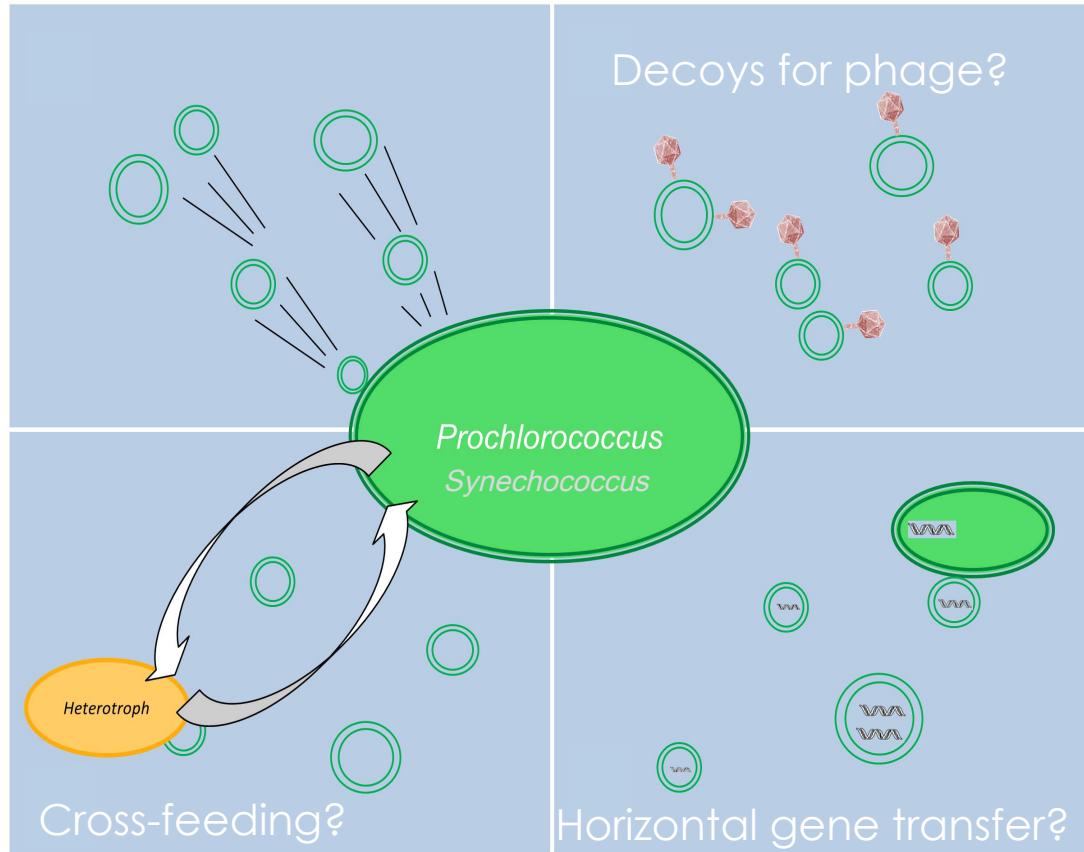


Image from Dave Scanlan

# Overview

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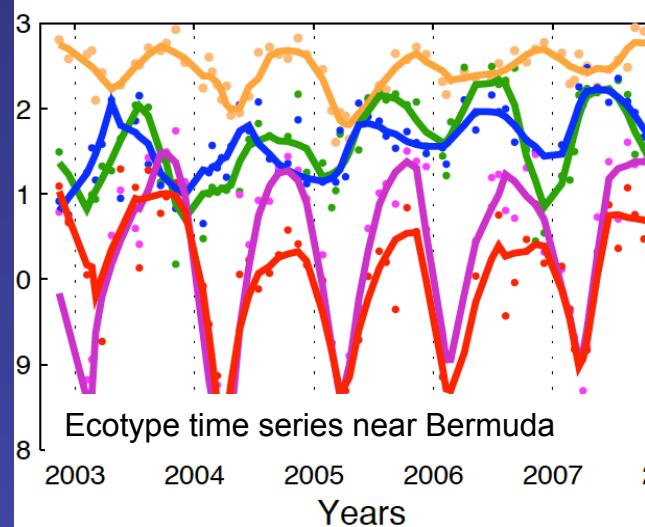
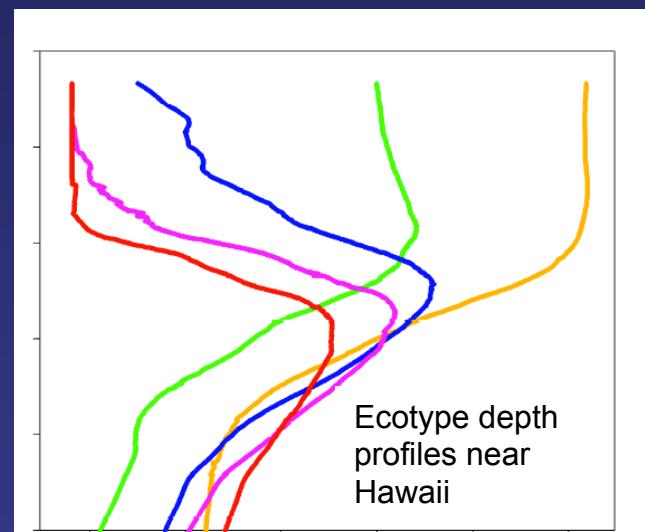
- ◆ History
- ◆ The Cell
- ◆ Niche Dimensions of *Prochlorococcus*
  - Light and Temperature
  - Genomics and Niche Dimensions:
    - Phosphorus
    - Nitrogen
    - Iron
- ◆ The Community
- ◆ Marine Vesicles!
- ◆ Integrative Systems Biology

