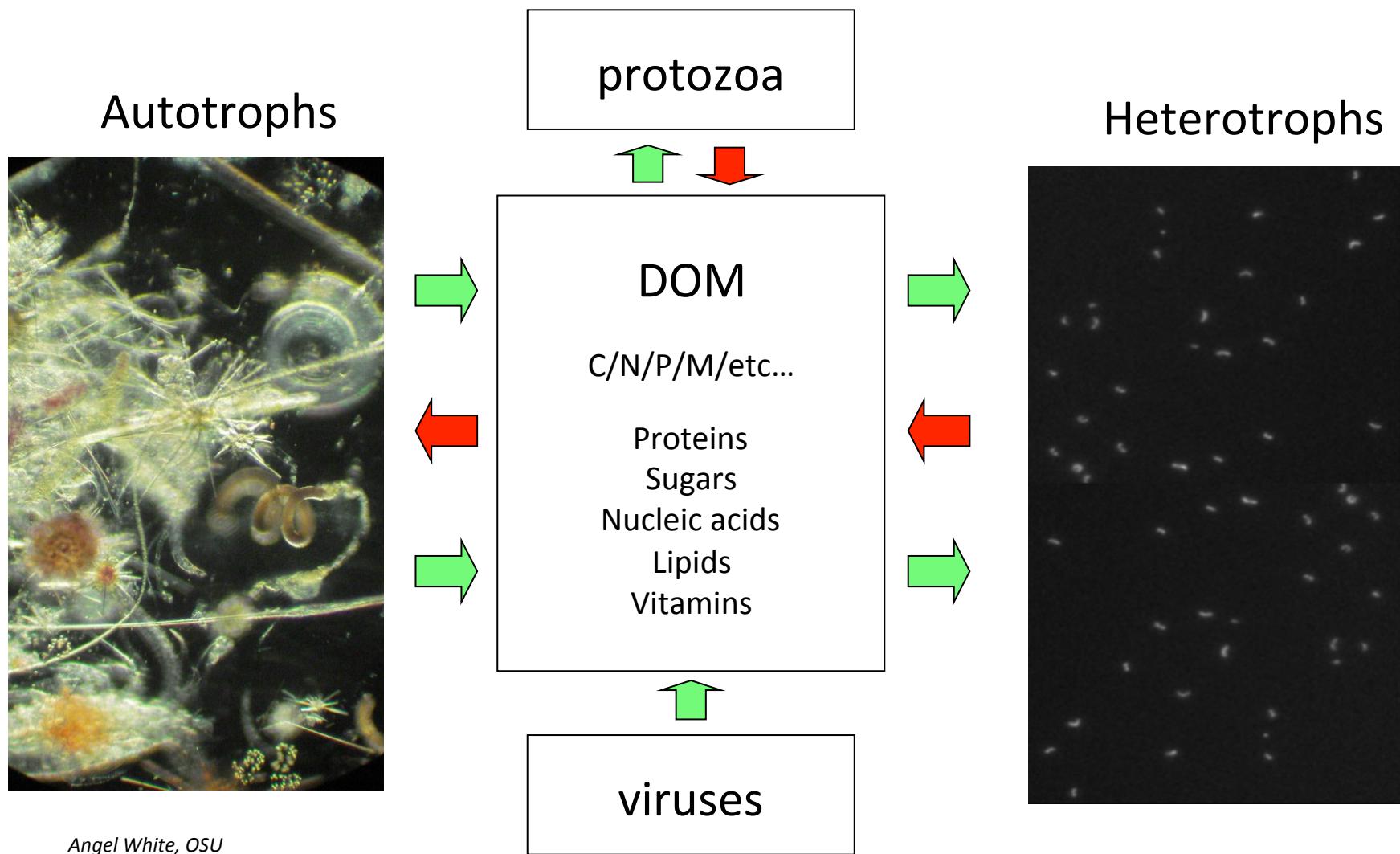
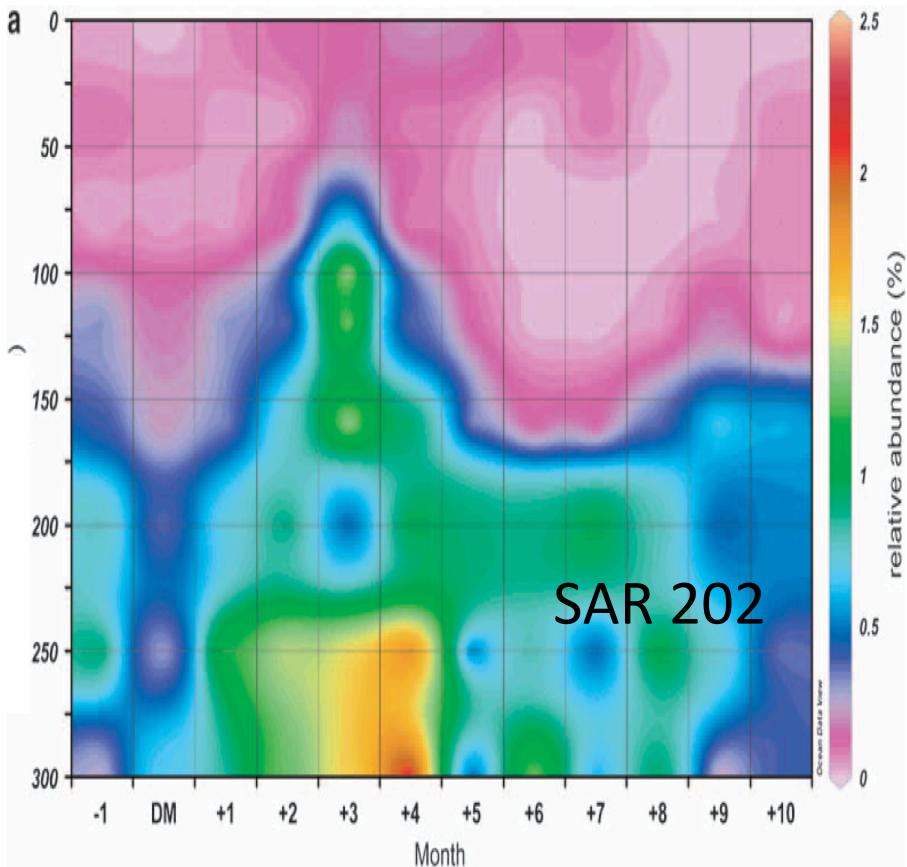
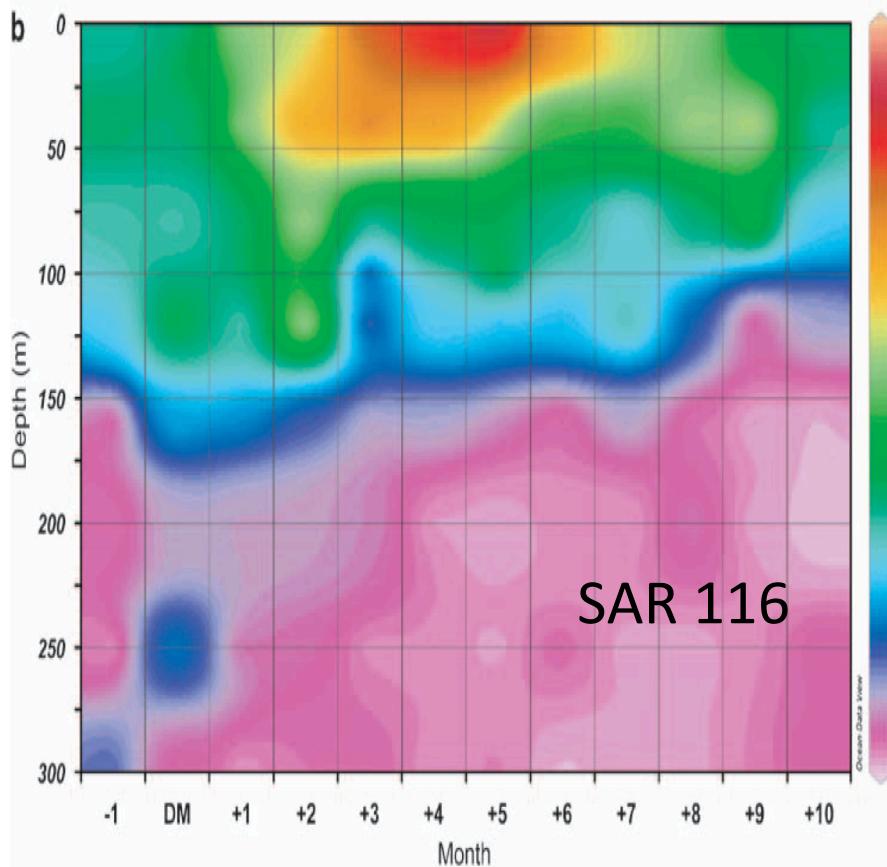


# Coupling microbial diversity and activity with dissolved organic matter production and cycling within the microbial loop.



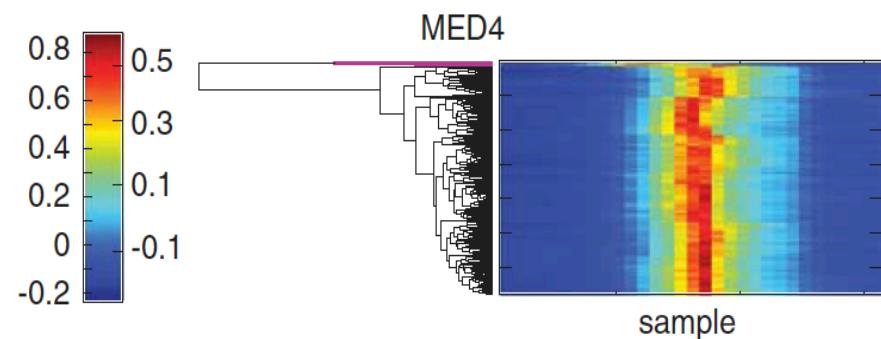
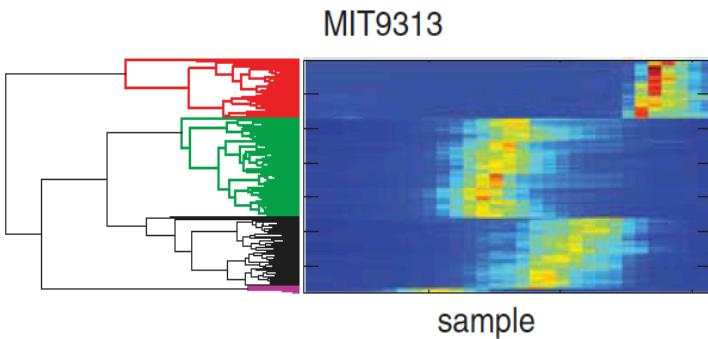
# How do we explain the seasonal and vertical structure of heterotrophic microbial communities in the ocean?



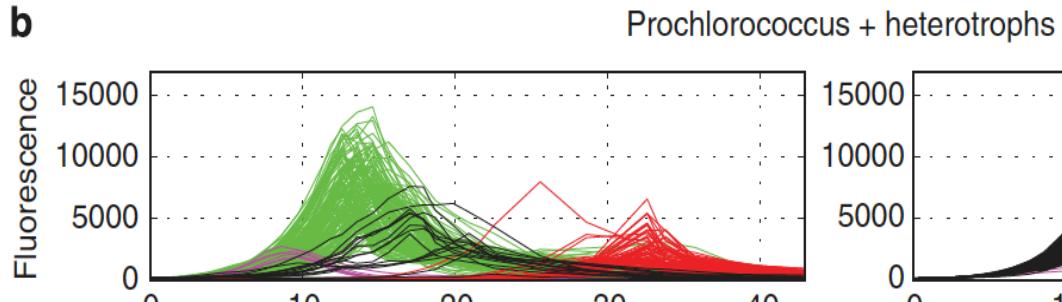
*“Interactions between bacterioplankton cells and natural DOM pool are difficult to measure... Although it has not yet been possible to measure heterotrophic interactions between microbial populations and DOM at a comprehensive molecular level, described patterns of DOM concentration and reactivity at the surface layer suggest they are factors that influence the distributions of SAR 11 populations.....” (Carlson et al., ISME 2009)*

# Do heterotrophs influence the growth of photoautotrophs?

a



b



Color legend  
for co-culture  
outcomes:

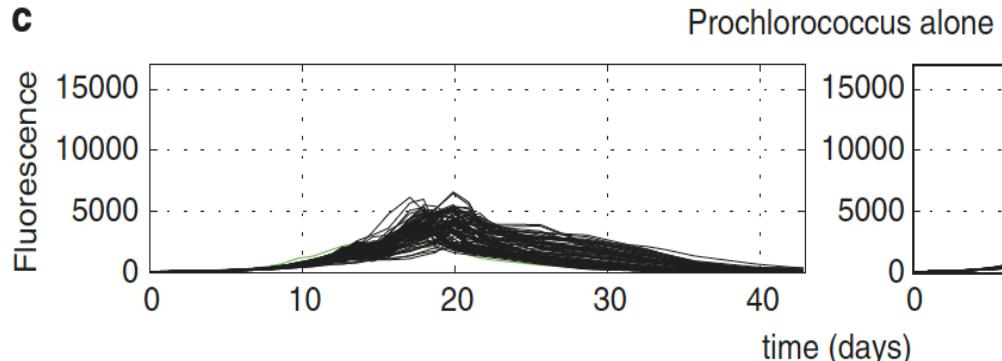
Early

Intermediate

Early arrested

Late

c



# Approaches

- 1) *Perturbation experiments: Add a specific (known) organic substrate and measure changes in microbial diversity, abundance, etc.*
- 2) *High throughput screening of natural organic matter fractions via pure cultures of heterotrophs and photoautotrophs.*
- 3) *Use genomic based reconstruction of metabolic pathways in pure cultures to determine substrate specificity – confirm via pure culture experiments.*
- 4) *Whole community metabolism experiments- add natural organic matter to seawater and use community transcriptomics to deduce which metabolisms are up-regulated in which organisms.*

# Isolation of Phytoplankton derived DOM

Triplicate axenic cultures  
(known biological source)



