

# Behavior, physiology and the niche of marine phytoplankton

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Halifax, Nova Scotia, Canada B3H 4R2

2014 C-MORE Summer Training Course  
“Microbial Oceanography:  
Genomes to Biomes”

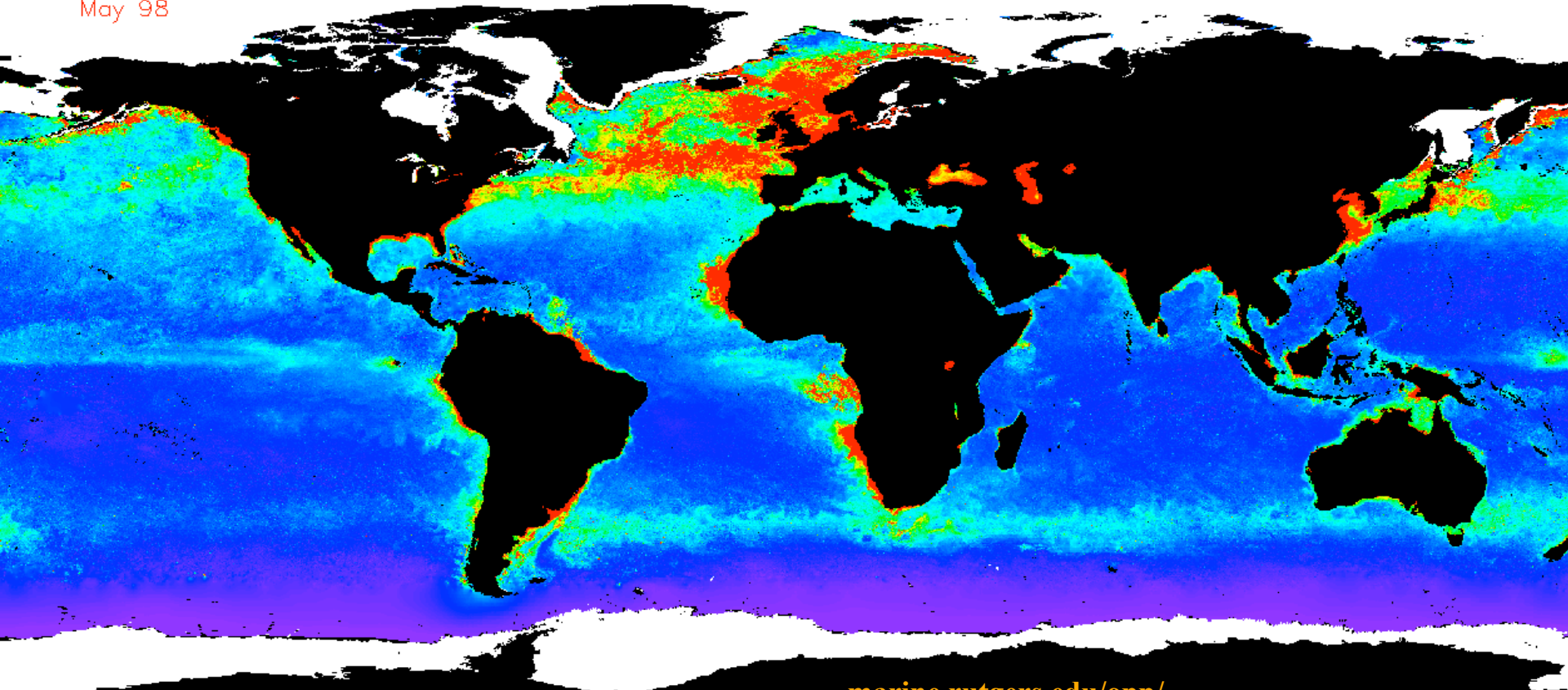
Ocean Tracking Network



# A principal goal of microbial oceanography

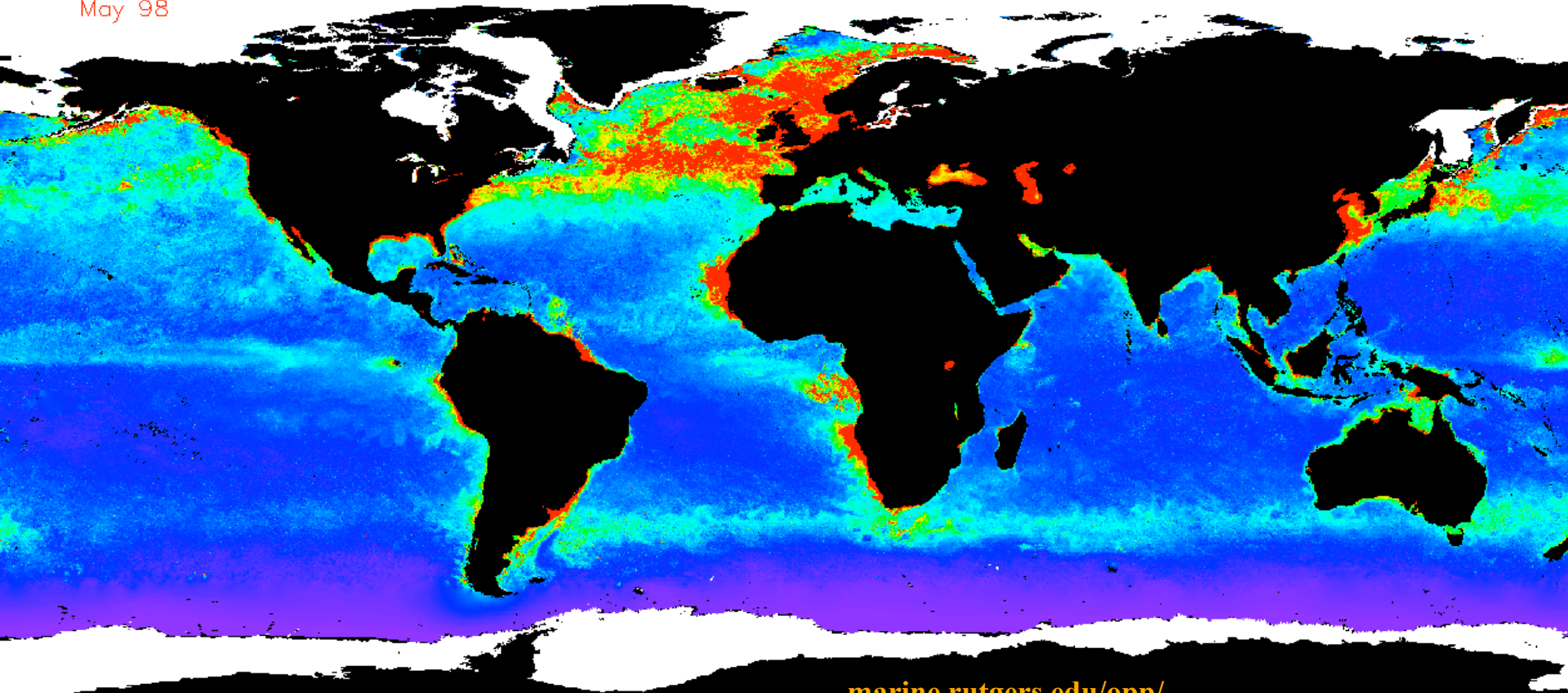
Describing and explaining the  
*distributions* and *activities* of marine microbes

May 98



...and using this information to describe the *causes* and *consequences* of variations in key biogeochemical processes