# The micro-scale complexity is humbling...

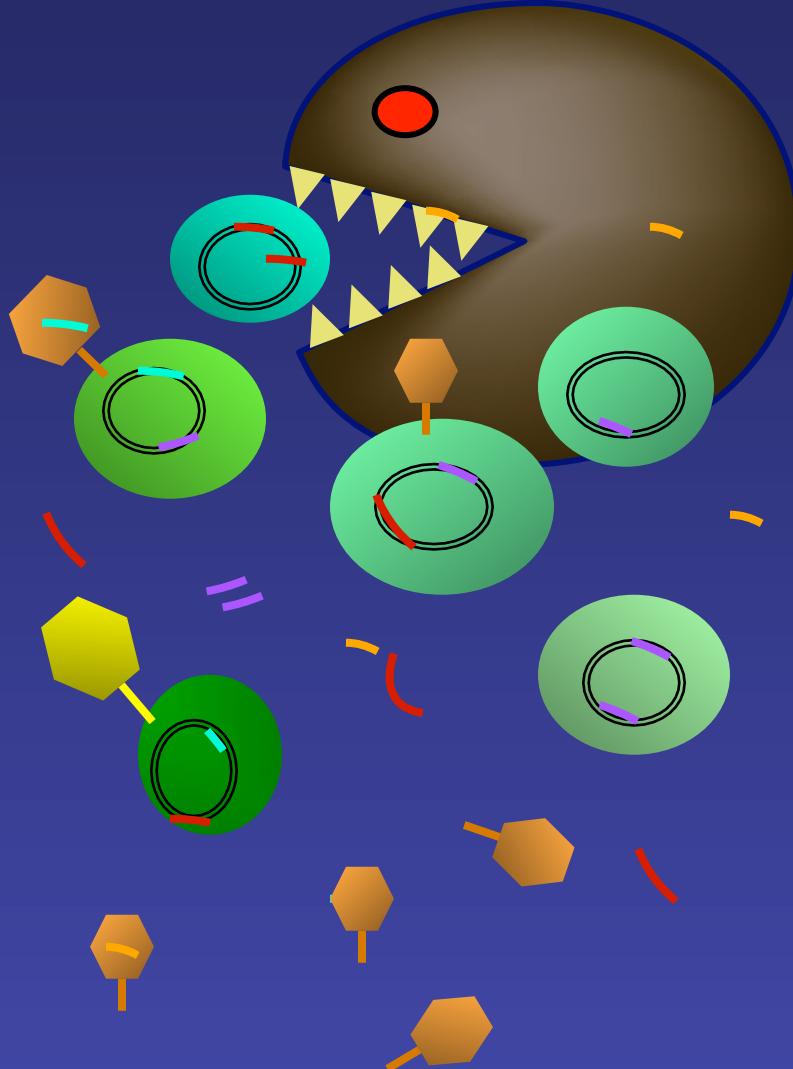


Have come to view the system as  
A loose network of  
'dissolved information'  
temporarily housed  
in microbes and viruses