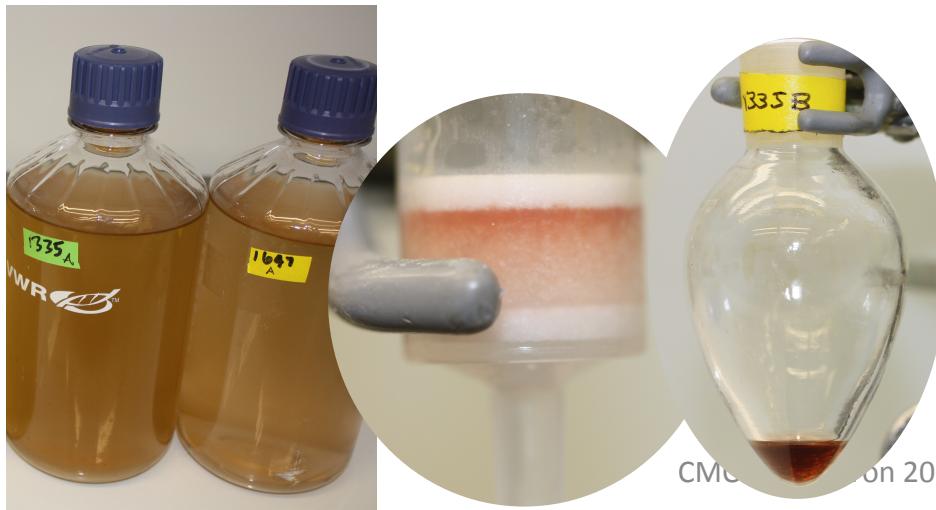
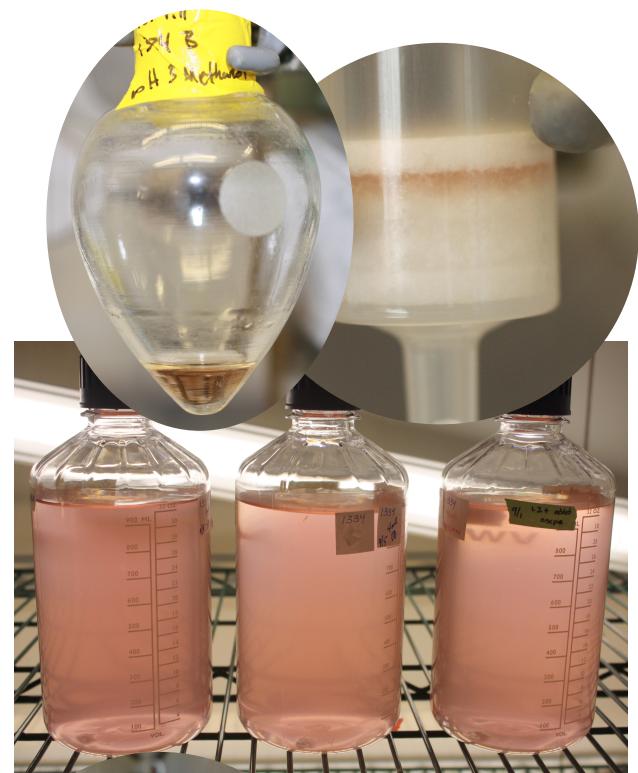
Biomass removal



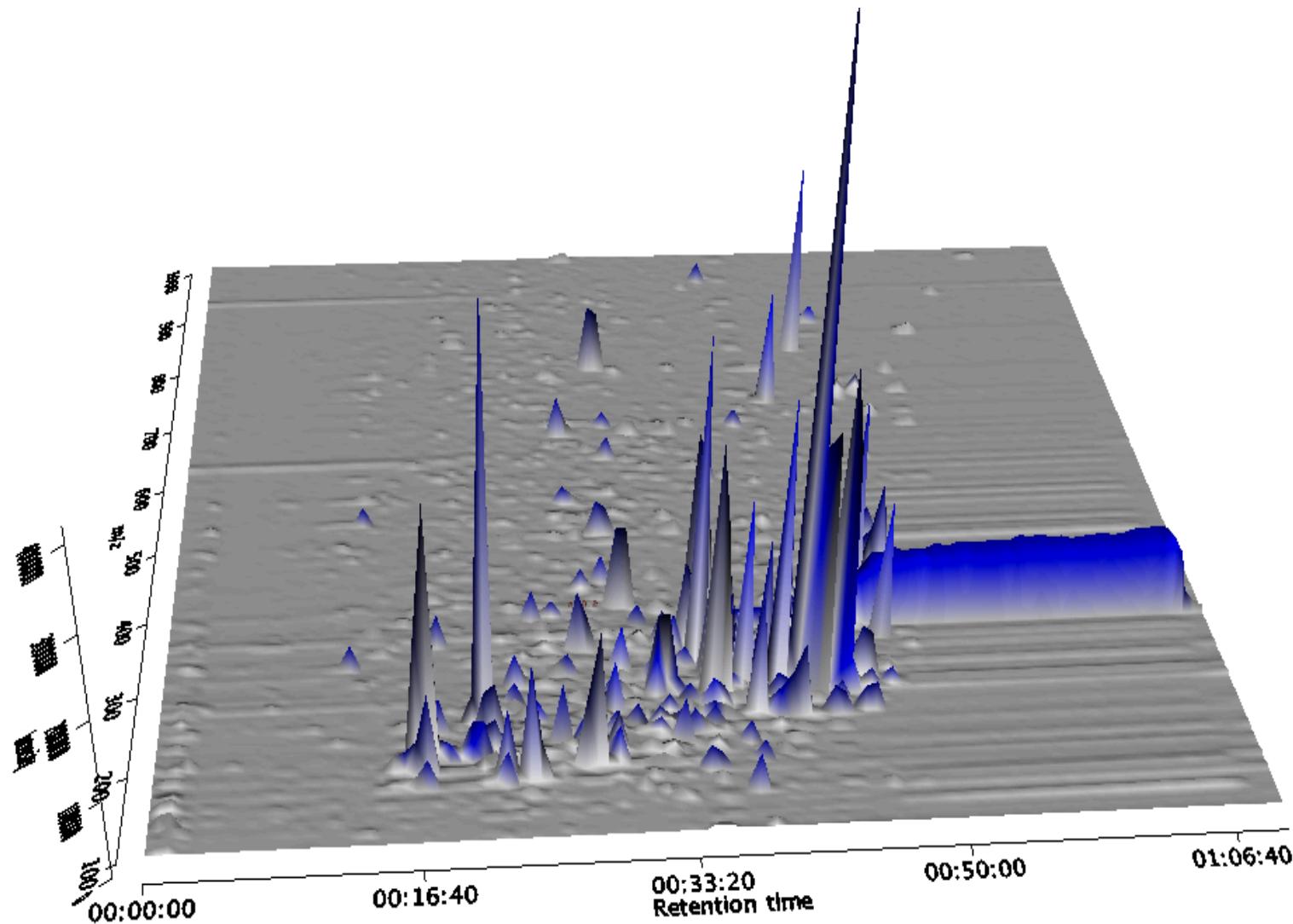
Solid-phase extraction



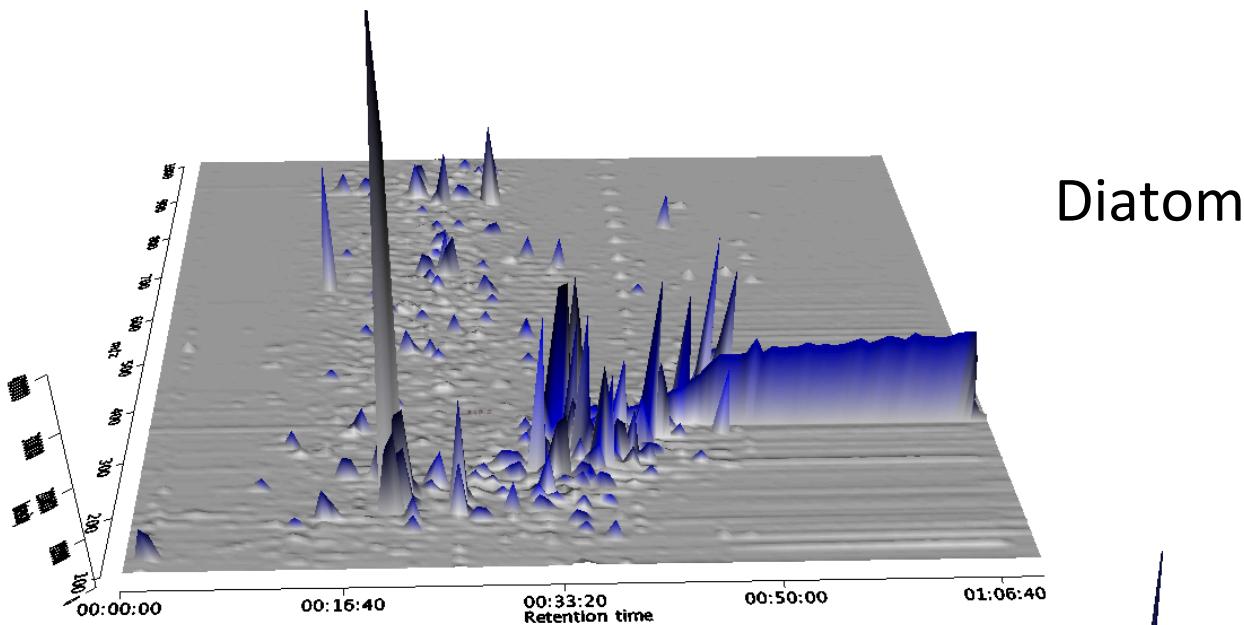
Labile material



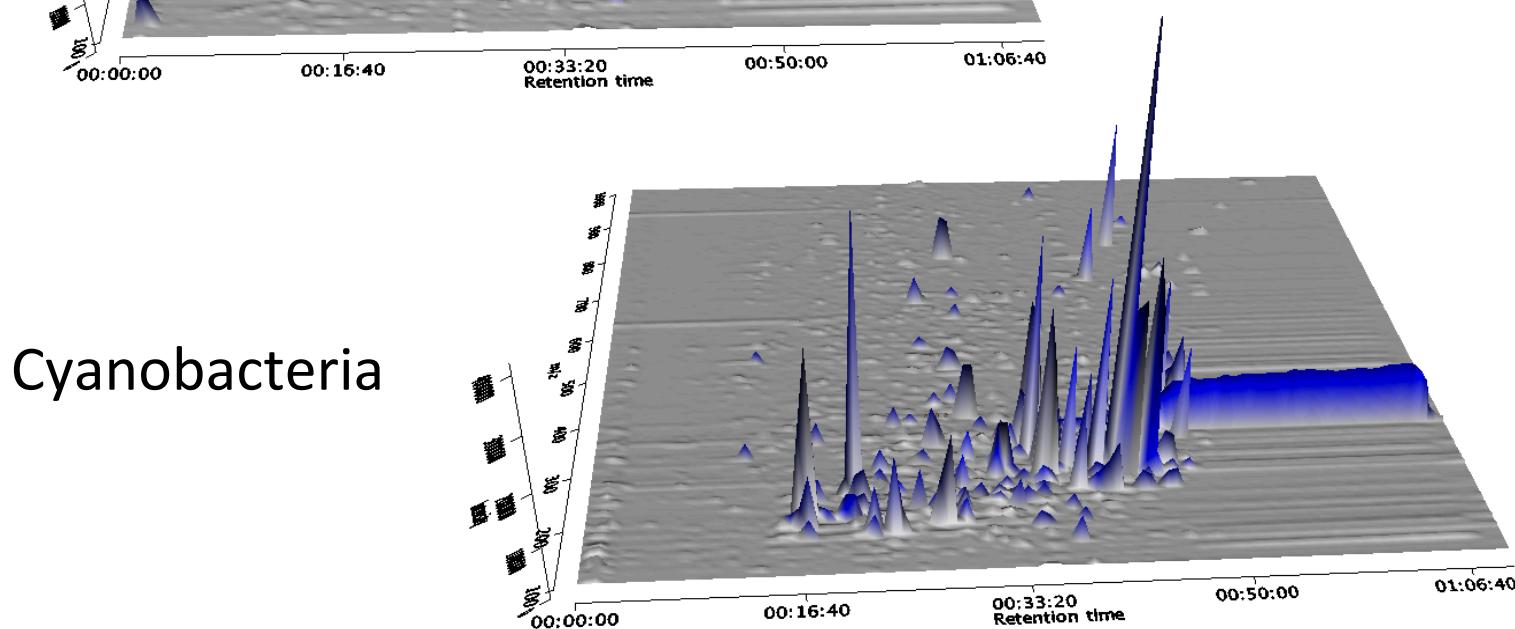
# Chemical Diversity by chromatography/mass spectrometry