May 98

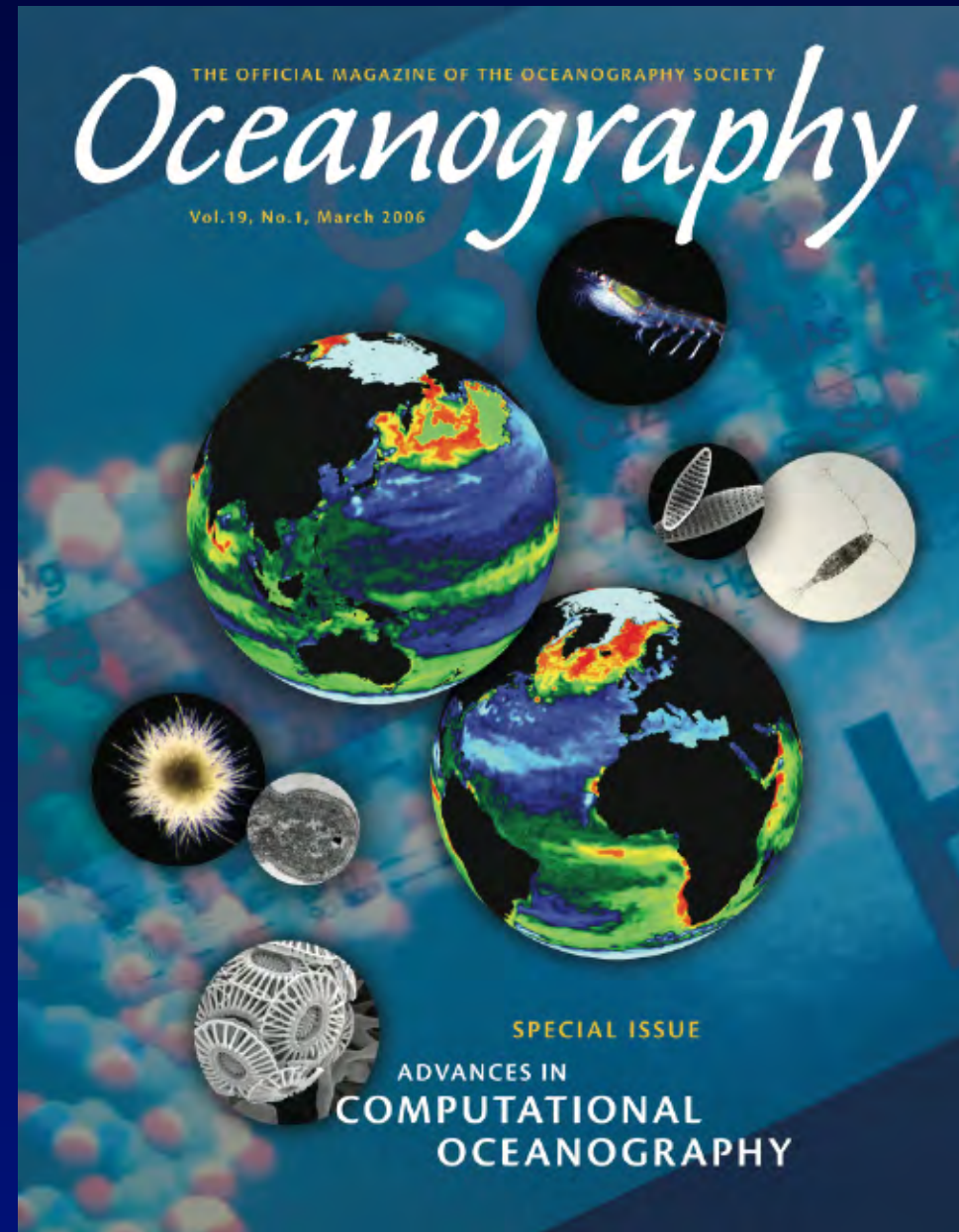


# Key biogeochemical processes

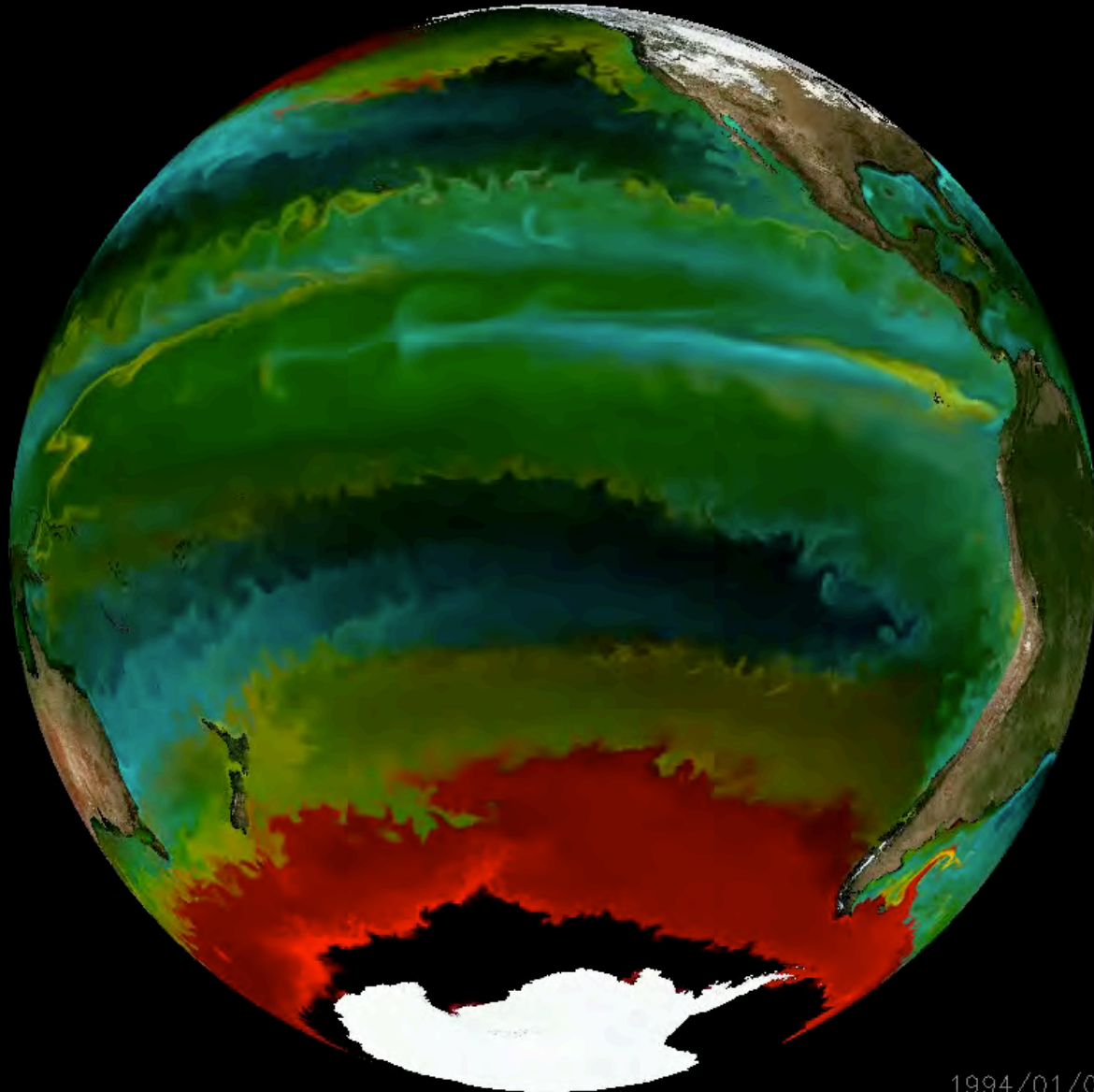
- Primary production
- Nitrogen fixation
- Denitrification
- Trace gas production
- ...and the many other processes that make those processes possible

This can be achieved only through an integrated approach

*The role of the oceans in Earth systems ecology, and the effects of climate variability on the ocean and its ecosystems, can be understood only by observing, describing, and ultimately predicting the state of the ocean as a physically forced ecological and biogeochemical system.*



# Models integrate information

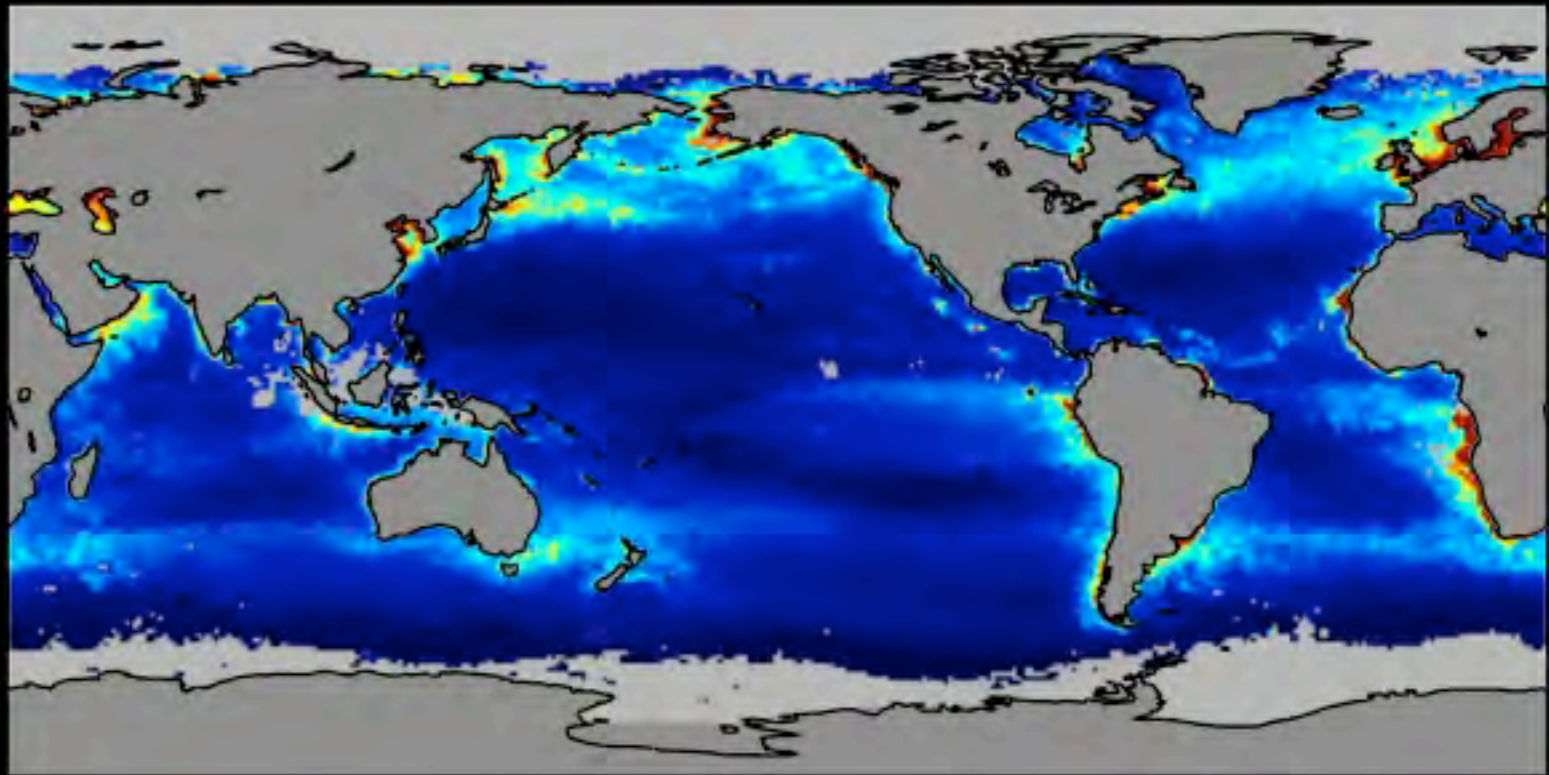


Mick Follows

1994/01/01

John Cullen: C-MORE 2014

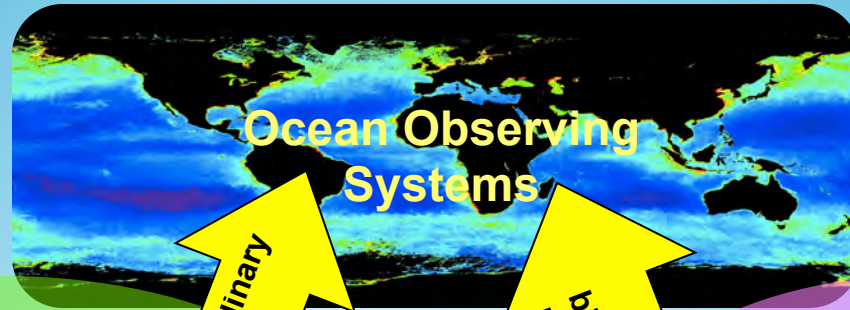
So do you



Year



# One Ultimate Target



Genomics & the other 'omics

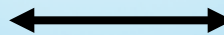
Integrated interdisciplinary sensor systems

Physical Chemical & Biological Oceanography

biogeochemical models  
Assimilative models

Hydrological & Atmospheric Optics [+Acoustics]

Process Studies



Theory / Models

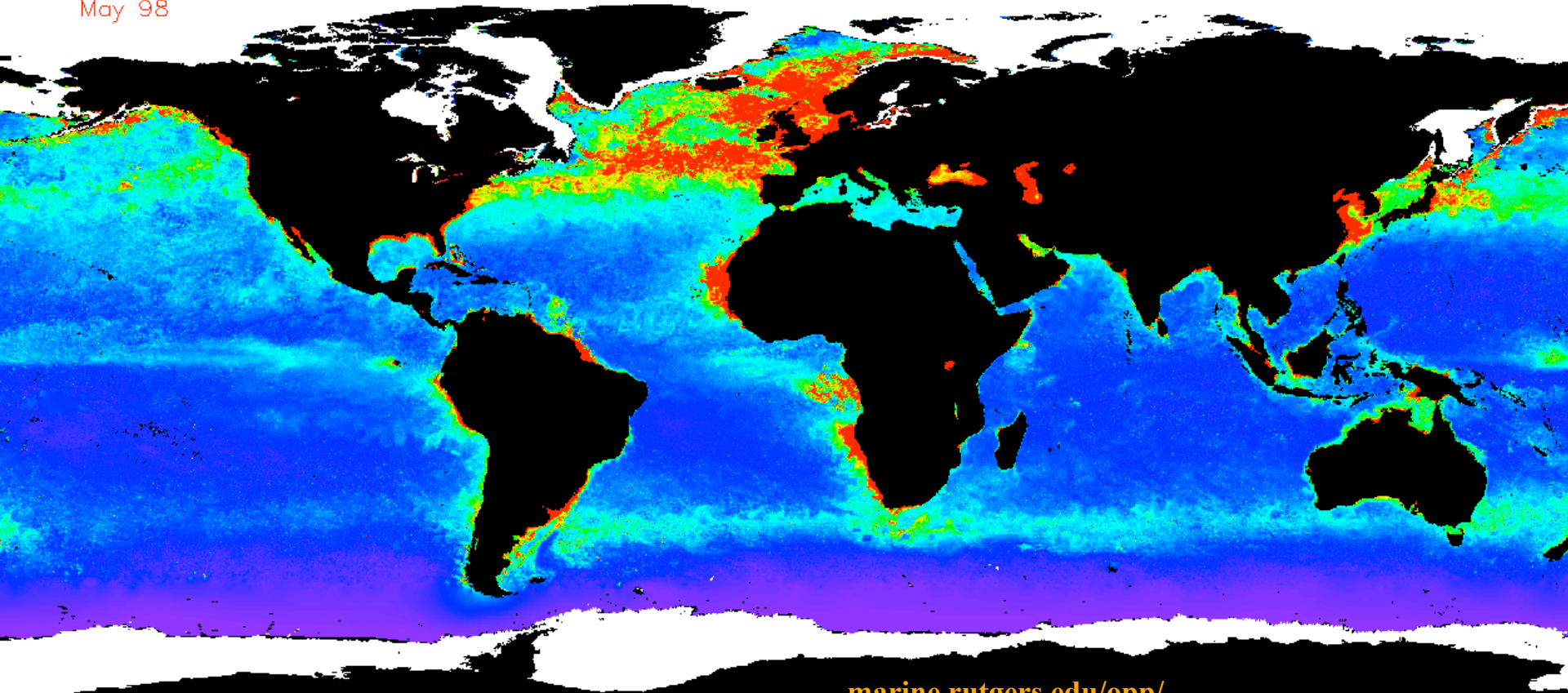
Physiological Ecology Including Gene Expression