...the stability of the emergent patterns is awe inspiring

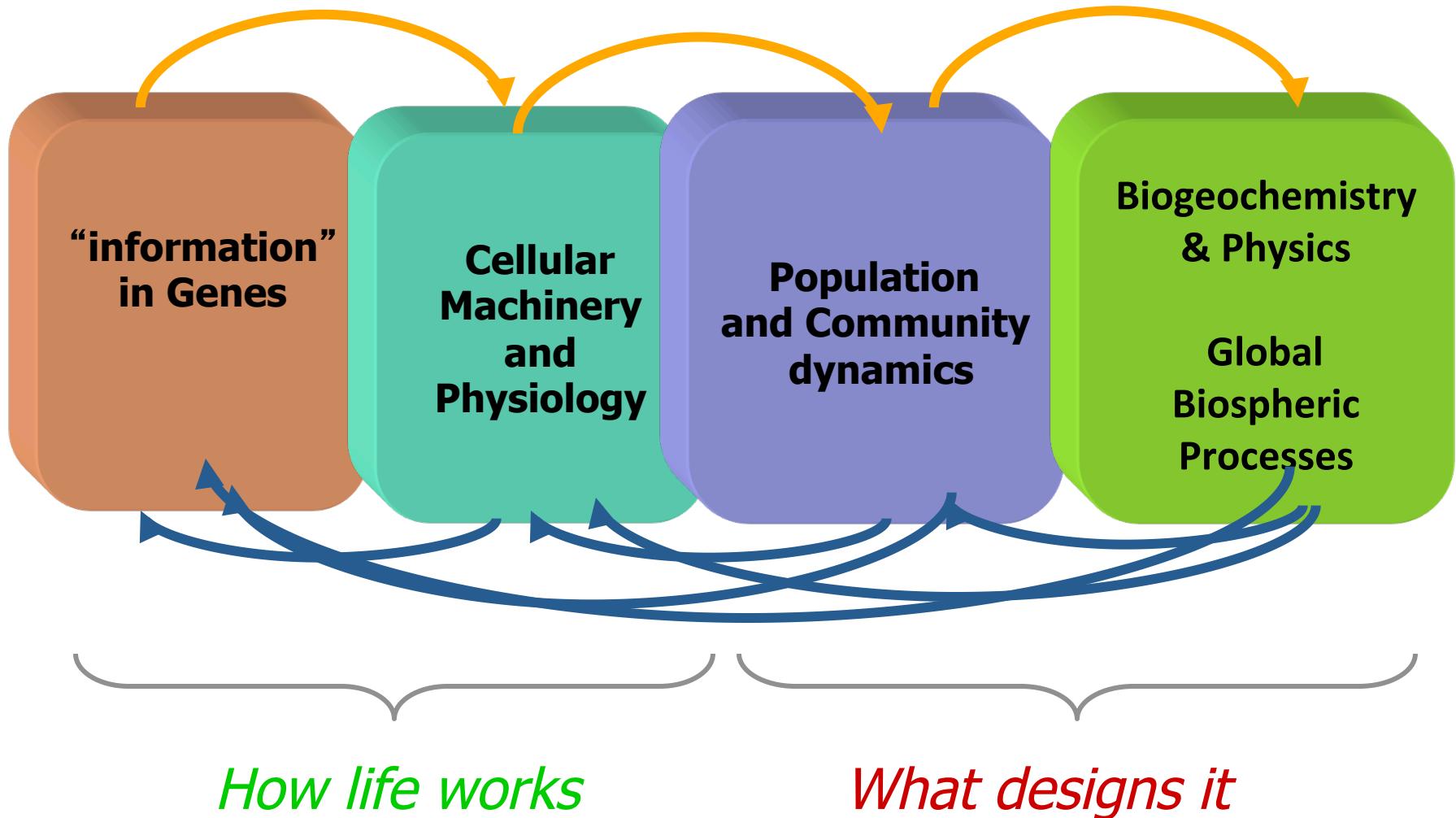


# It's a co-evolved, self-organizing, COMPLEX SYSTEM...



What are the  
assembly rules?

# Toward a “New Biology” Integrative Systems Biology



*"Our task now is to resynthesize biology;*

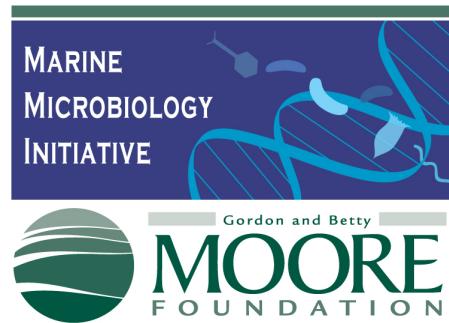
*...put the organism back into its environment;*

*...connect it again to its evolutionary past;*

*...and let us feel that complex flow that is organism, evolution, and environment united."*

**– Carl Woese, 2004**

# Many thanks to...



Rob Olson

Lisa Moore  
Gabrielle Rocap

Allison Coe  
Debbie Lindell

Erik Zinser  
Zackary Johnson

Rex Malmstrom  
Sebastien Rodrigue

Jason Bragg  
Maureen Coleman

Katherine Huang  
Matt Sullivan

Greg Kettler  
Anne Thompson

Jessie Thompson  
Katya Fros-Moniz

Andres Cubillos-Ruiz  
Nadav Kashtan

Steve Biller

Adam Martiny  
Kun Zhang  
Paul Berube  
Jake Waldbauer  
Luke Thompson  
Qinglu Zeng  
Libusha Kelly  
Peter Weigle

Mick Follows  
Mak Saito  
Eric Webb  
Eric Alm  
Ed DeLong  
George Church  
Bruce Birren  
Matt Henn

And many more!