# Chemical Diversity by chromatography/mass spectrometry

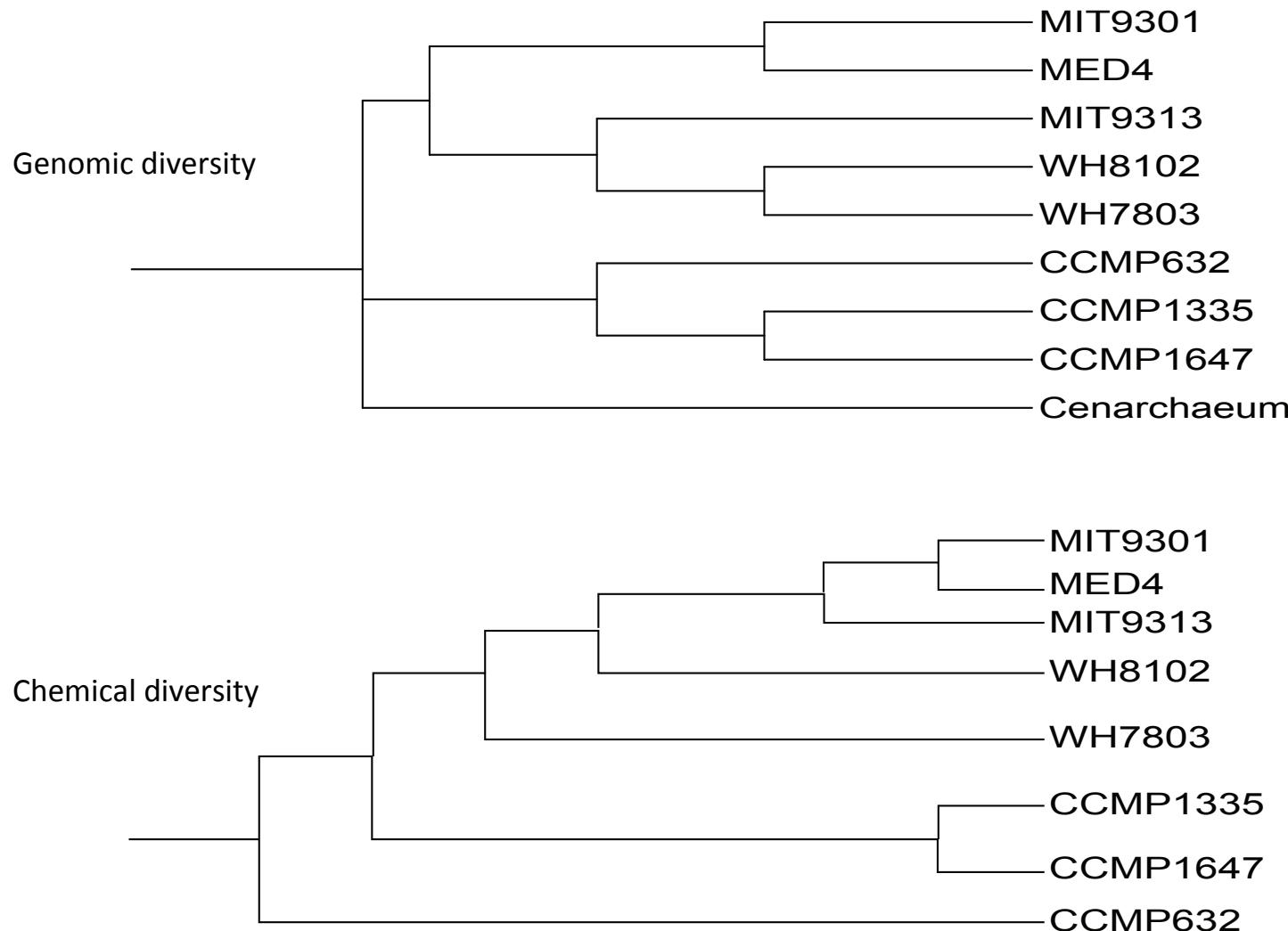


Diatom

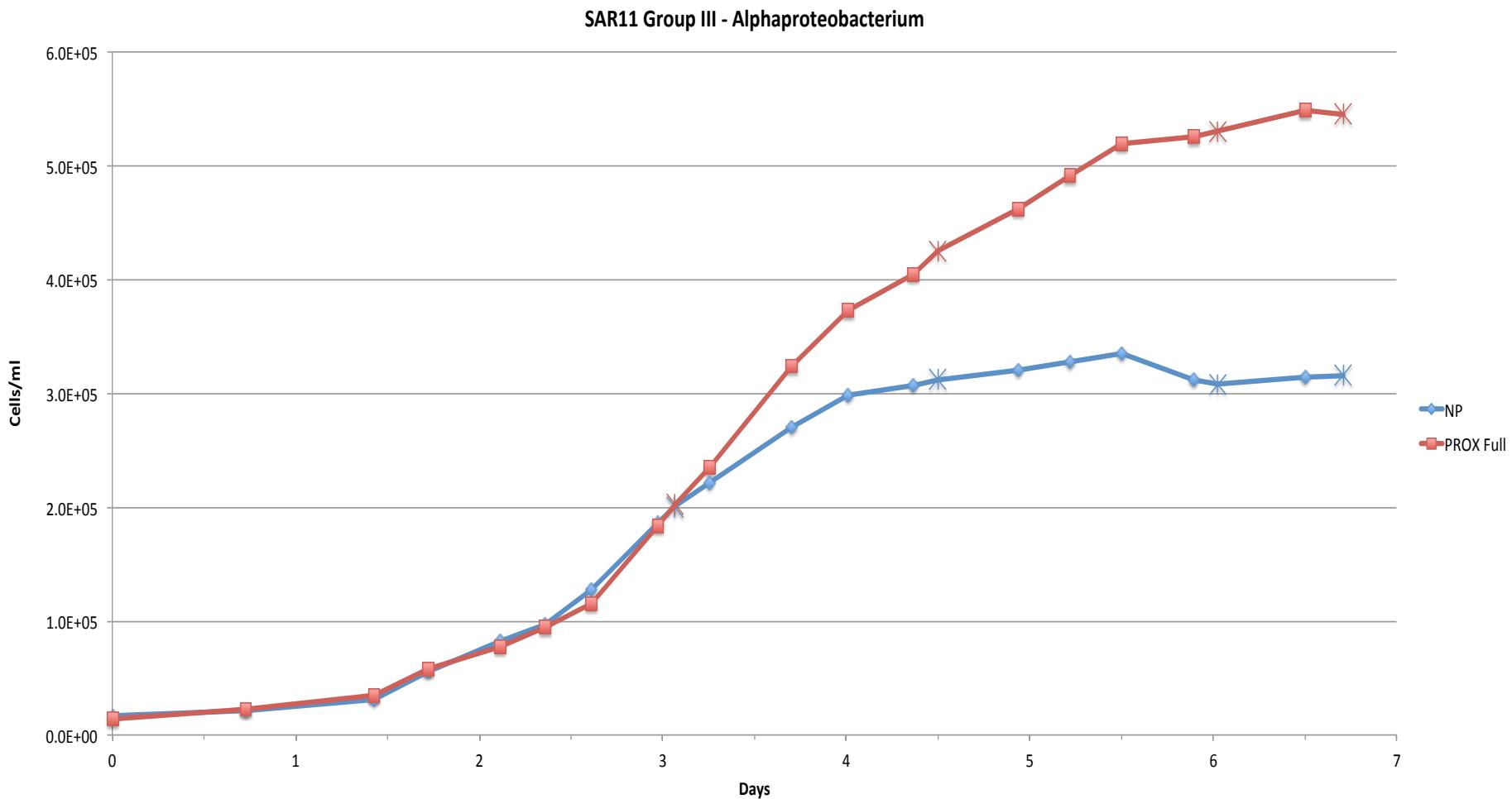


Cyanobacteria

# Chemical diversity tracks biological diversity

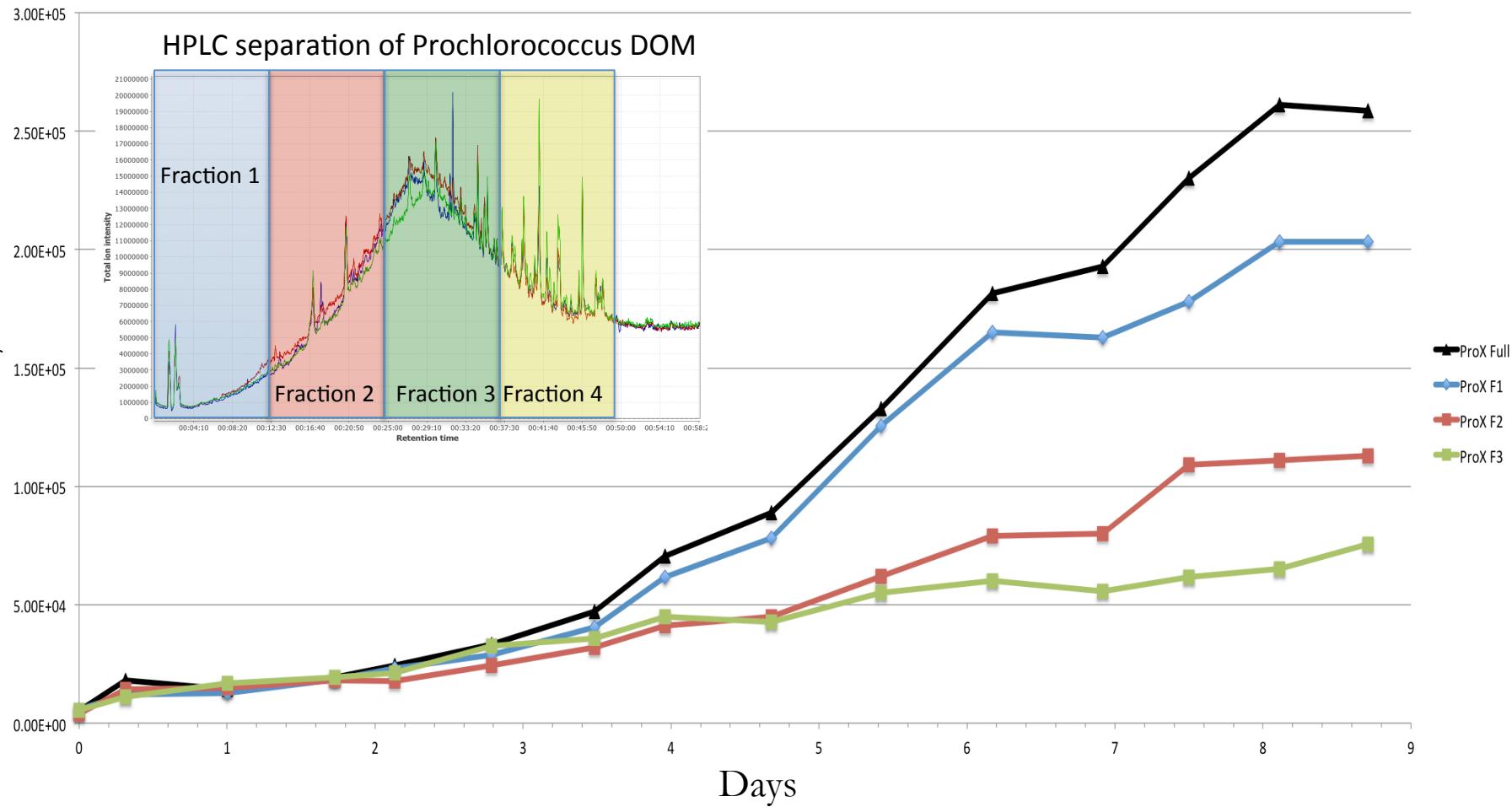


# Liability Screening

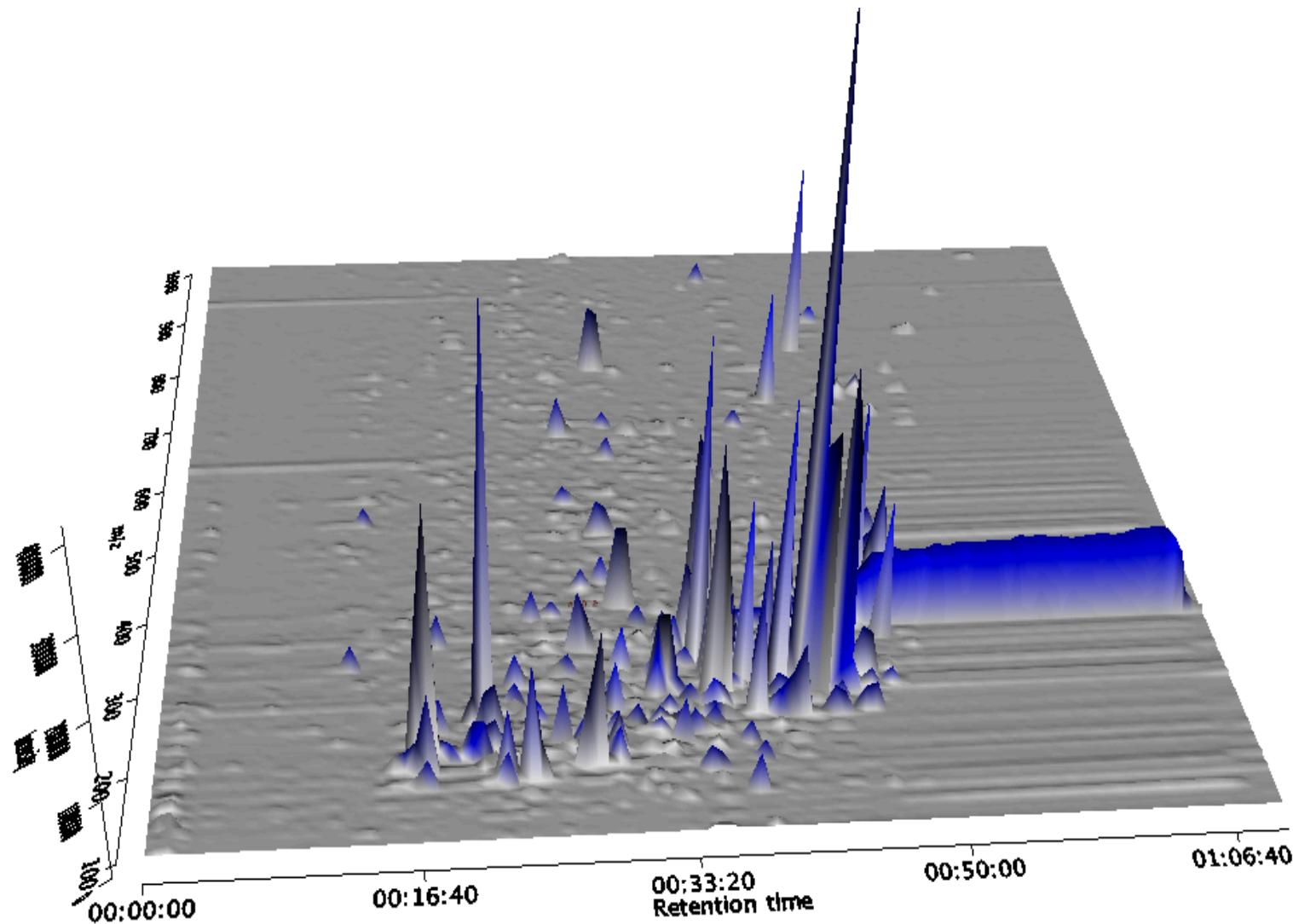


# Lability Screening

SAR11 Group III - Alphaproteobacterium

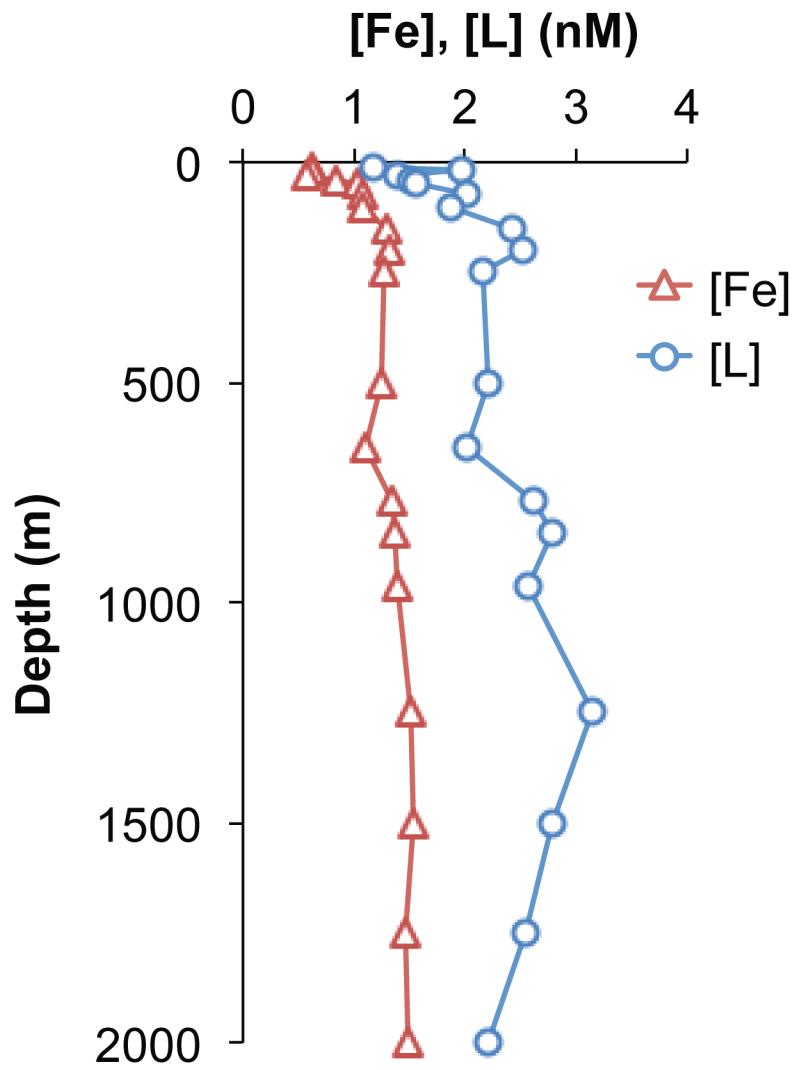


# Chemical Diversity by chromatography/mass spectrometry



# Marine Iron Ligands

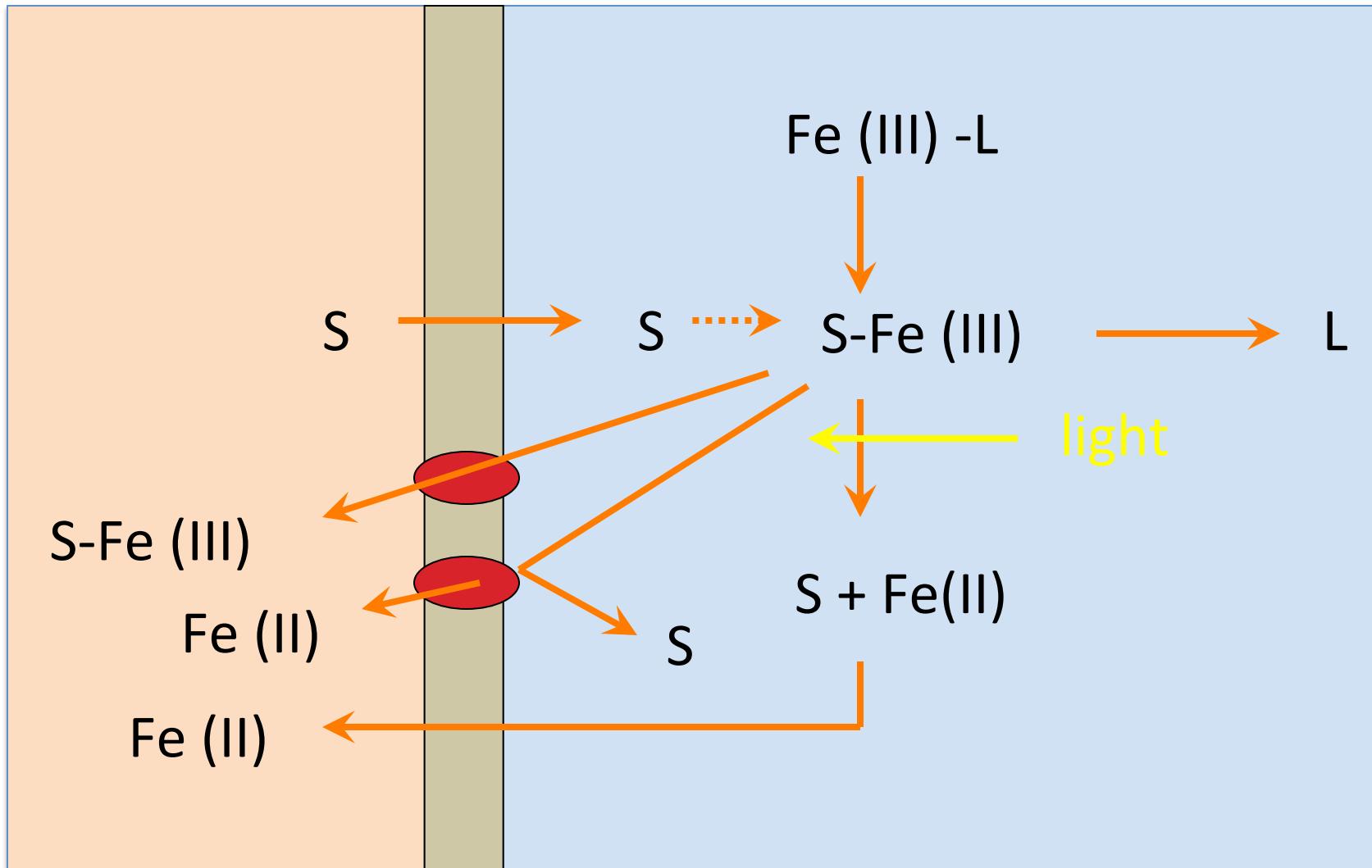
- Concentration of Fe ligands nearly always exceeds dissolved Fe.
- Conditional stability constant  $\text{LogK}_{\text{FeL}}$  between 10 -13.
- Suggests that >99% of dissolved Fe is complexed.



# Iron uptake mechanisms by bacterioplankton

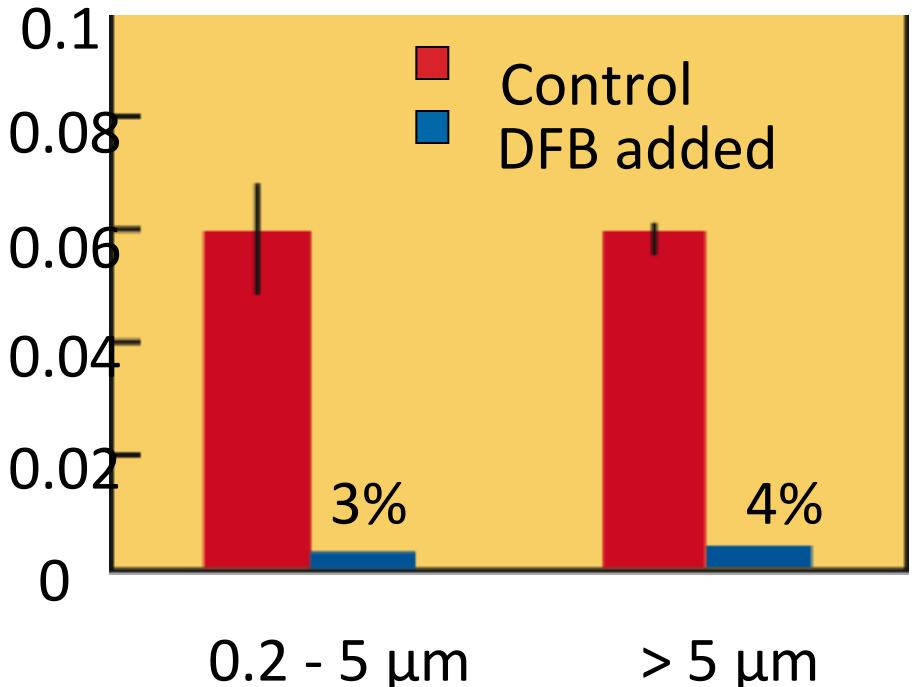
Internal

External

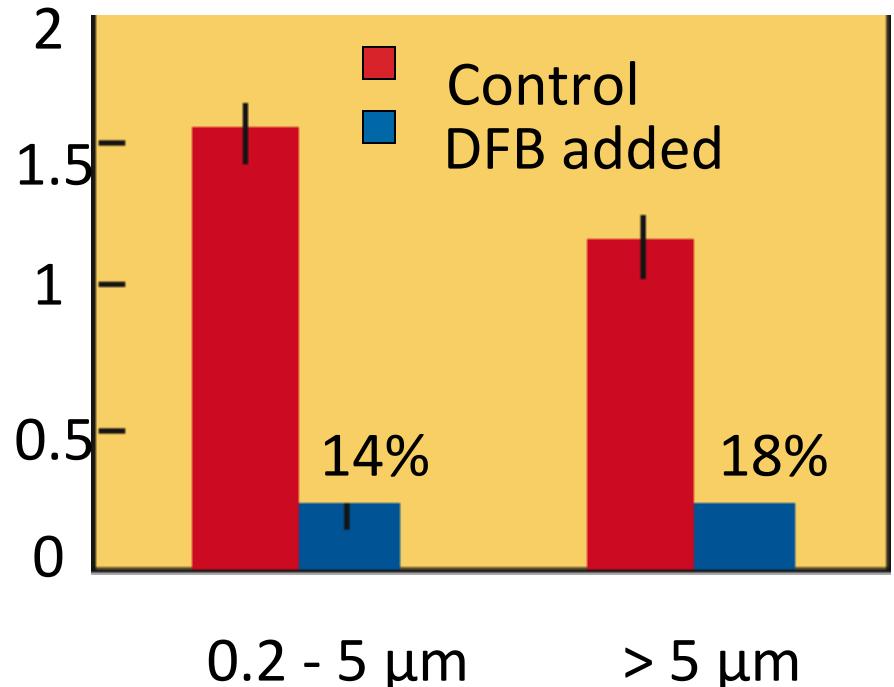