# Biological oceanography and phytoplankton ecology

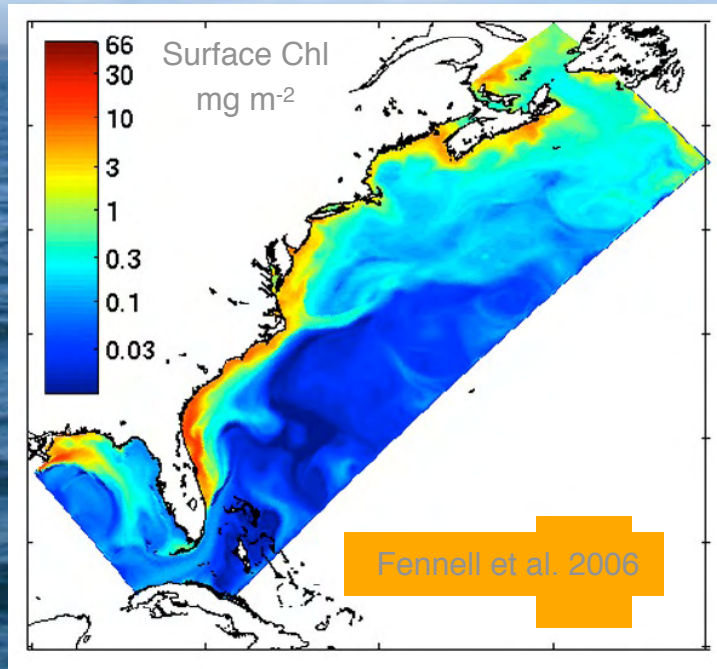
Describe the *causes* and *consequences* of variations in primary productivity (and food web structure)

May 98

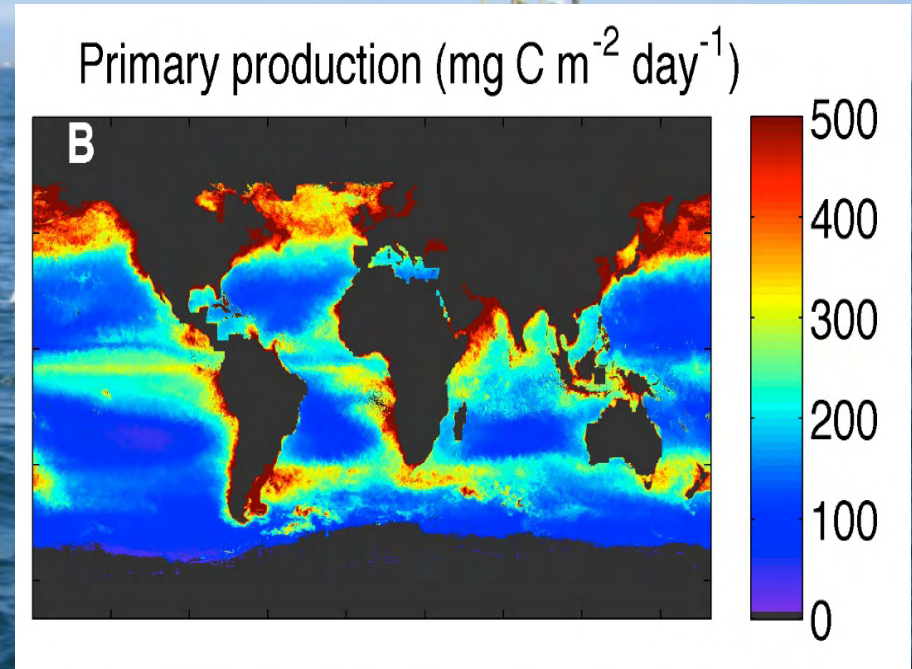


# An overview of established approaches to marine modelling

## Prognostic Model



## Diagnostic Model



# Approach #1: Observation, analysis, inference (empirical, diagnostic models)

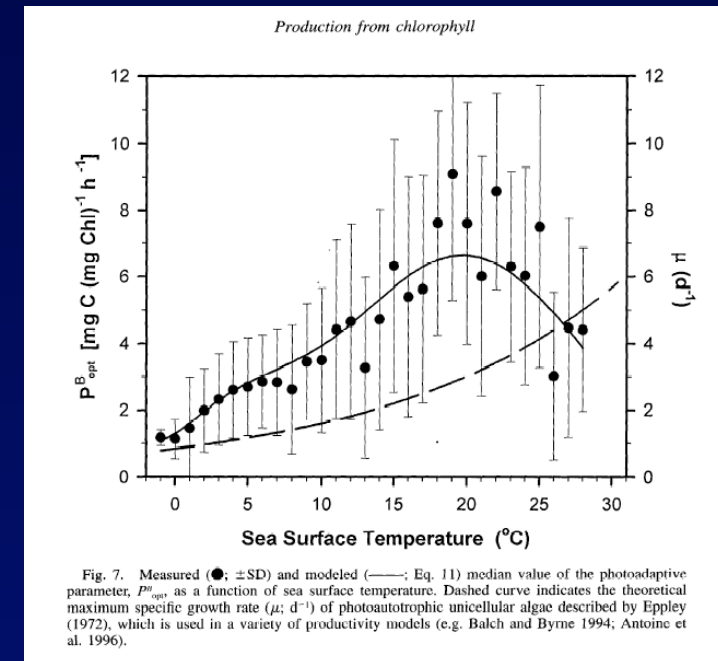
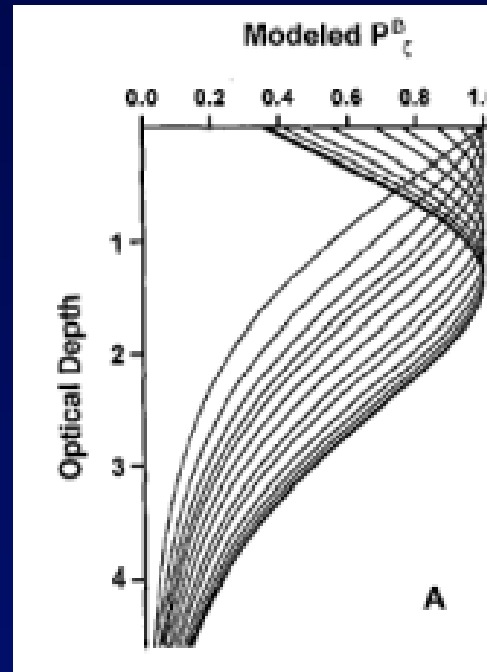
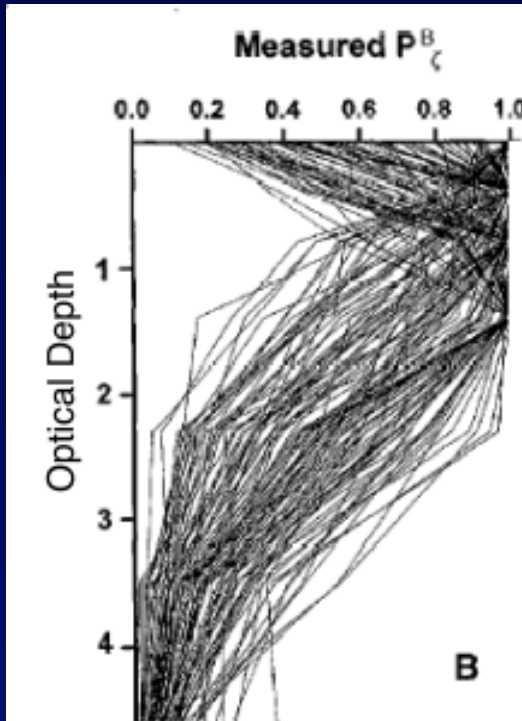


Fig. 7. Measured (●;  $\pm$ SD) and modeled (—; Eq. 11) median value of the photoadaptive parameter,  $P_{opt}^B$  as a function of sea surface temperature. Dashed curve indicates the theoretical maximum specific growth rate ( $\mu$ ;  $d^{-1}$ ) of photoautotrophic unicellular algae described by Eppley (1972), which is used in a variety of productivity models (e.g. Balch and Byrne 1994; Antoine et al. 1996).

Behrenfeld and Falkowski 1997b L&O

Modeling the pattern in the measurements — not necessarily primary productivity!

# Biospheric Primary Production During an ENSO Transition

Michael J. Behrenfeld,<sup>1\*</sup> James T. Randerson,<sup>2</sup>  
Charles R. McClain,<sup>1</sup> Gene C. Feldman,<sup>1</sup> Sietse O. Los,<sup>3</sup>  
Compton J. Tucker,<sup>1</sup> Paul G. Falkowski,<sup>4</sup> Christopher B. Field,<sup>5</sup>  
Robert Frouin,<sup>6</sup> Wayne E. Esaias,<sup>1</sup> Dorota D. Kolber,<sup>4</sup>  
Nathan H. Pollack<sup>7</sup>

Result: Quantitative predictions that  
are as good as the data &  
assumptions that go into them