# Manipulation of Fe availability in seawater by the addition of a siderophore (DFB)

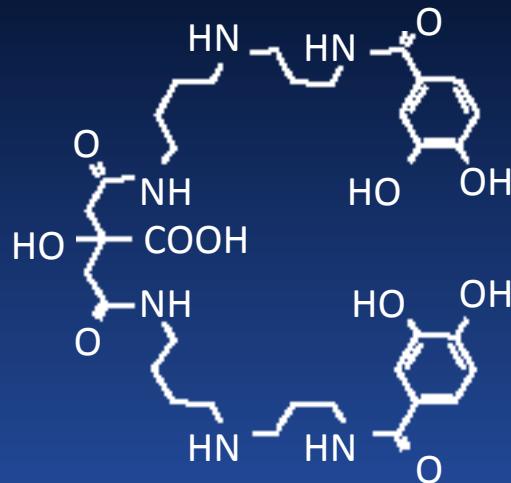
Fe uptake ( $\text{pmole ml}^{-1}$ )



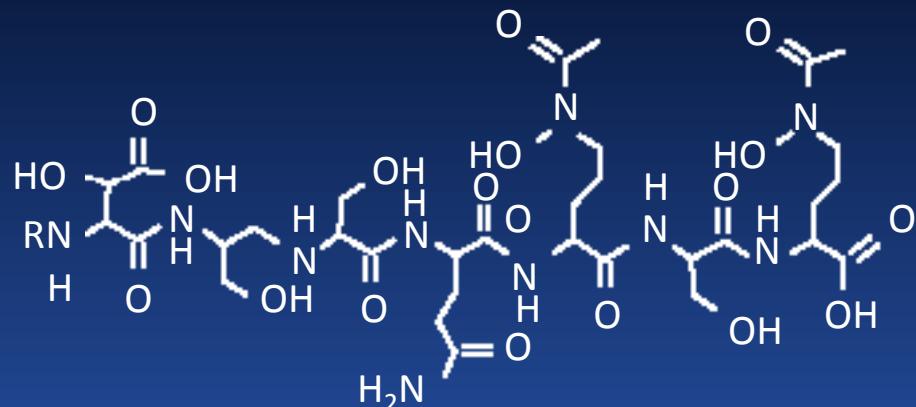
Carbon uptake ( $\text{mgC L}^{-1}$ )



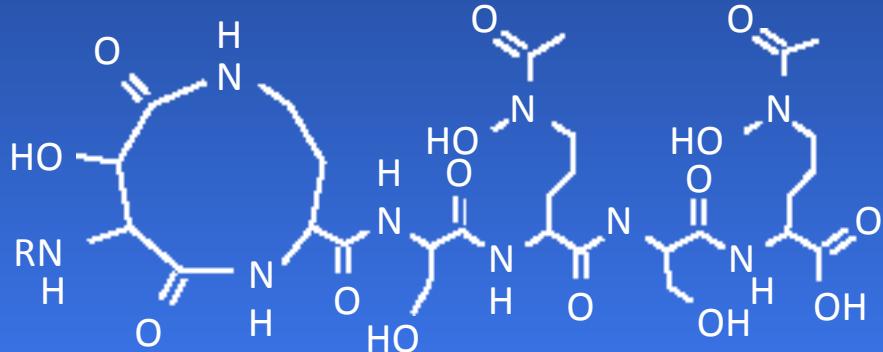
# Siderophores in Marine Bacteria



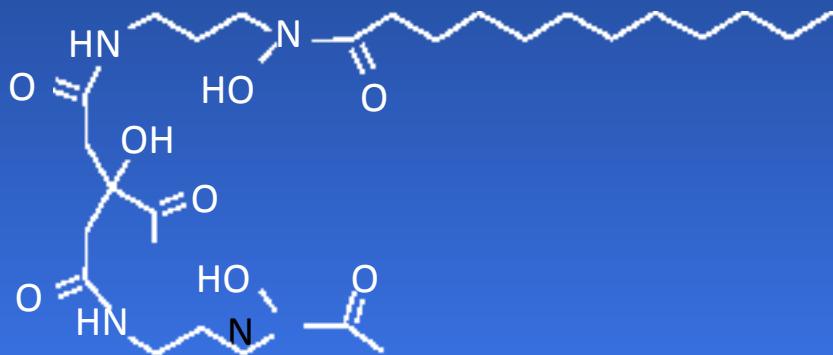
Petrobactin



Aquachelins

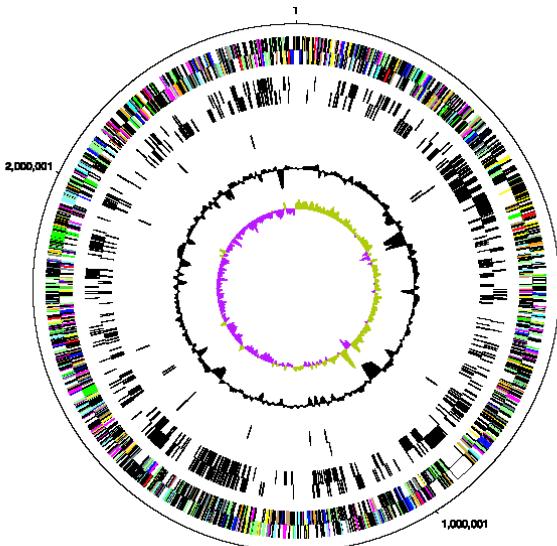


Marinobactins



Synechobactin

Then again there are good reasons to think that siderophores are not present as Fe-binding ligands in seawater...



Genomes of *Synechococcus* WH8102 and *Prochlorococcus* do not have detectable systems for siderophore production or uptake.

The energetics and efficiency of siderophore production and uptake in dilute media has been questioned in model calculations.

Only one marine cyanobacteria has been shown to make siderophores. NO siderophores have been isolated from seawater.