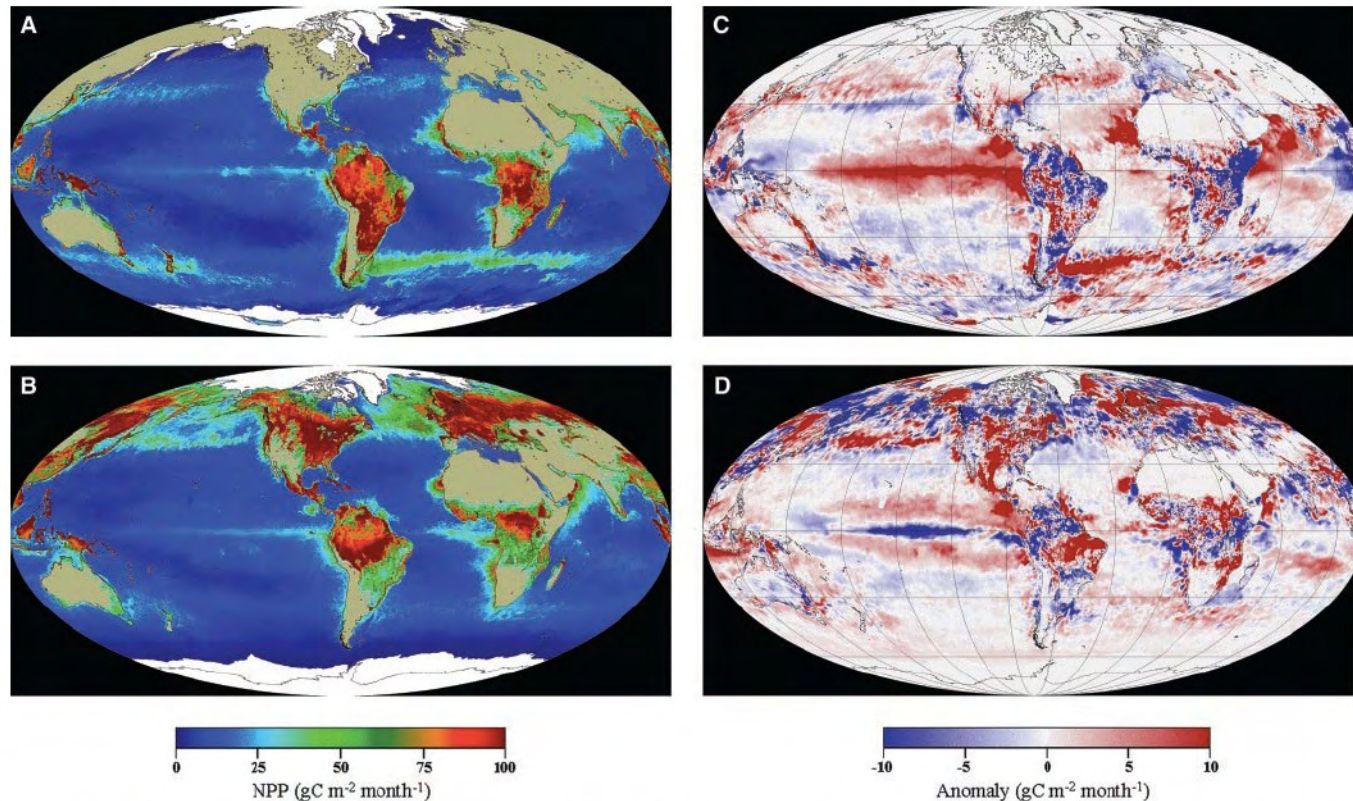
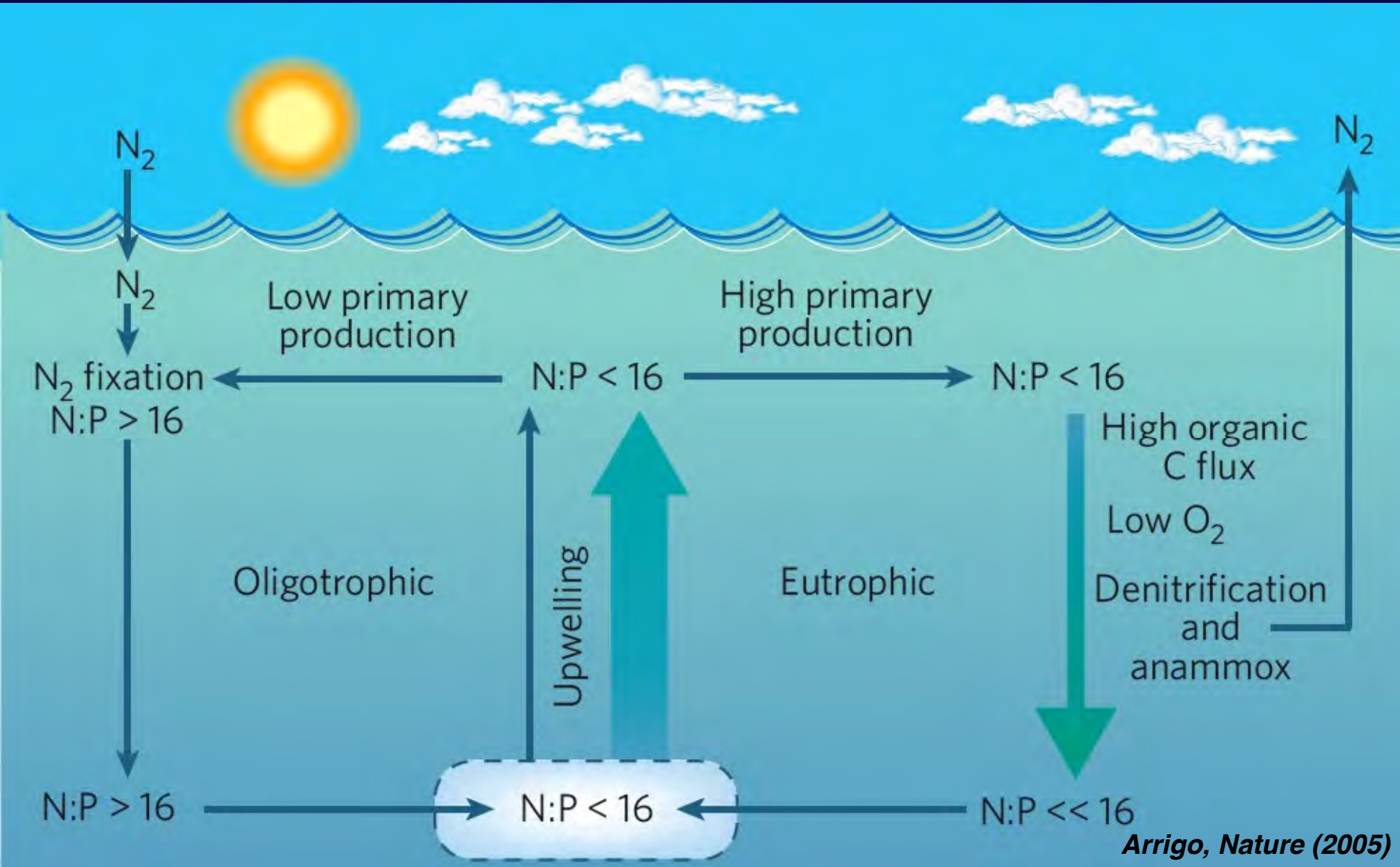


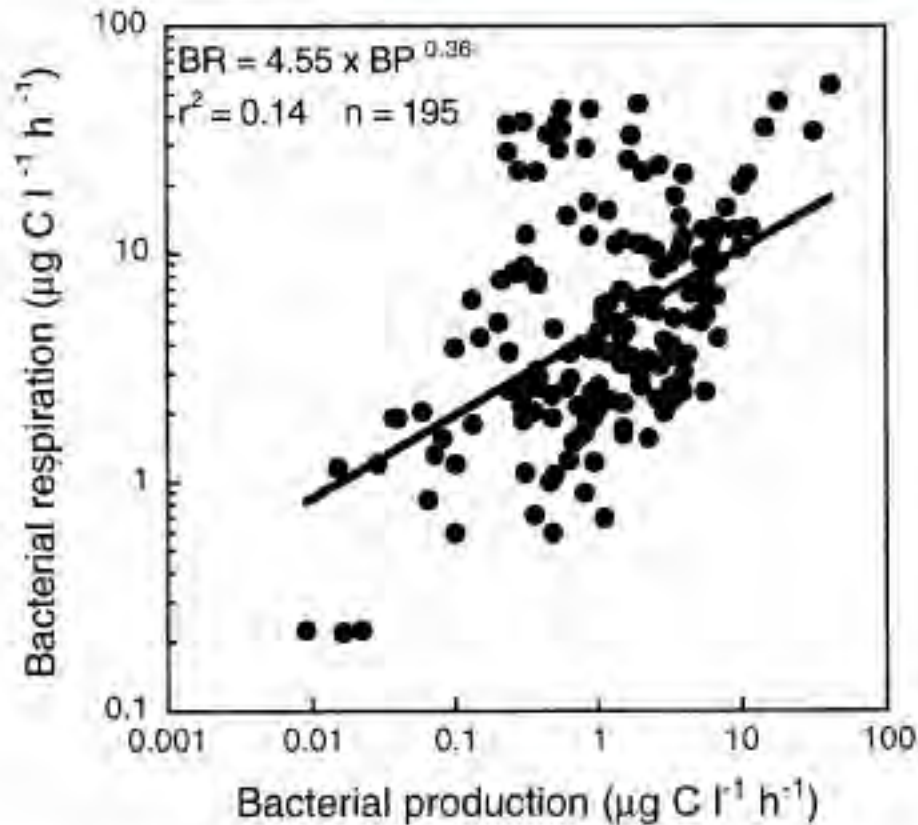
Fig. 2. Seasonal average and interannual differences in biospheric NPP ( $\text{g C m}^{-2} \text{ month}^{-1}$ ) estimated with SeaWiFS data and the integrated 3D NPP model (A). Interannual differences in NPP (B).

to La Niña conditions resulted in substantial regional changes in NPP, as illustrated by interannual differences in Austral summer NPP (i.e., summer NPP for December 1999 to February 2000 minus summer NPP

# Approach #2: Observation, analysis, inference (qualitative, mechanistic, predictive models)

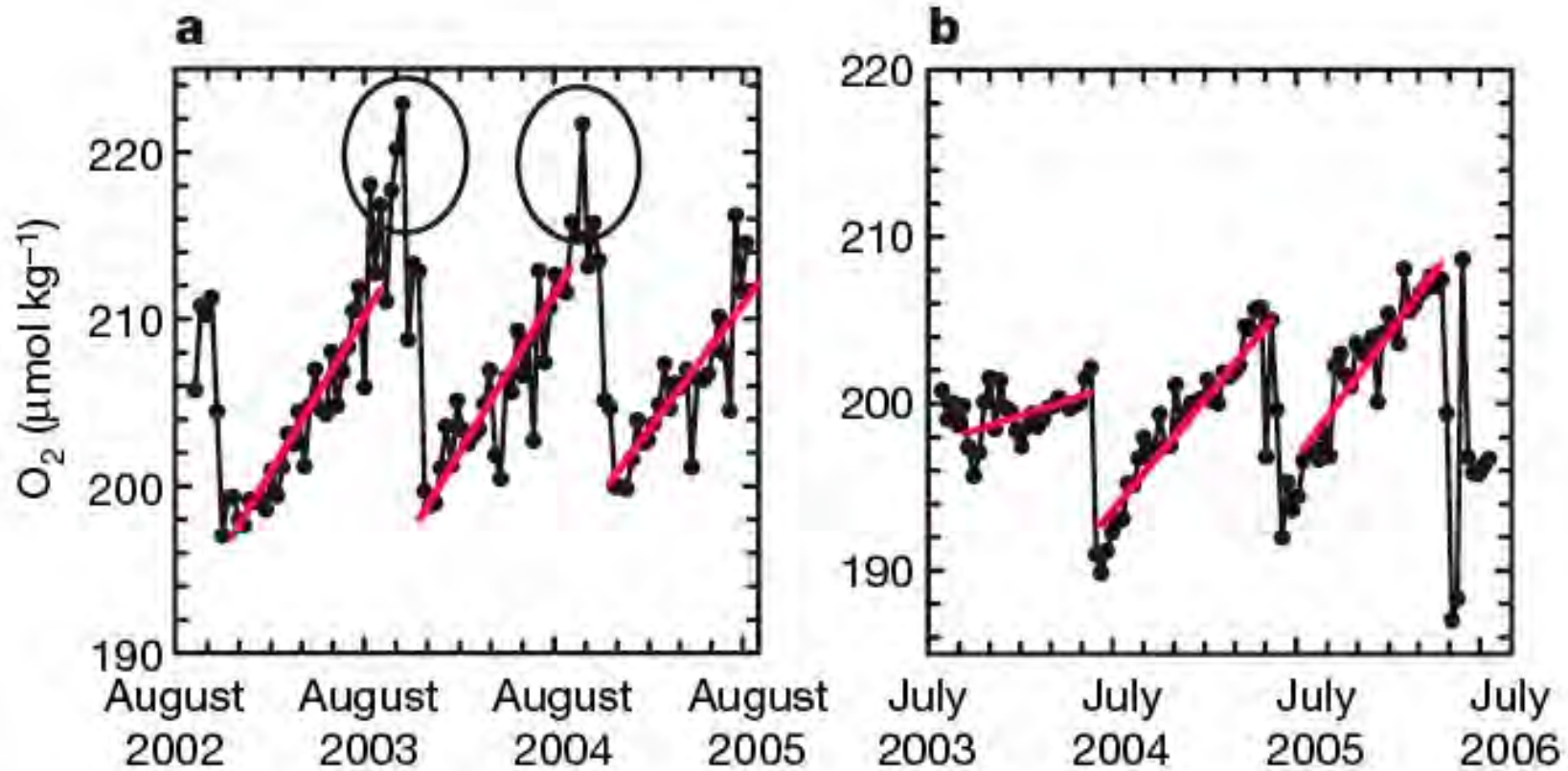


# The testing of qualitative, mechanistic, predictive models may be messy



**Figure 4.** Bacterial respiration as a function of bacterial production in aquatic ecosystems. The data are paired observations of bacterial respiration (*BR*) and production (*BP*); the sources of these data appear in Table 1. The line is the least-squares fit to the log-transformed data.

But predictions can be tested with appropriate observations



**Figure 3 | Oxygen concentrations in the SOM versus time. Oxygen**

Riser and Johnson Nature 2008

# Diagnostic models can be used to test hypotheses rather than to consolidate and interpret observations

## ARTICLE

doi:10.1038/nature09403

### Ocean nutrient ratios governed by plankton biogeography

Thomas S. Weber<sup>1</sup> & Curtis Deutsch<sup>1</sup>

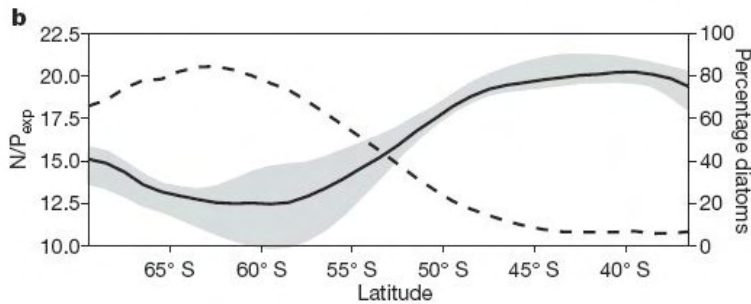


Figure 3 | Diagnosed nutrient export ratios. a, Spatial pattern of  $N/P_{exp}$ ,

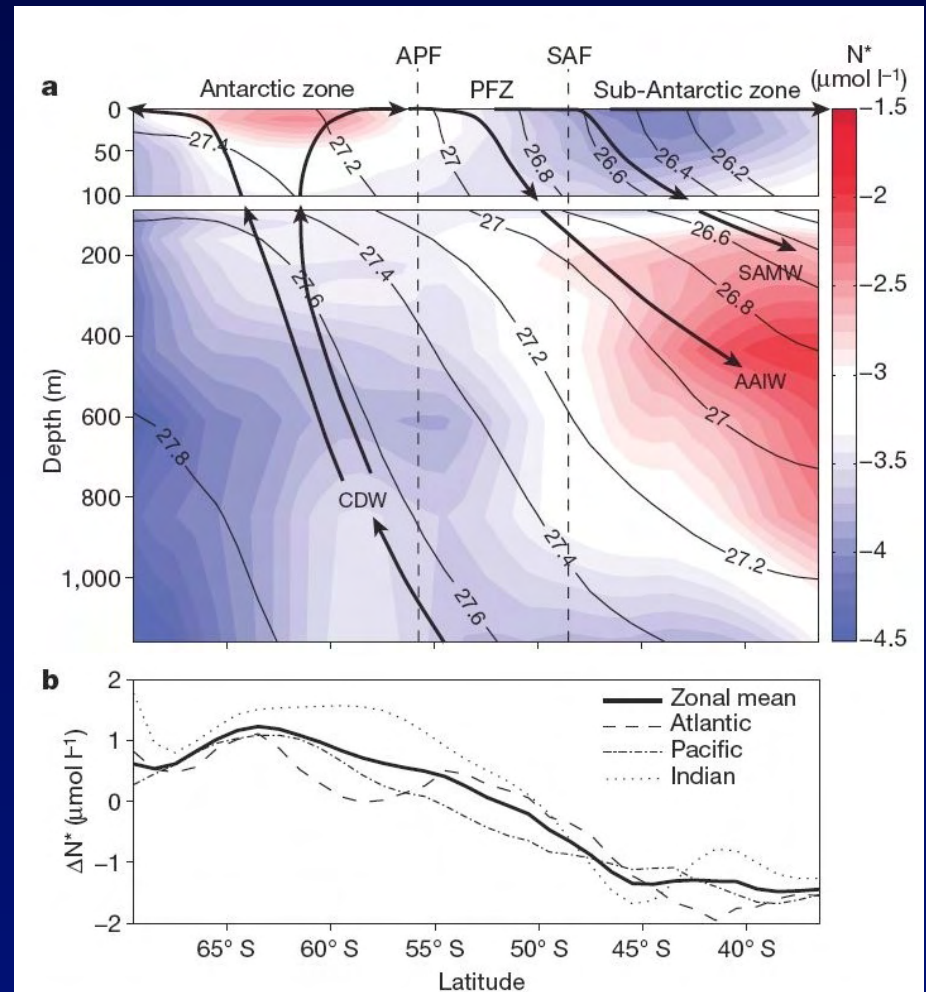


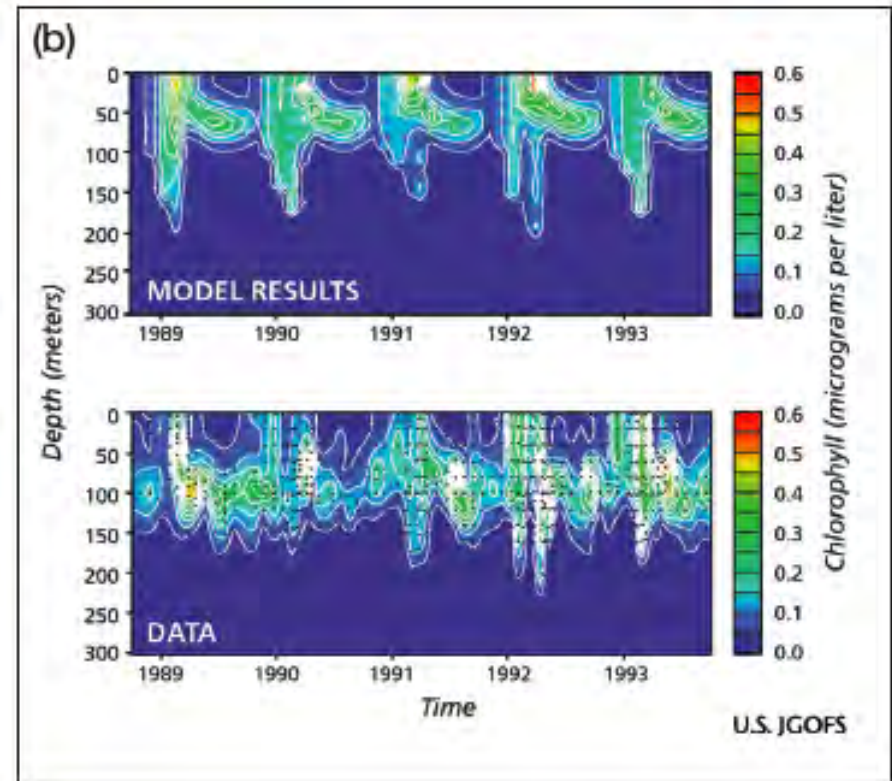
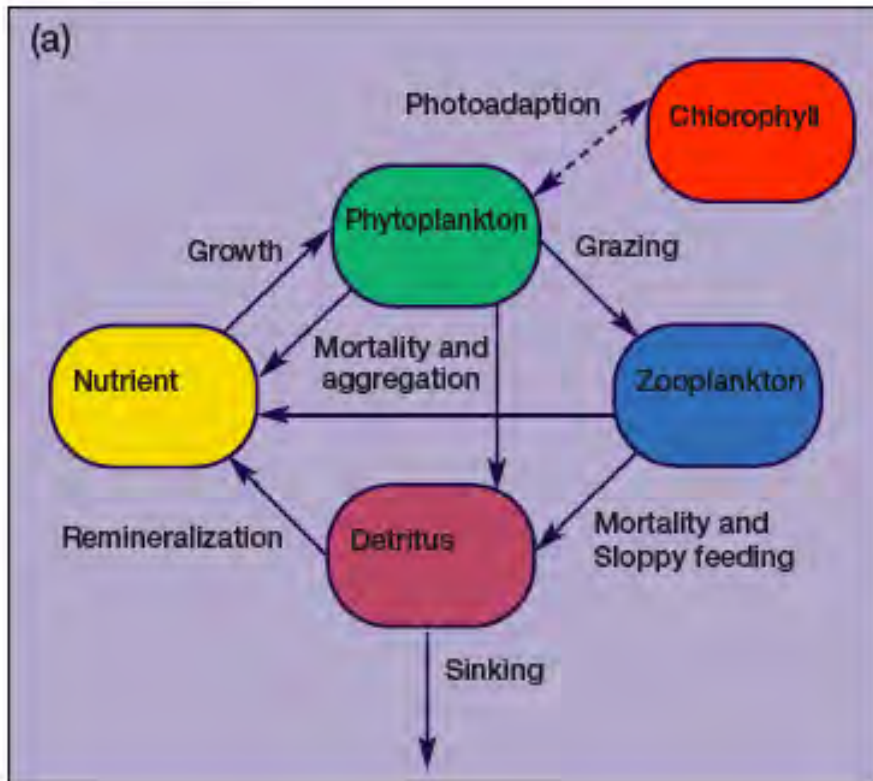
Figure 1 | Observed  $N^*$  distribution in the Southern Ocean. a, Zonal mean



# Approach #3: Prognostic models (quantitative, mechanistic, predictive models) -- e.g., BOCGMs

Ocean genomics

SC Doney et al.



Doney, S. C., M. R. Abbott, J. J. Cullen, D. M. Karl, and L. Rothstein. 2004. From genes to ecosystems: the ocean's new frontier. *Frontiers in Ecology and the Environment* 2: 457-466.