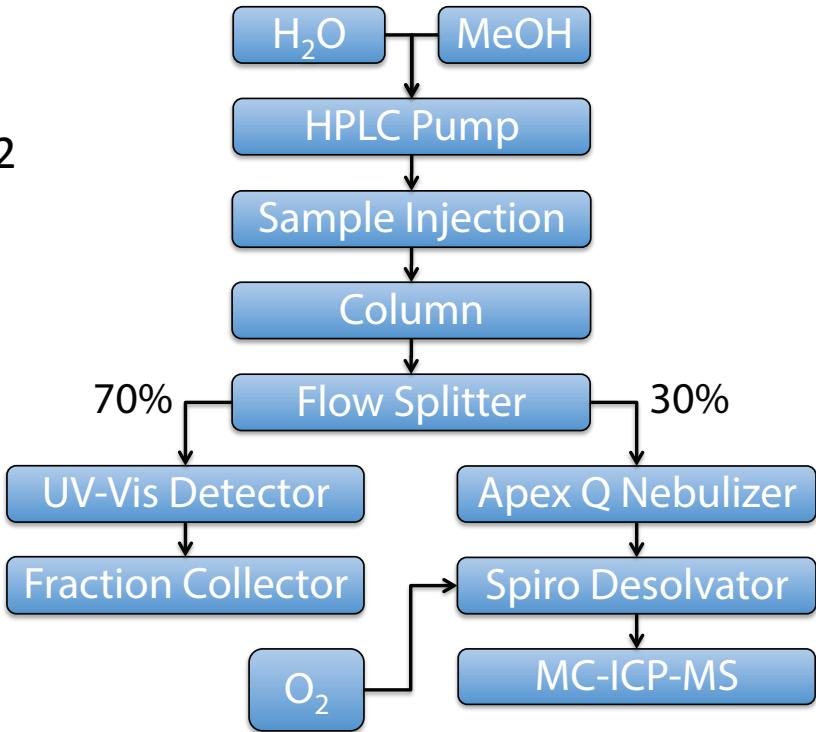
# Applications

## 1. Cyanobacteria cultures

- *Synechococcus* strain PCC 7002
- *Prochlorococcus*

## 2. Natural Seawater

- Subtropical Pacific



# Applications

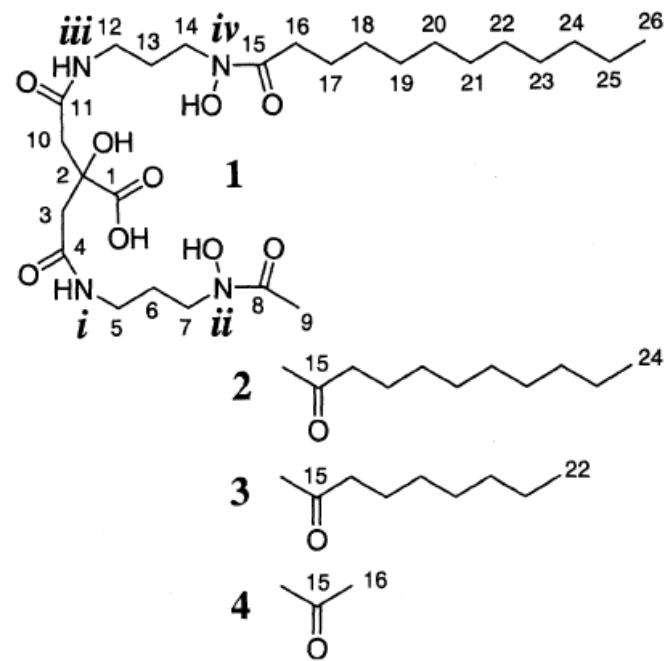
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## 1. Cyanobacteria cultures

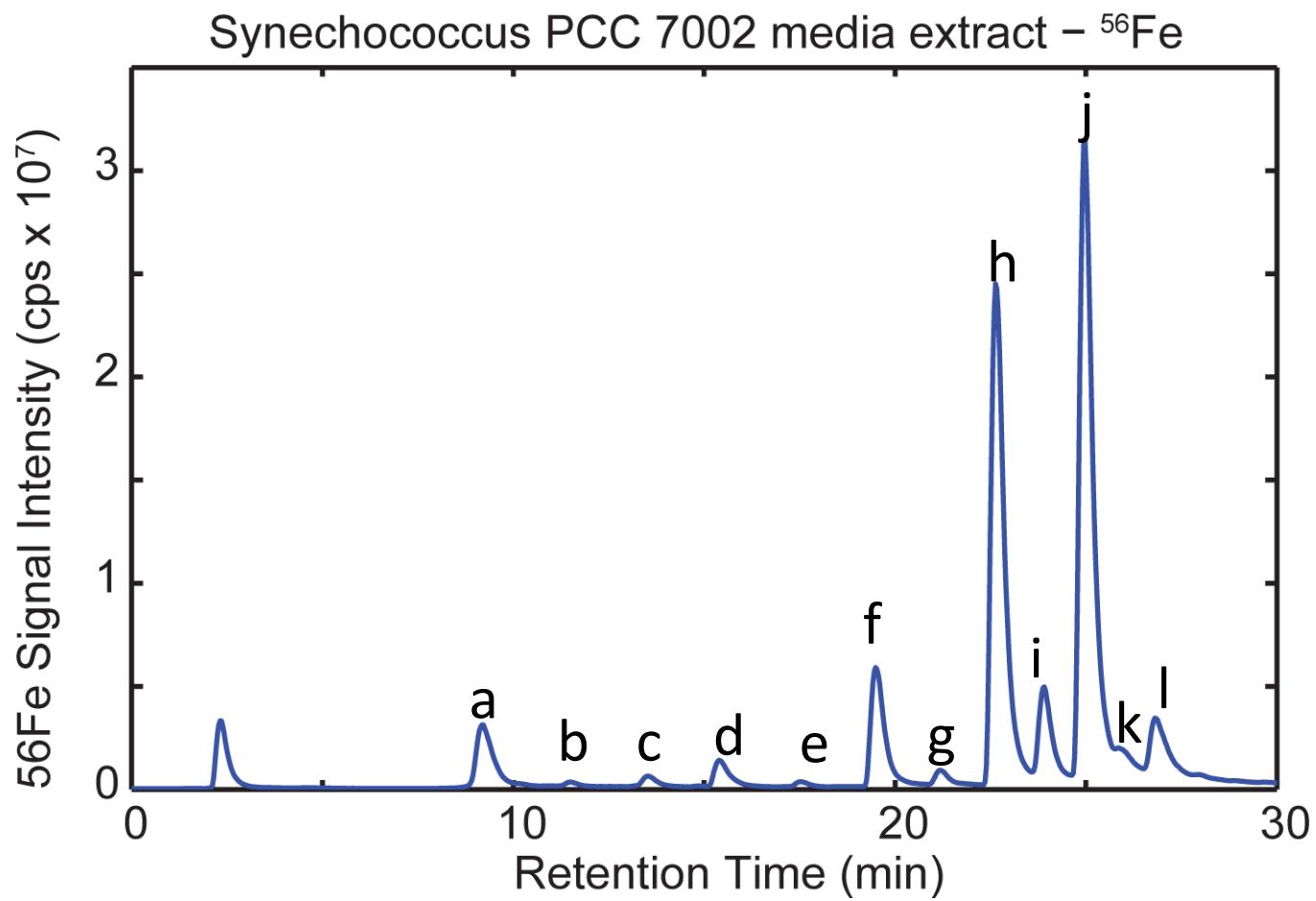
- ***Synechococcus* strain PCC 7002**
- *Prochlorococcus*

## 2. Natural Seawater

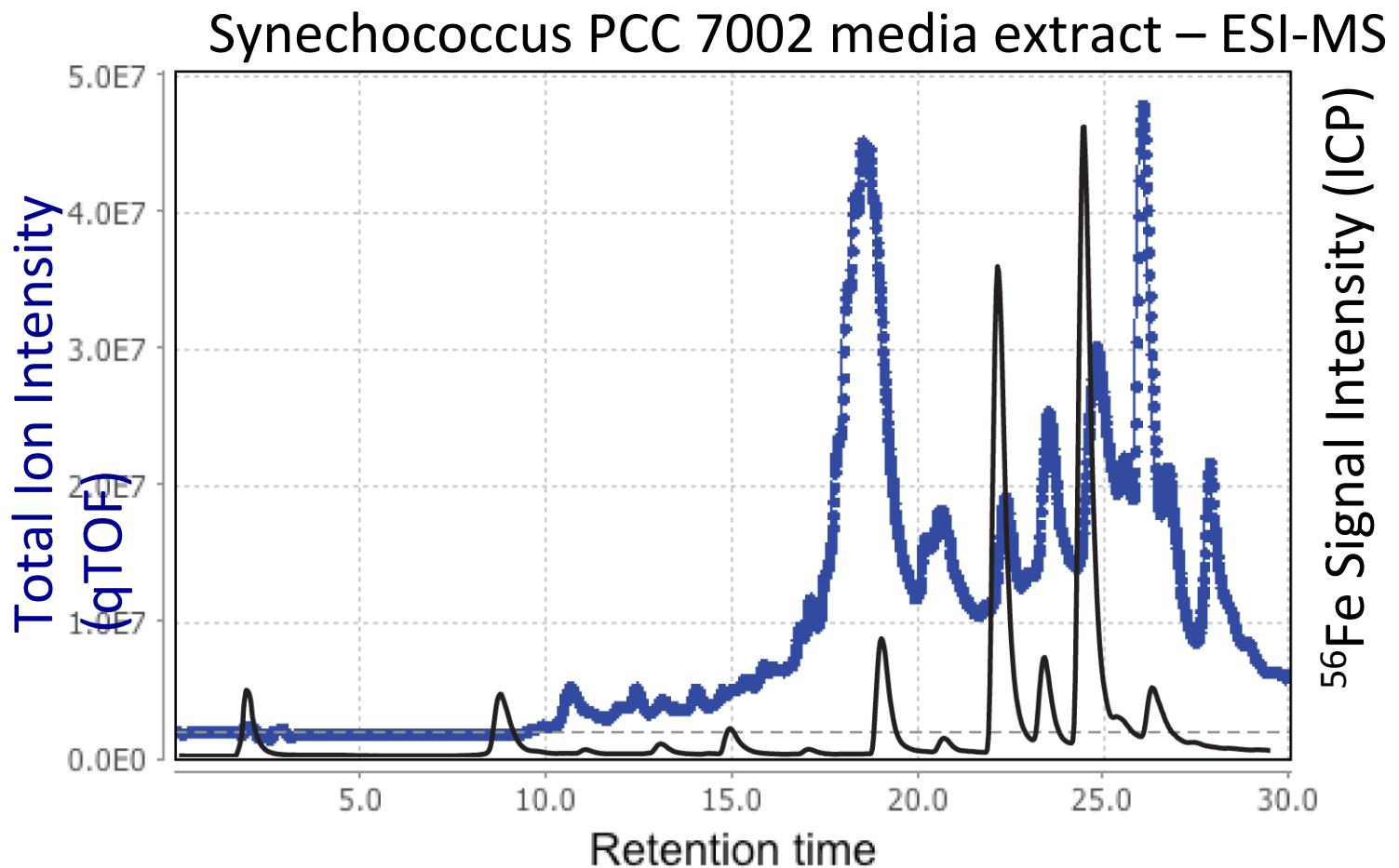
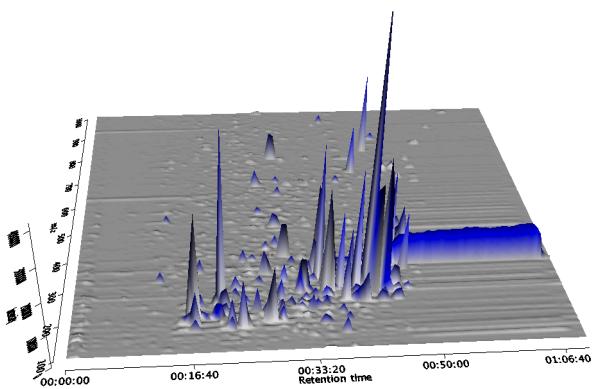
- Subtropical Pacific