John Cullen: C-MORE 2014

# Complexity is added to increase realism and to test hypotheses

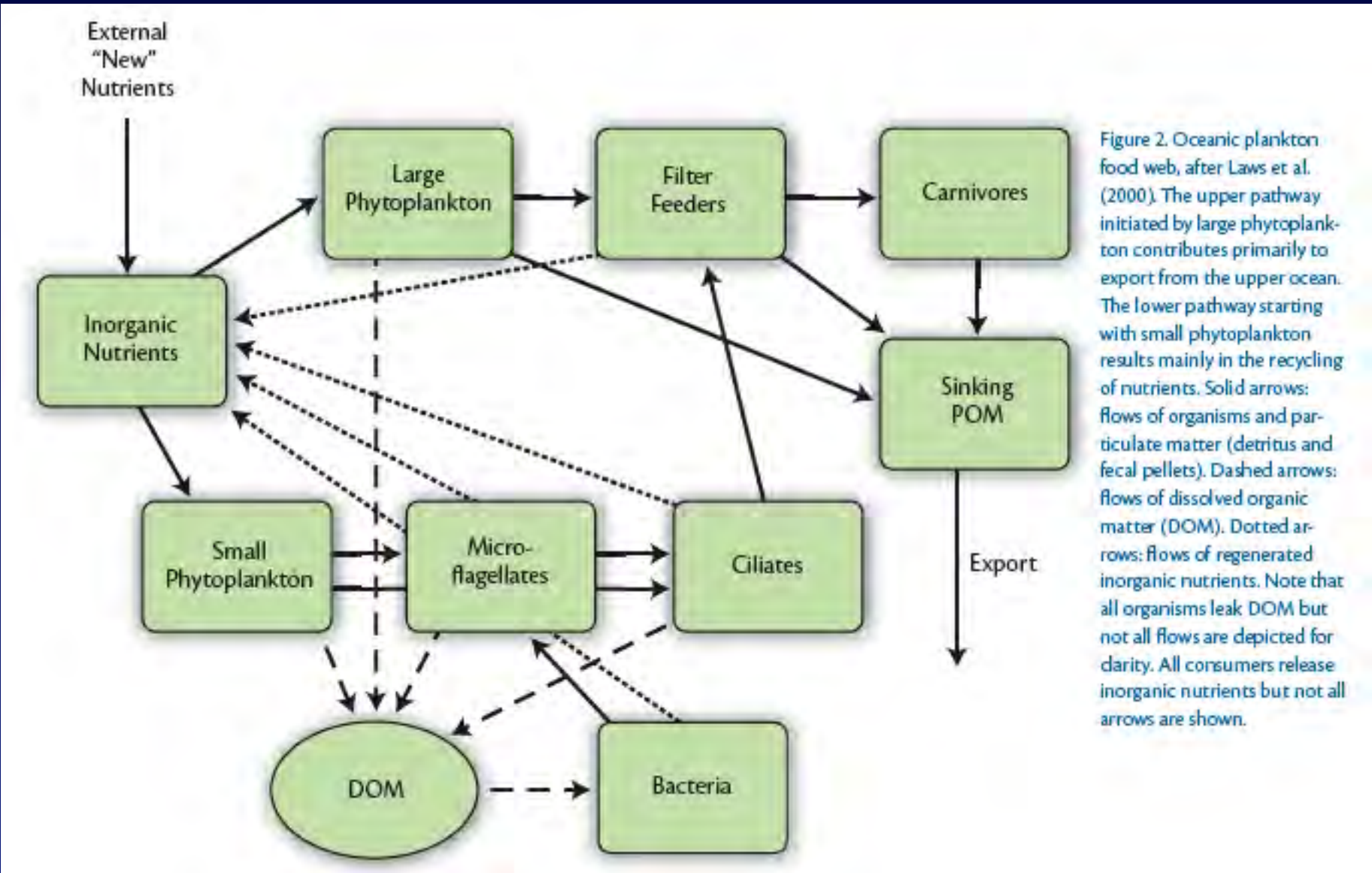
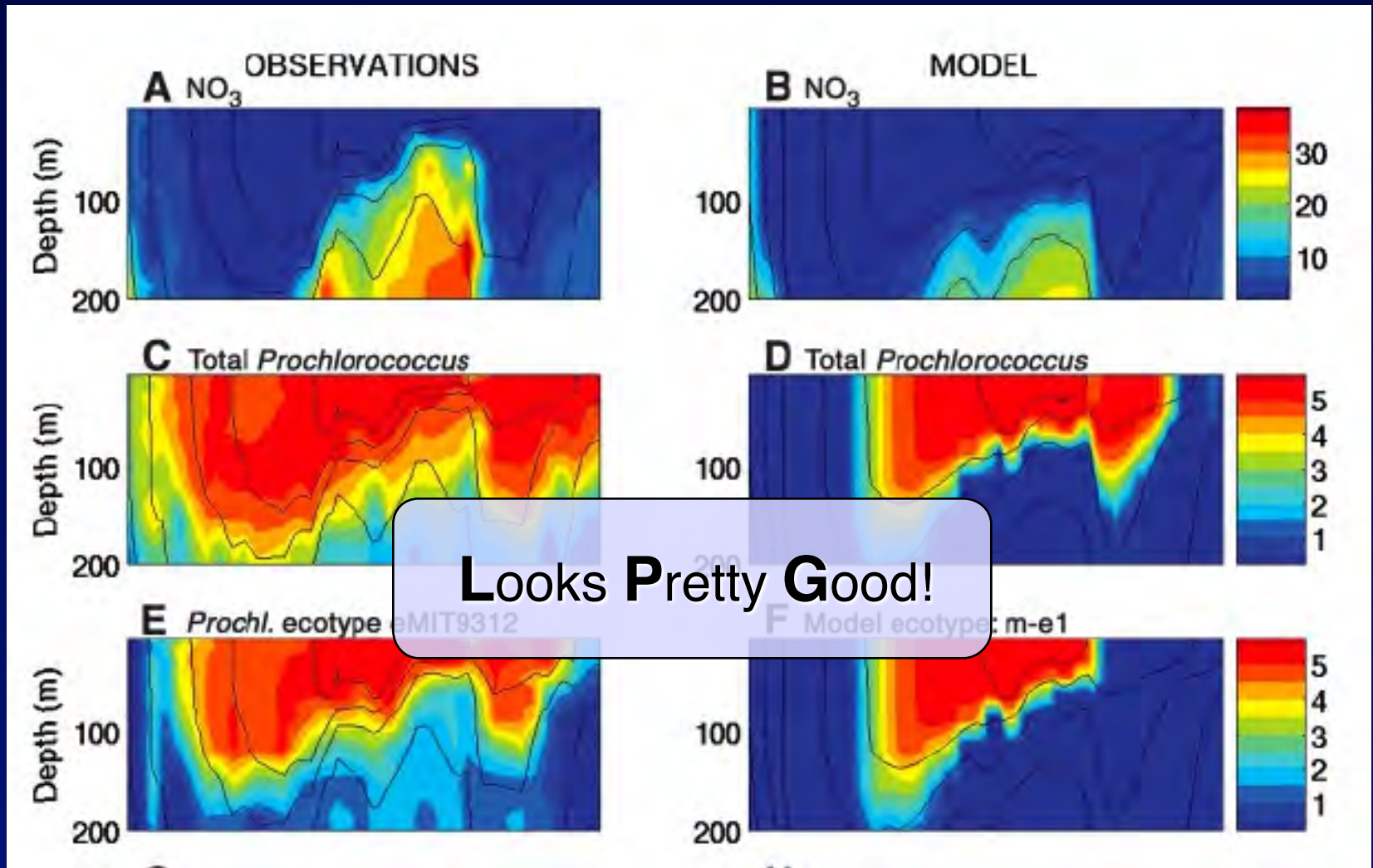


Figure 2. Oceanic plankton food web, after Laws et al. (2000). The upper pathway initiated by large phytoplankton contributes primarily to export from the upper ocean. The lower pathway starting with small phytoplankton results mainly in the recycling of nutrients. Solid arrows: flows of organisms and particulate matter (detritus and fecal pellets). Dashed arrows: flows of dissolved organic matter (DOM). Dotted arrows: flows of regenerated inorganic nutrients. Note that all organisms leak DOM but not all flows are depicted for clarity. All consumers release inorganic nutrients but not all arrows are shown.

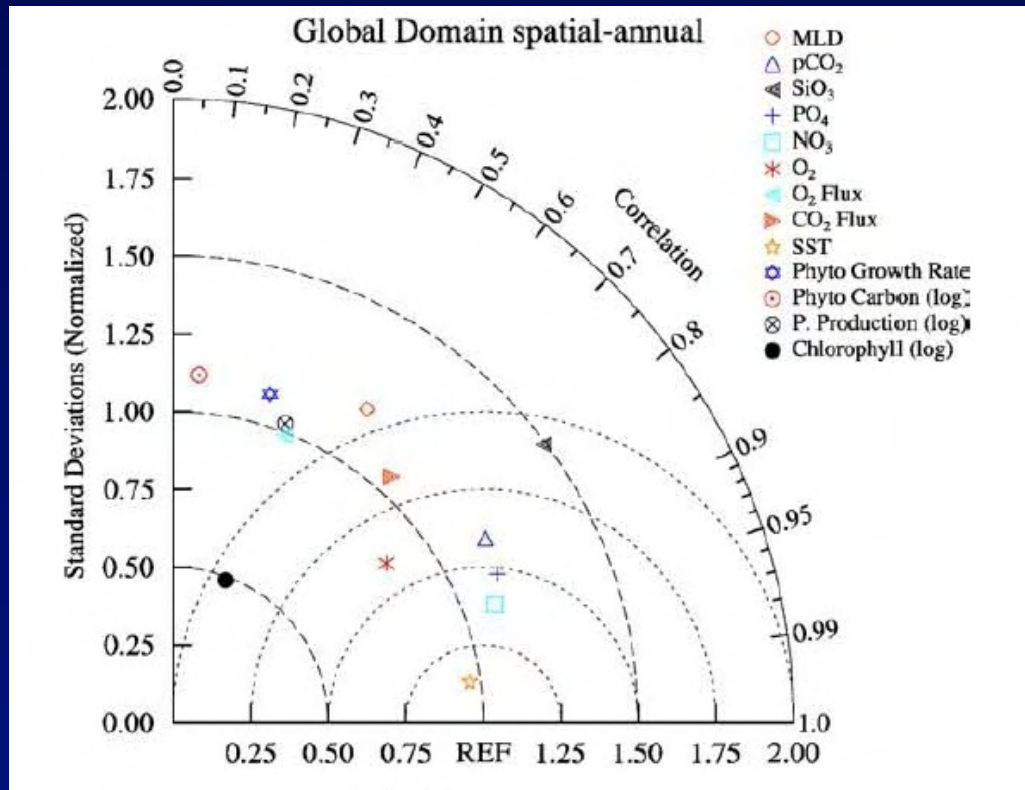
# Conventionally tested by the “LPG” criterion — but that is changing



Follows, M. J., S. Dutkiewicz, S. Grant, and S. W. Chisholm. 2007. Emergent biogeography of microbial communities in a model ocean. *Science* 315: 1843-1846.