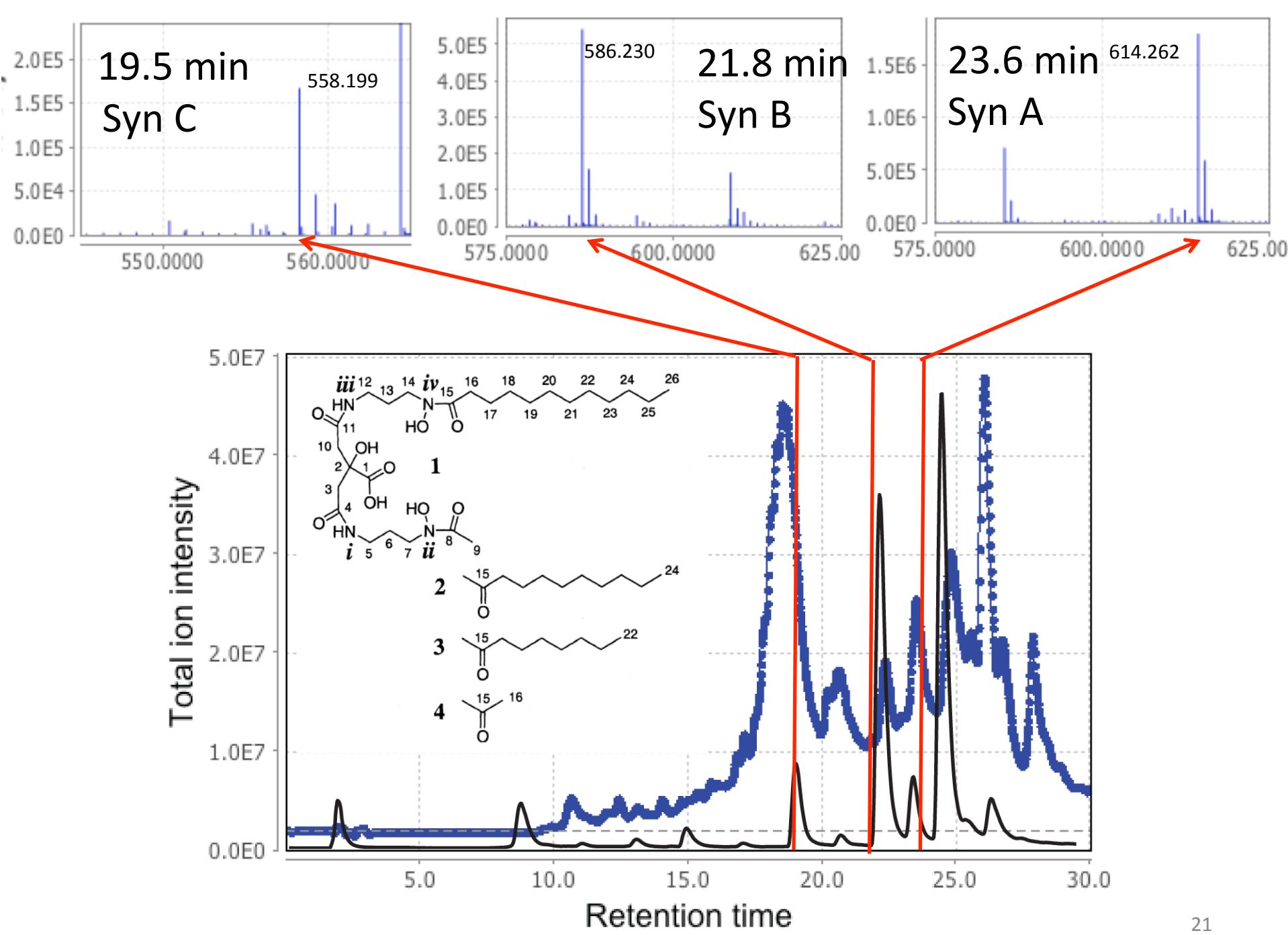


# Synechobactins

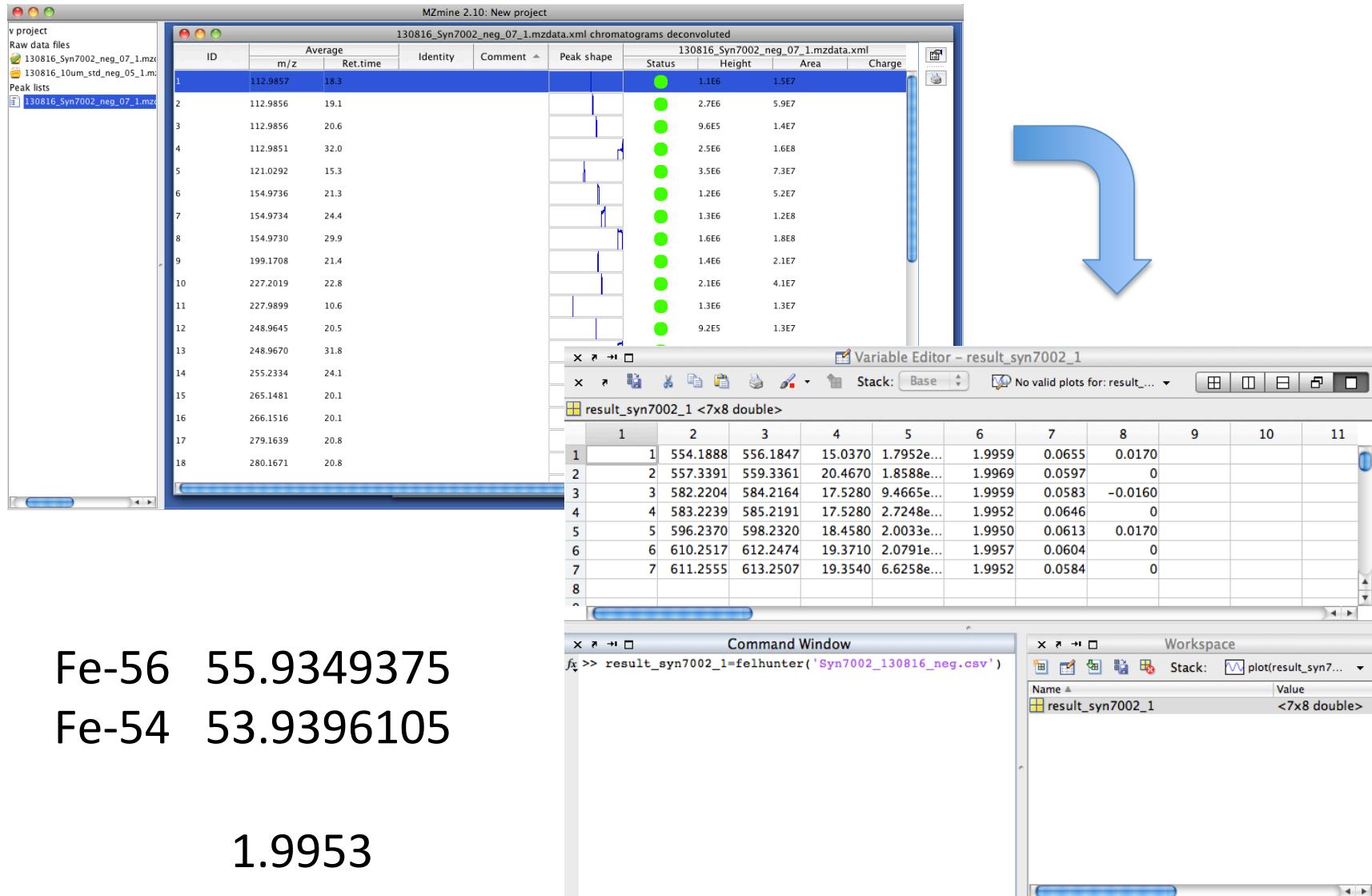


# Synechobactins

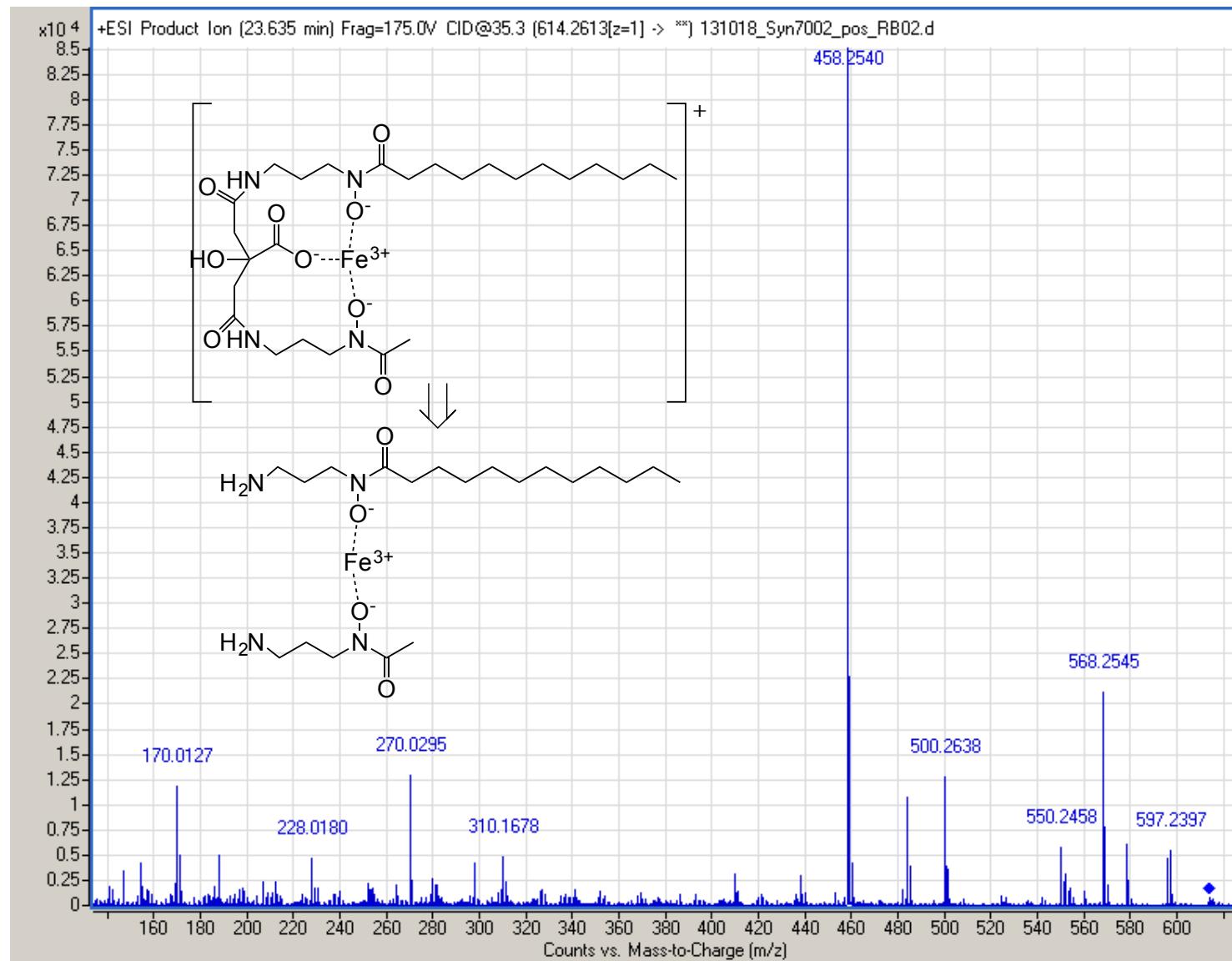




# Data processing



# MS/MS



# Novel Siderophore Discovery

Synechococcus 7002 Ligand list

ICP-MS Peak	Mass (M/Z)	Retention Time (min)	MS/MS Fragments	Ligand I.D
a	474.1	9.5	428.1	Schizokinin
c	672.3	13.4	572.3	Unknown
d	654.3	15.5		Unknown
f	558.2	19.4	512.2	Synechobactin C
h	586.2	21.8	540.2	Synechobactin B
i	571.3	22.7		Syn A – CO <sub>2</sub>
j	614.3	23.5	568.3	Synechobactin A
k	1198.5	23.6		Unknown
l	642.3	24.9	596.3	Synechobactin D

# Applications

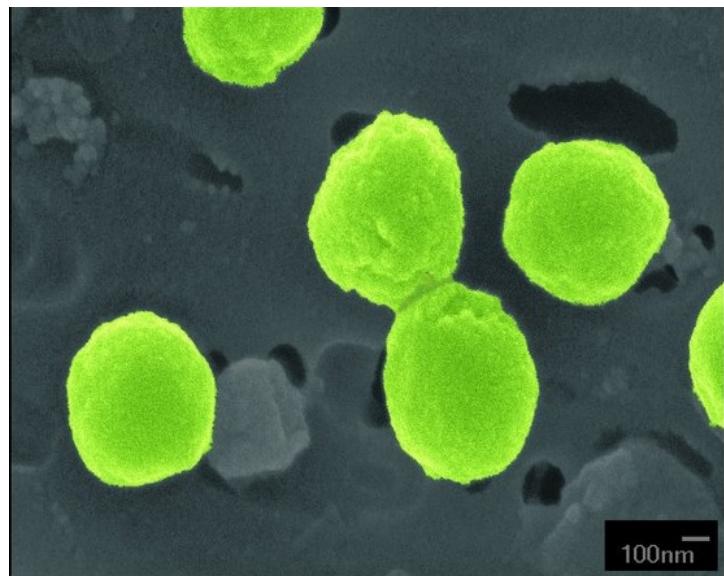
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## 1. Cyanobacteria cultures

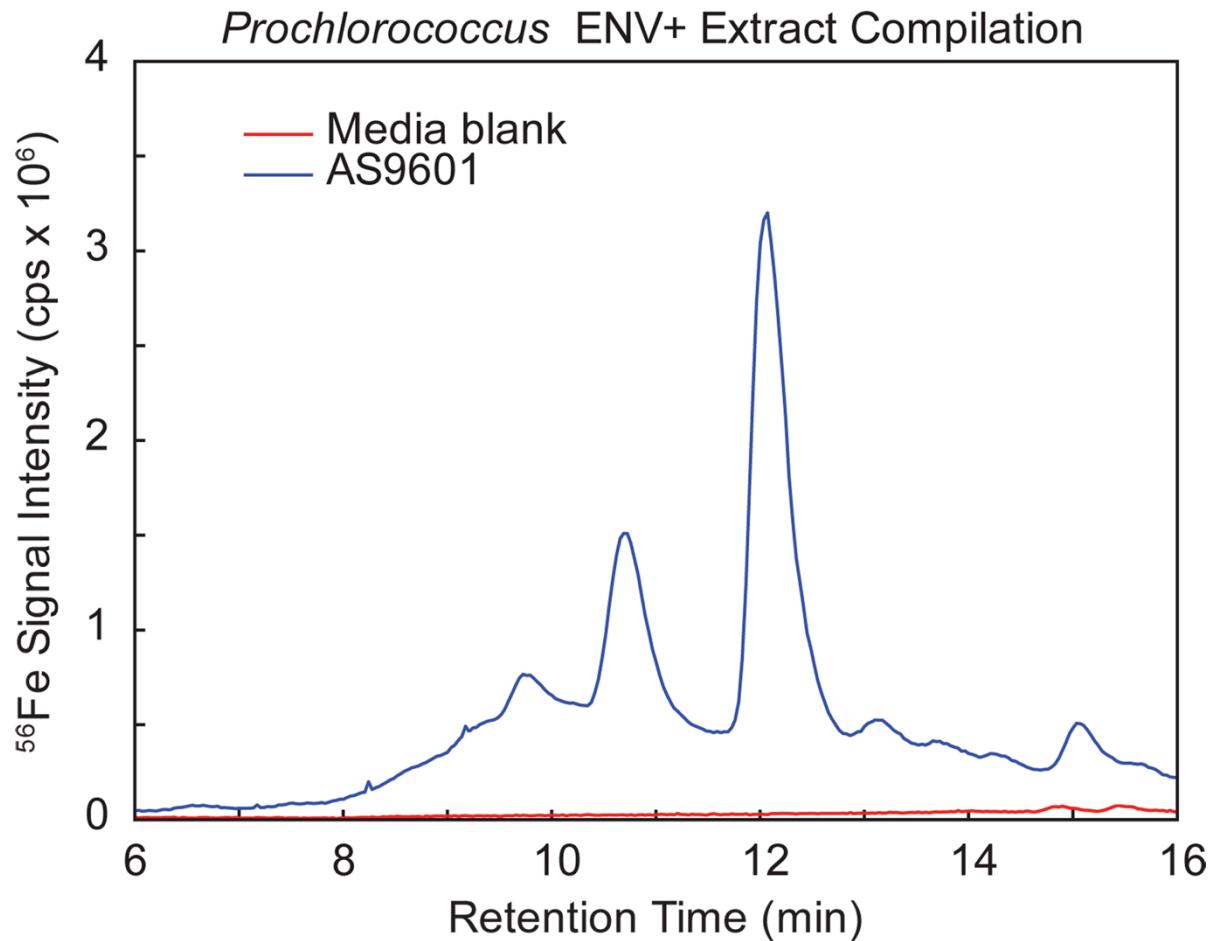
- *Synechococcus* strain PCC 7002
- ***Prochlorococcus***

## 2. Natural Seawater

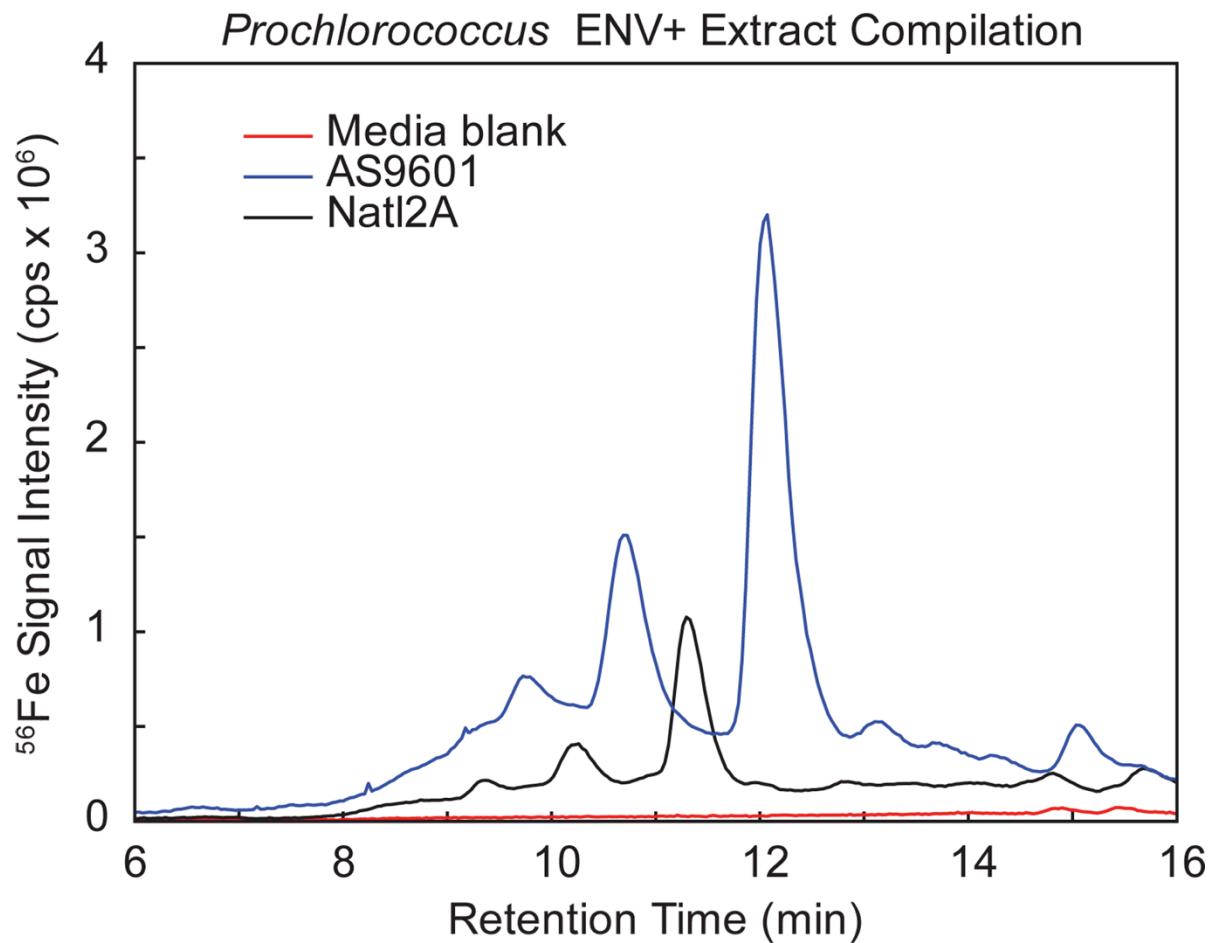
- Subtropical Pacific



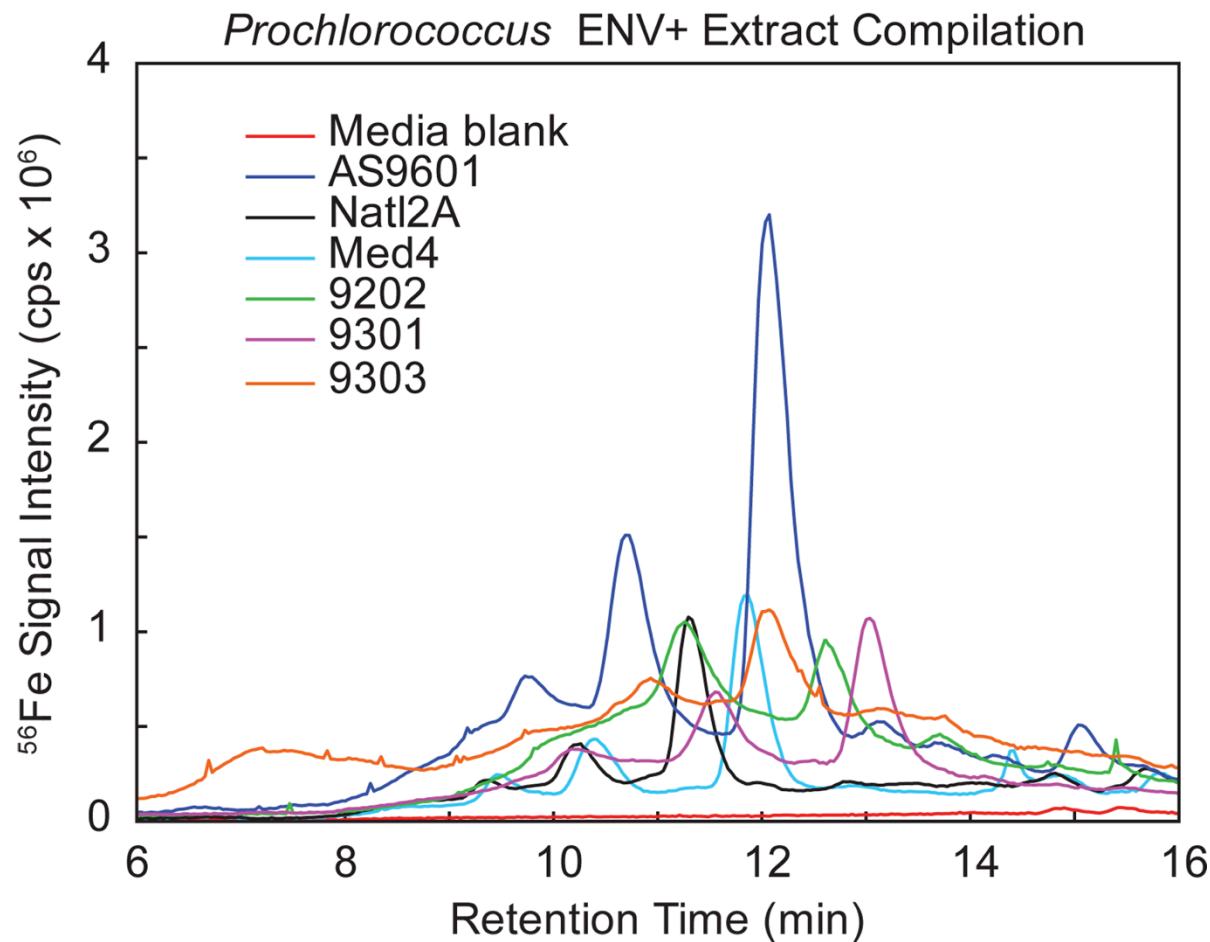
# *Prochlorococcus* Fe Ligands



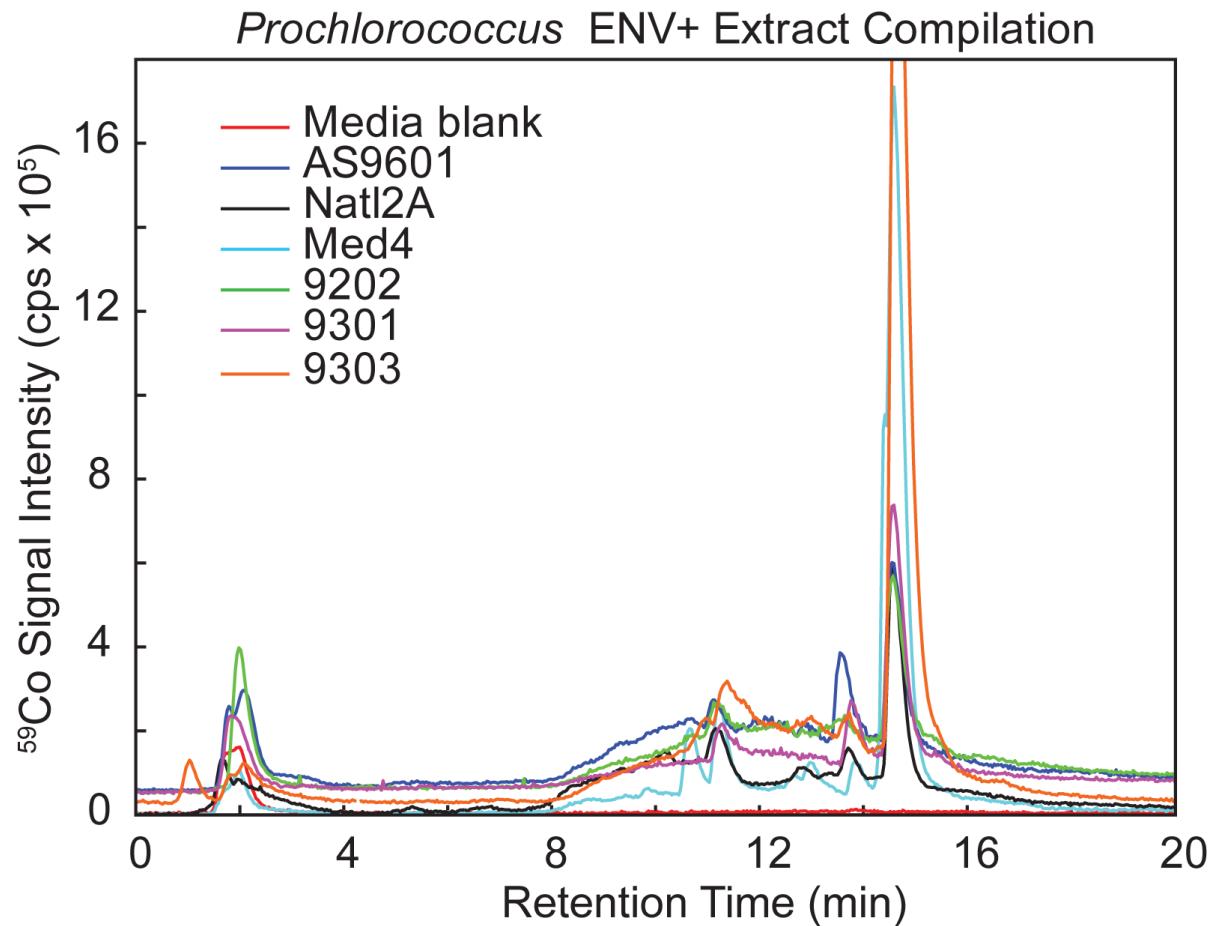
# *Prochlorococcus* Fe Ligands



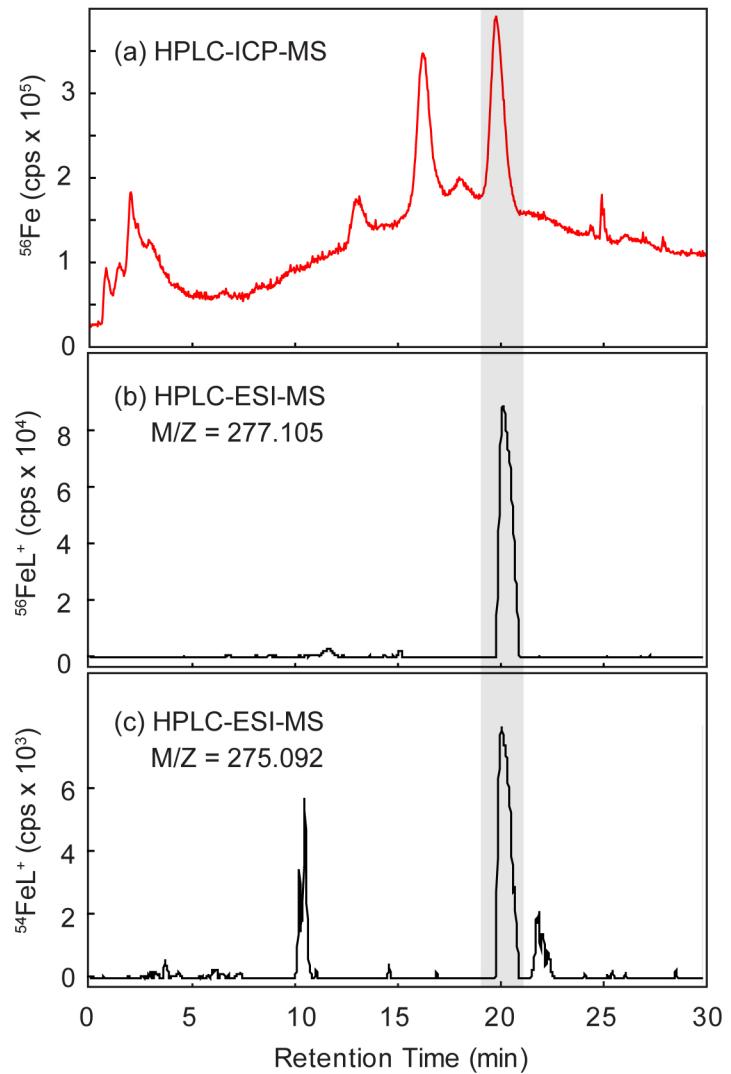
# *Prochlorococcus* Fe Ligands



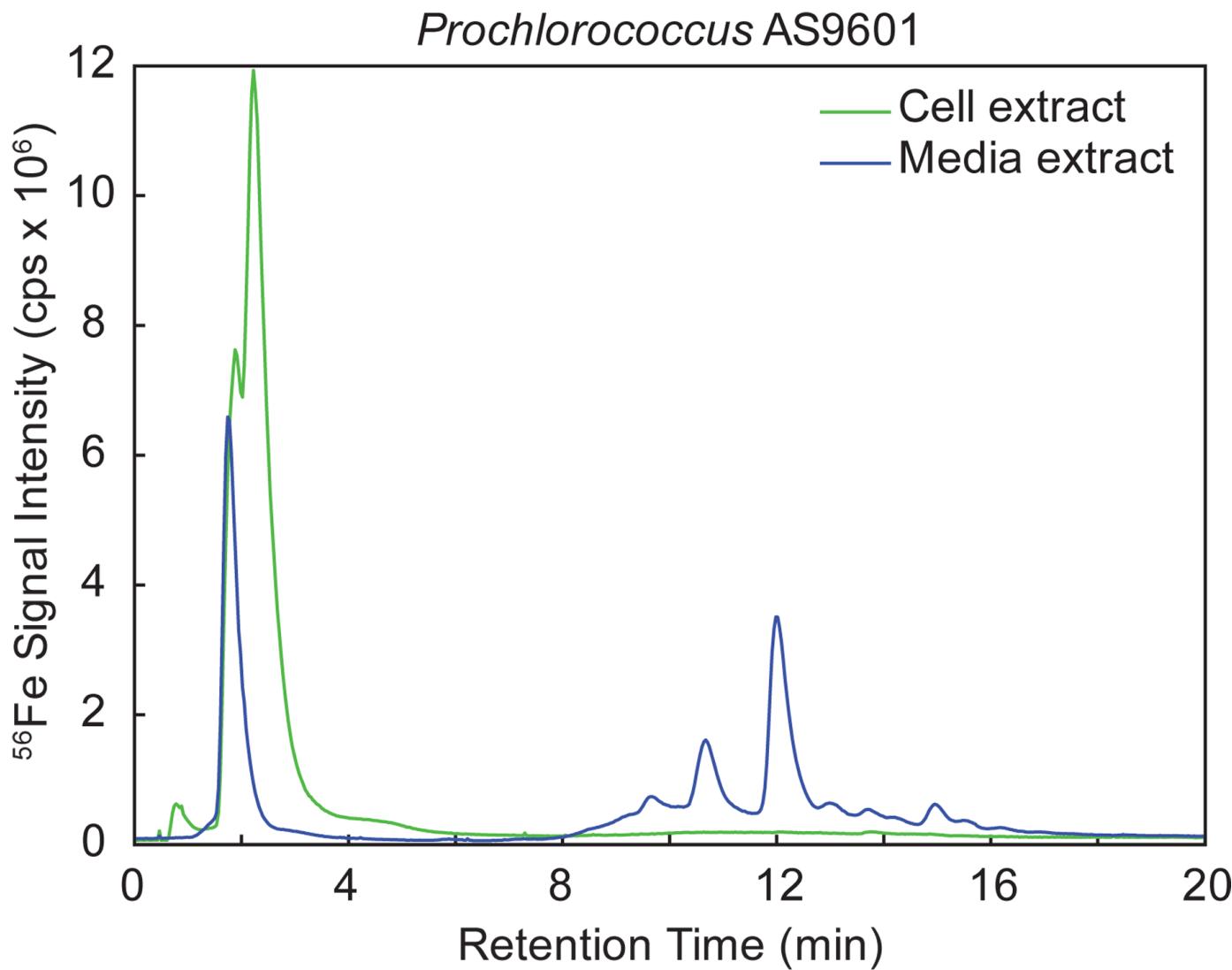
# *Prochlorococcus* Co Ligands



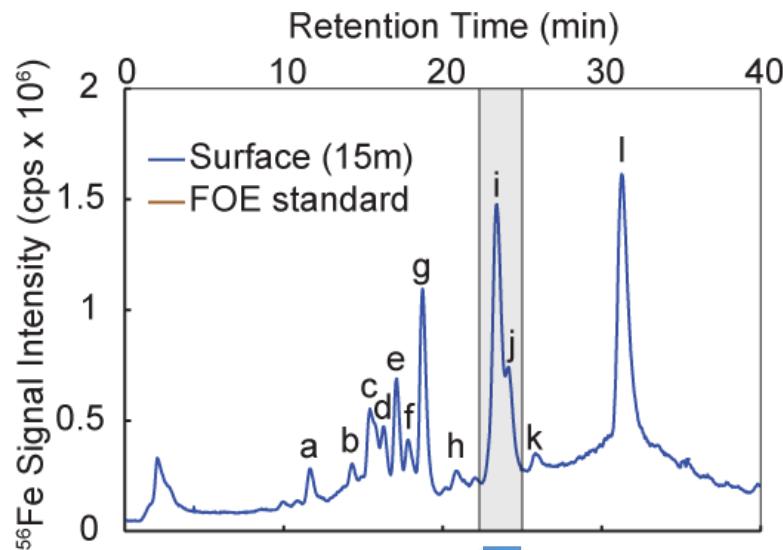
AS9601 Iron Ligand Characterization



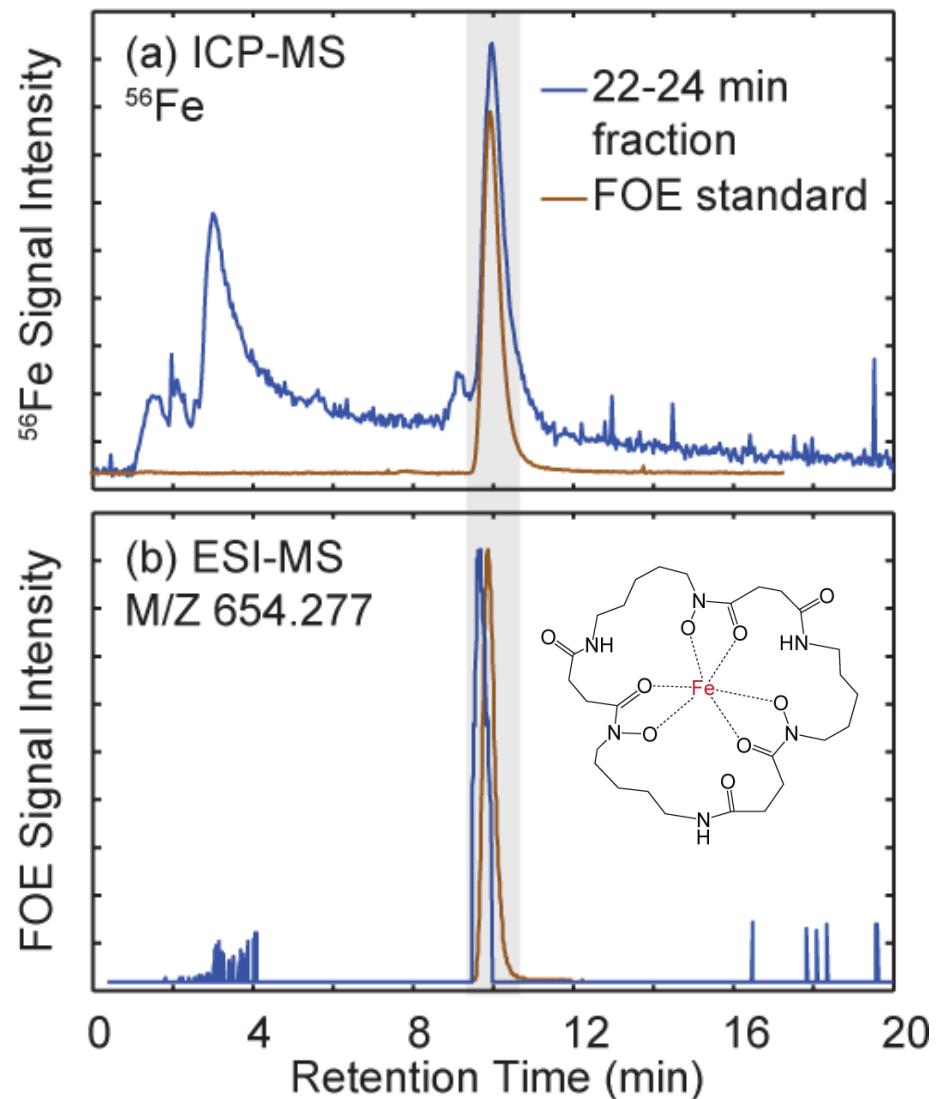
# *Prochlorococcus* Fe Ligands



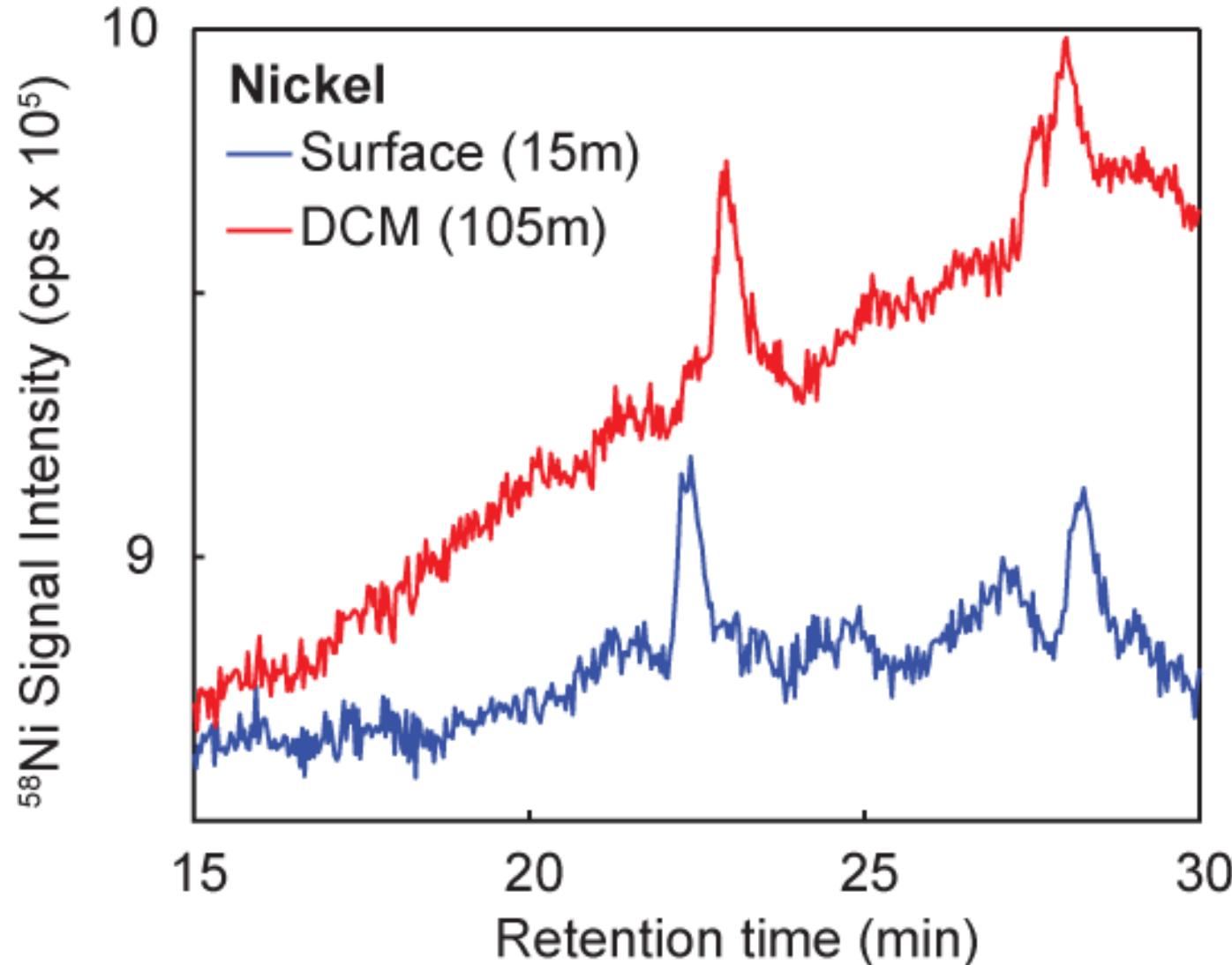
# Subtropical North Pacific (Hoe-Phor)



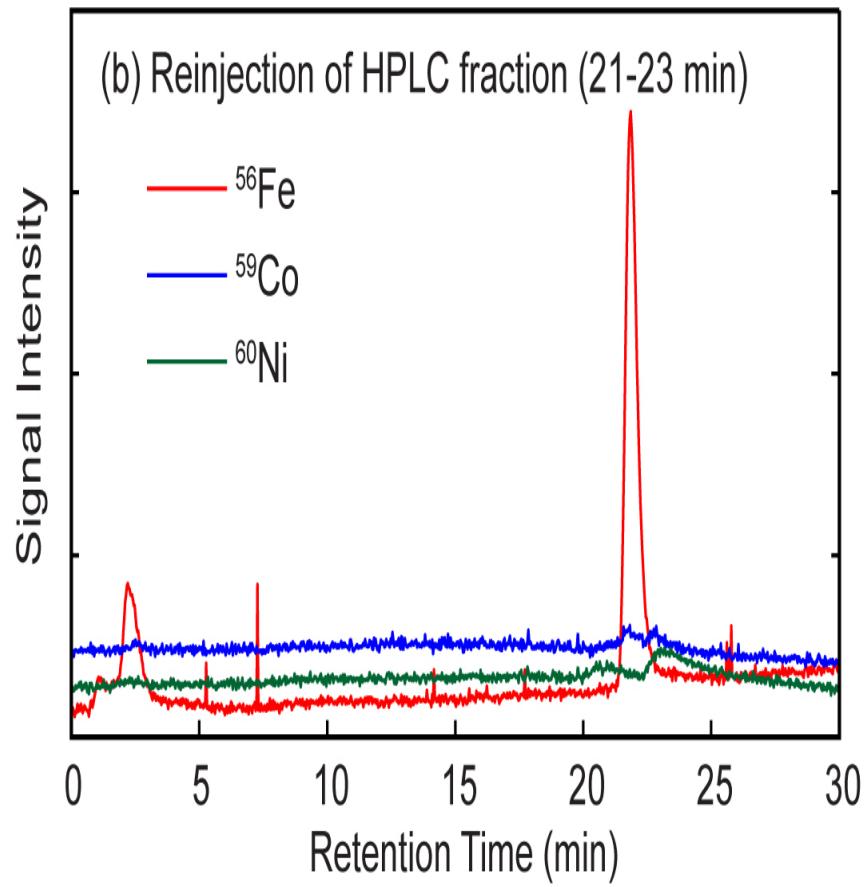
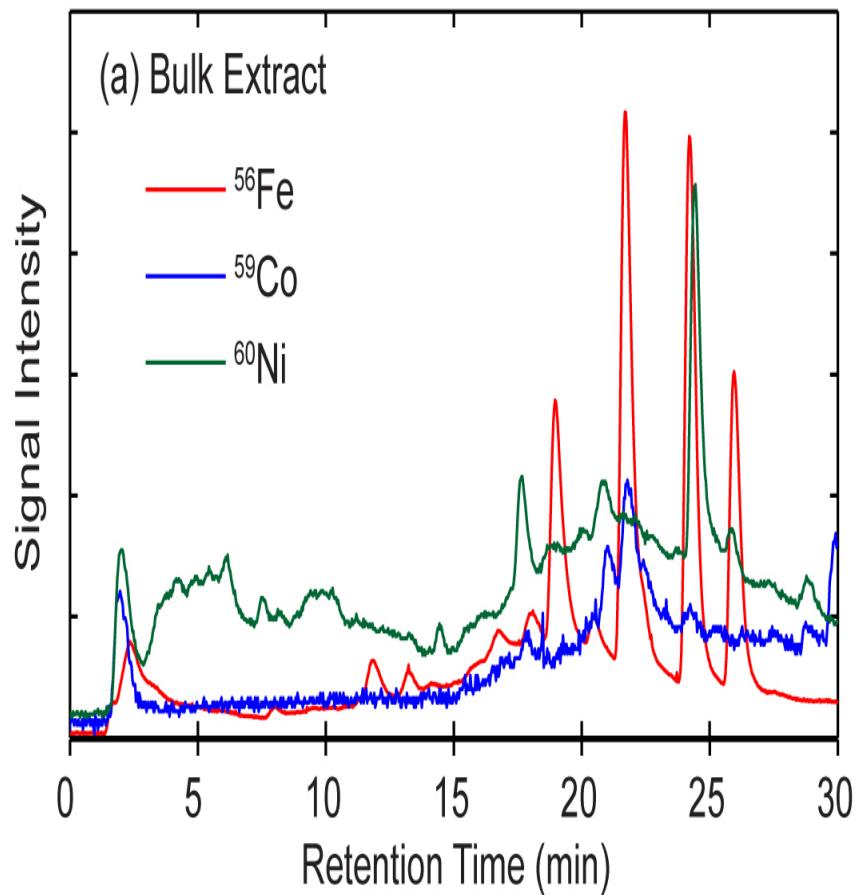
2<sup>nd</sup> separation  
on collected  
fractions



# Subtropical North Pacific (Hoe-Phor)



# Trace metal organic complexes in the South Pacific Subtropical Gyre



# Summary:

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- All marine microbes produce a very broad suite of organic compounds that are released from the cell (exometabolome) 100's-1000's of compounds from a single pure culture.
- To understand microbial interactions we somehow need to identify which of these compounds matter, and which don't. There are two approaches to this, untargeted and targeted.
- We typically don't find compounds, we find "features". We can turn a feature into a compound through spectral analysis.
- In the case of organometallics, we can use the metals to "find" the feature and begin to track it to source and biosynthetic pathway: link "omics" to geochemical cycling.