## Skill metrics for confronting global upper ocean ecosystem-biogeochemistry models against field and remote sensing data

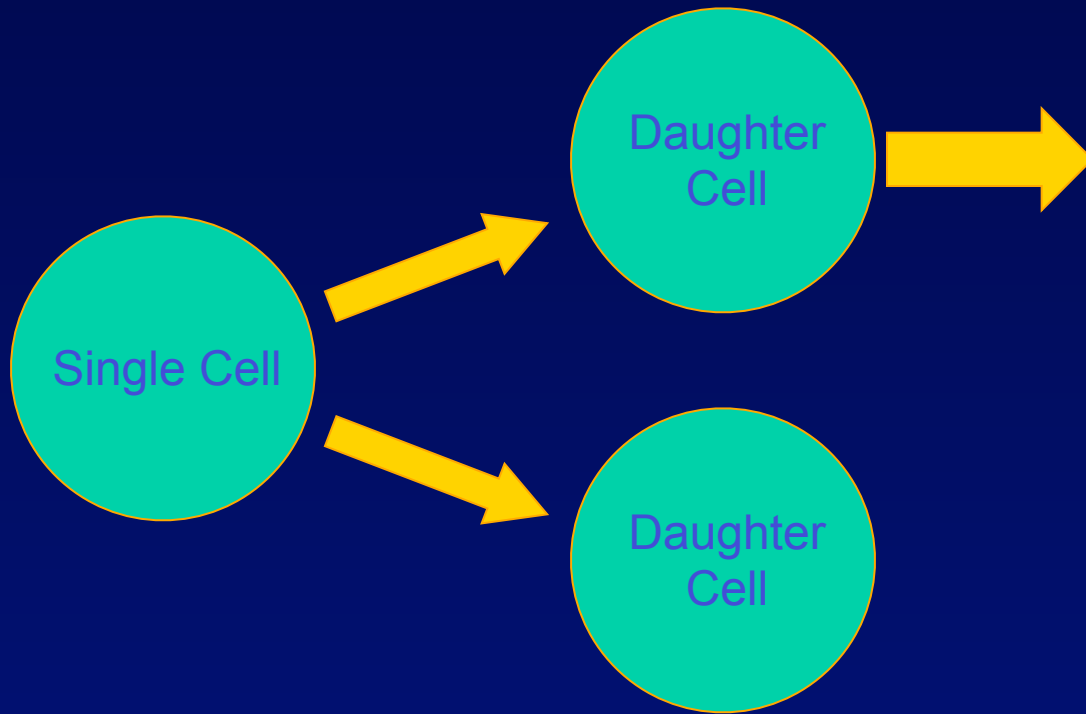
Scott C. Doney<sup>a,\*</sup>, Ivan Lima<sup>a</sup>, J. Keith Moore<sup>b</sup>, Keith Lindsay<sup>c</sup>, Michael J. Behrenfeld<sup>d</sup>,  
Toby K. Westberry<sup>d</sup>, Natalie Mahowald<sup>e</sup>, David M. Glover<sup>a</sup>, Taro Takahashi<sup>f</sup>



e.g., Taylor diagrams

# Fundamentally, ecosystem models should predict population dynamics

**Growth**



**Loss**

Accumulate (Bloom)

Be eaten

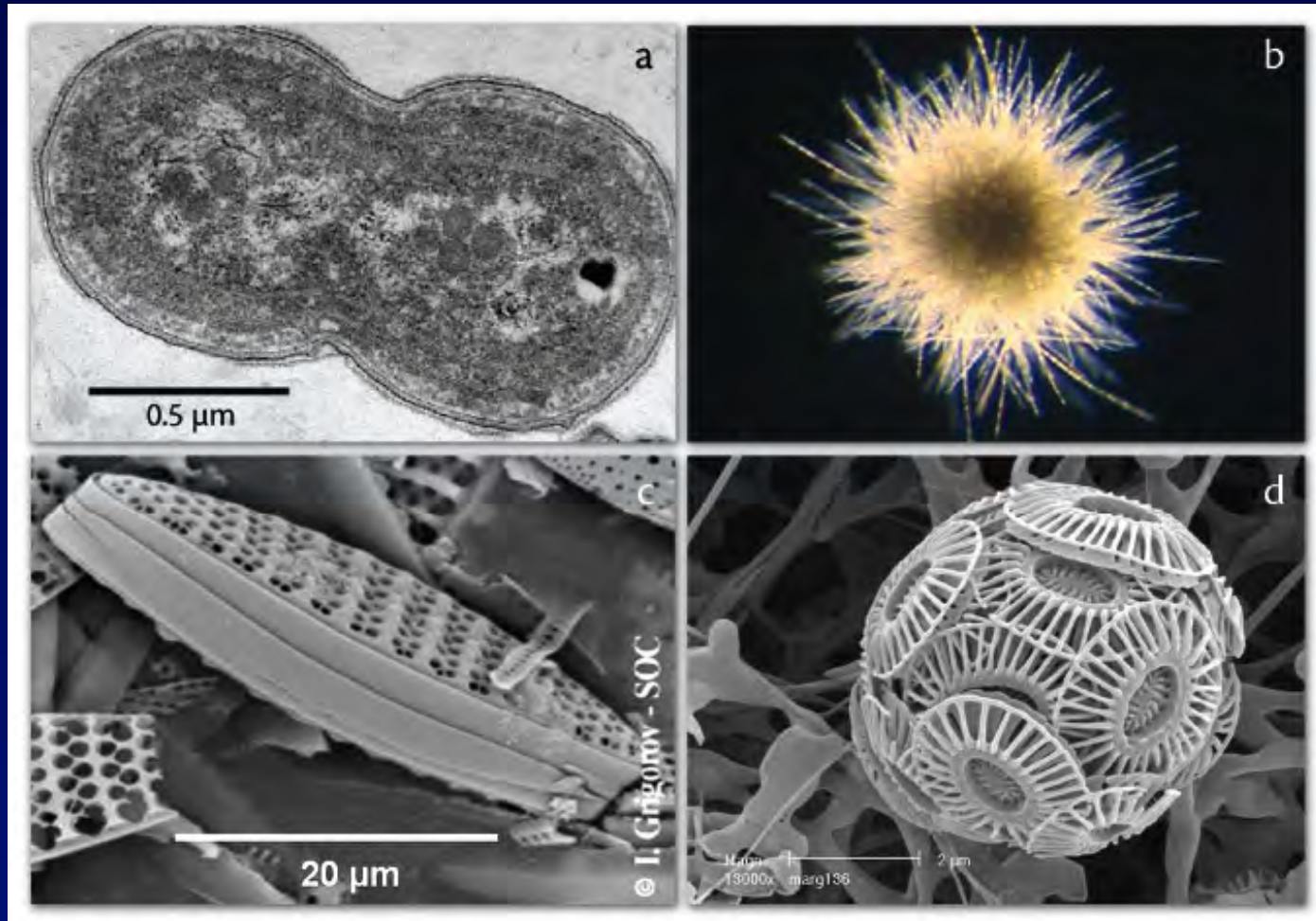
Blow up (viral lysis)

Sink

Die (e.g., apoptosis)

*A bit weird, because “population” refers to species, and many species are often lumped*

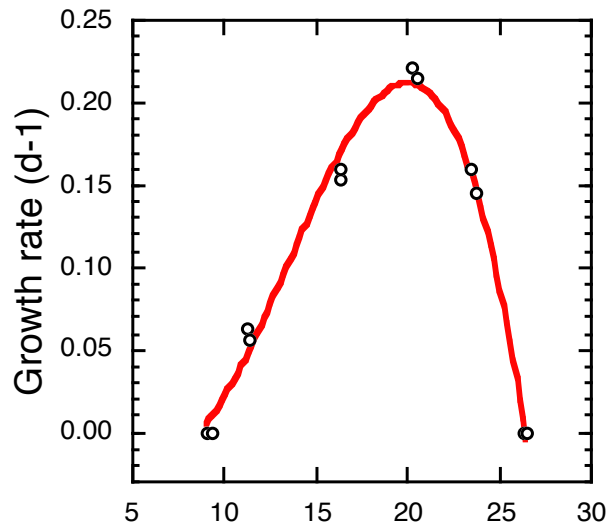
# Biogeochemical models must include functional groups



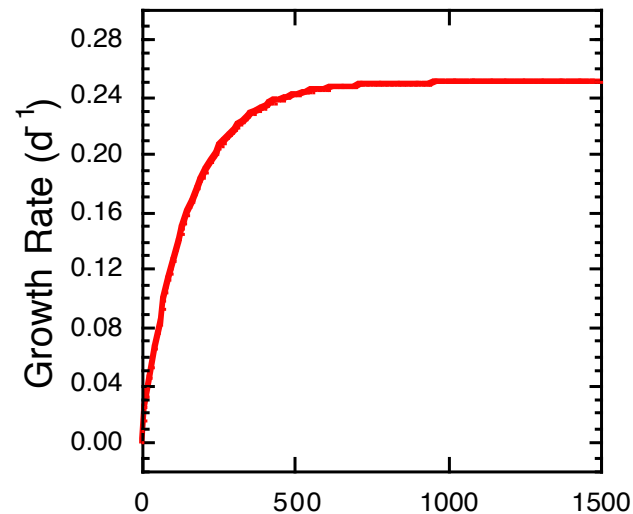
# Critical to know the Environmental Influences on the Growth of Phytoplankton:

Temperature  
Light  
Daylength  
Nutrients

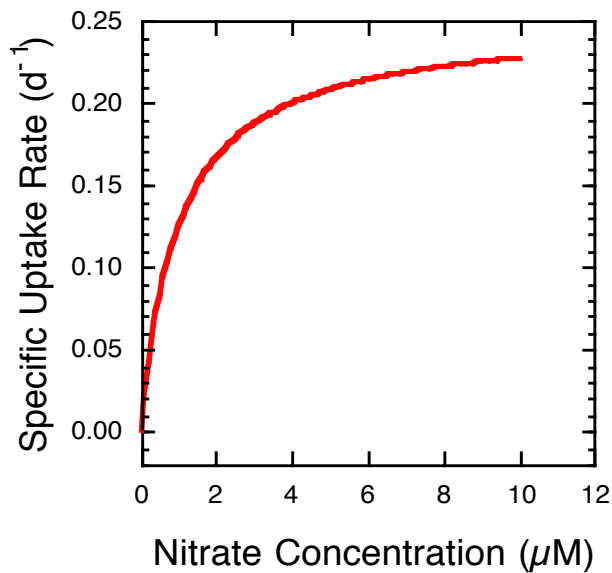
*Alexandrium ostenfeldii*



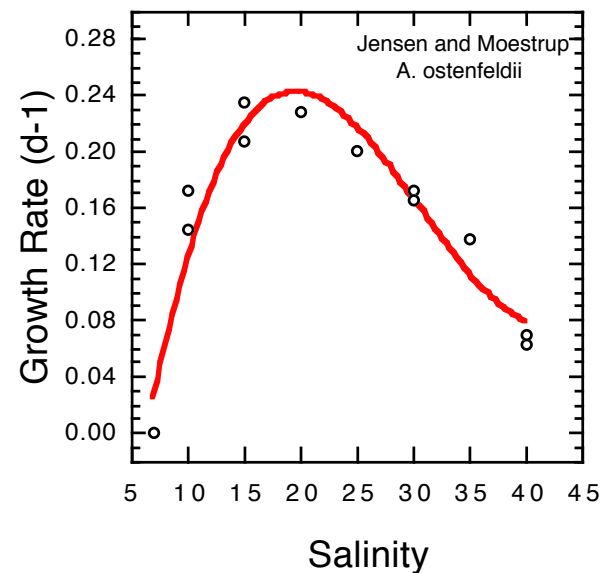
Growth vs Irradiance



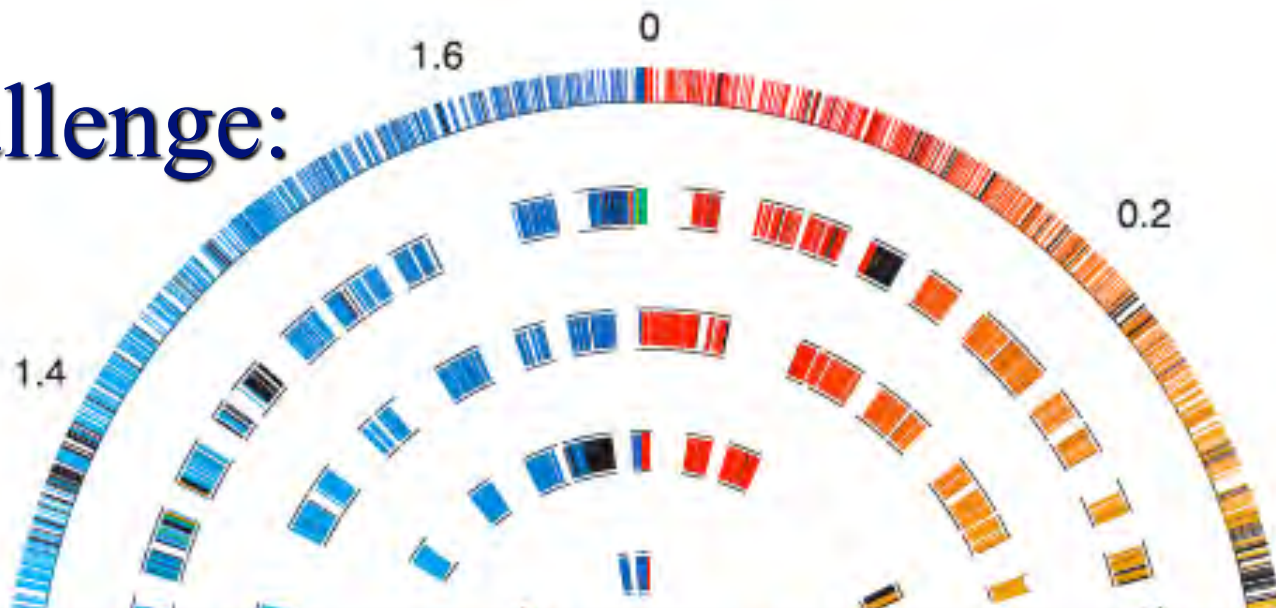
Nutrient Uptake Kinetics



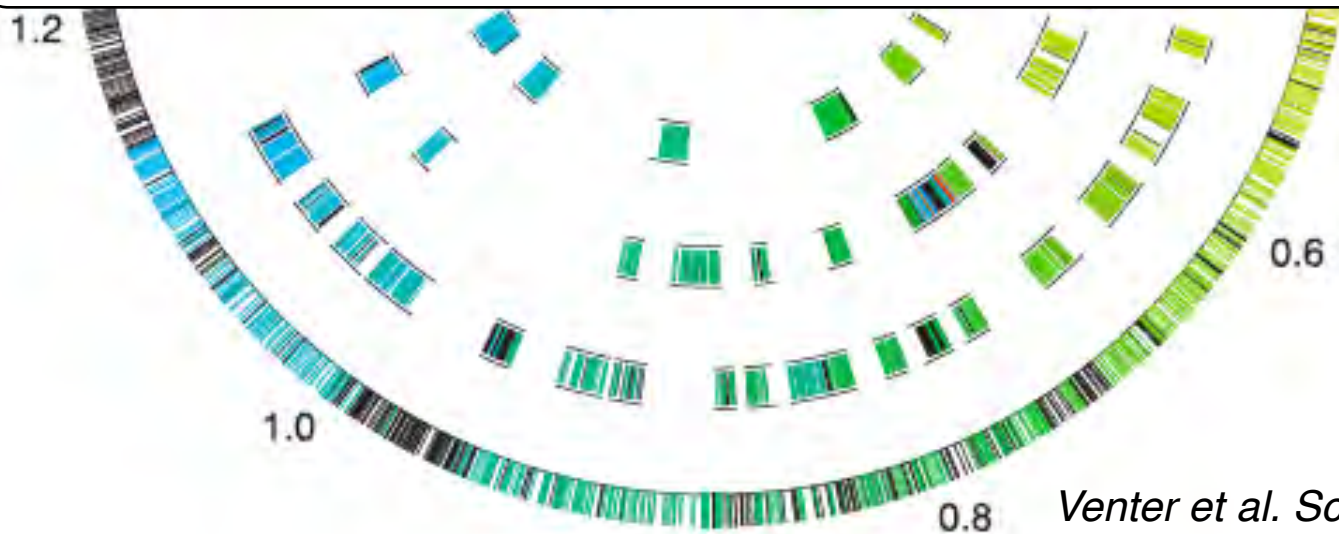
Effect of Salinity



# Challenge:



Figuring out what to do with exploding knowledge of biological complexity in the ocean



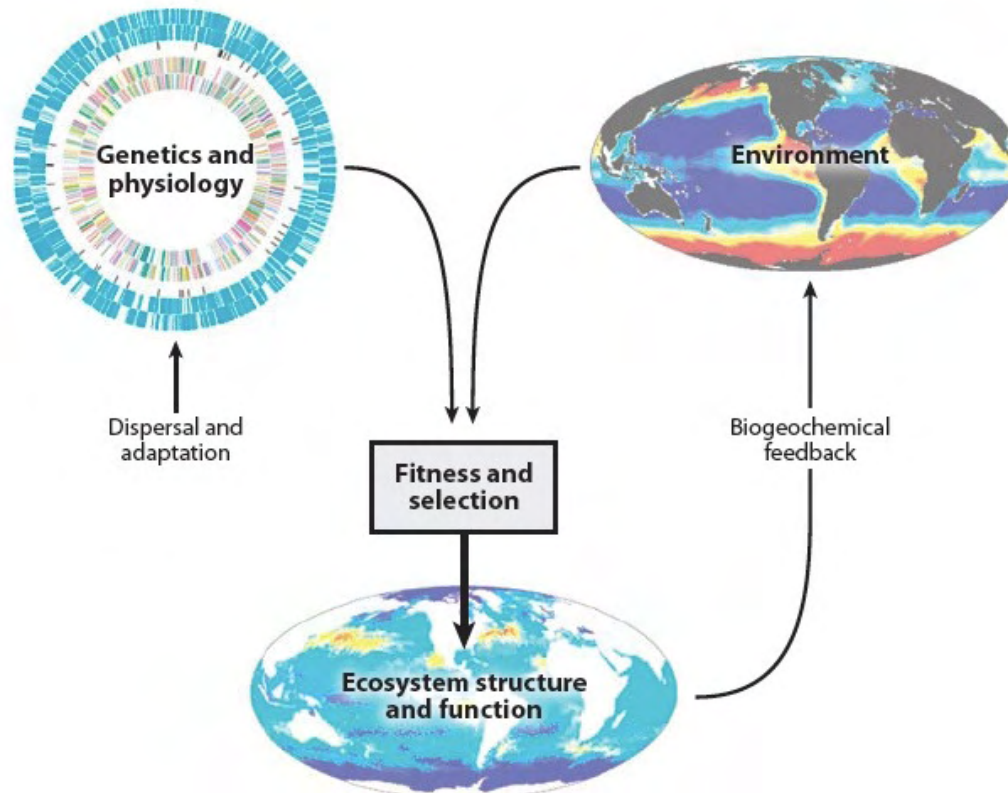


A big part of  
the answer is  
here

# Modeling Diverse Communities of Marine Microbes

Michael J. Follows and Stephanie Dutkiewicz

Earth, Atmospheric, and Planetary Sciences, Massachusetts Institute of Technology,  
Cambridge, Massachusetts 02139; email: mick@ocean.mit.edu, stephd@mit.edu



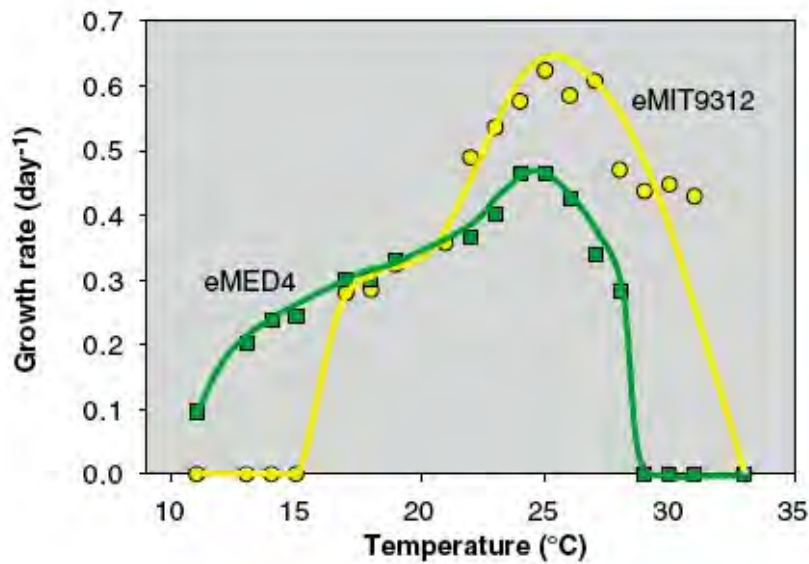
Annu. Rev. Mar. Sci. 2011. 3:427–51

The *Annual Review of Marine Science* is online at  
[marine.annualreviews.org](http://marine.annualreviews.org)

This article's doi:  
10.1146/annurev-marine-120709-142848

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1941-1405/11/0115-0427\$20.00

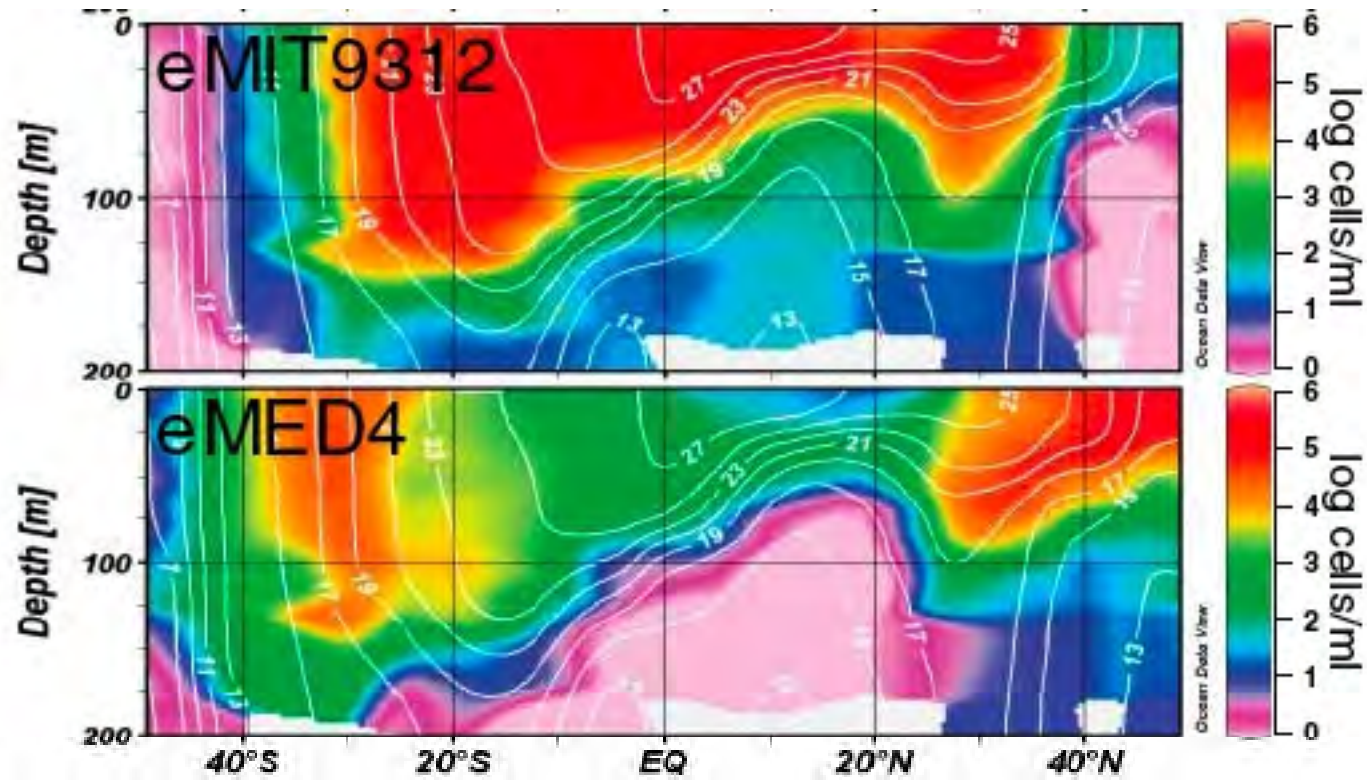


# Growth vs Temperature

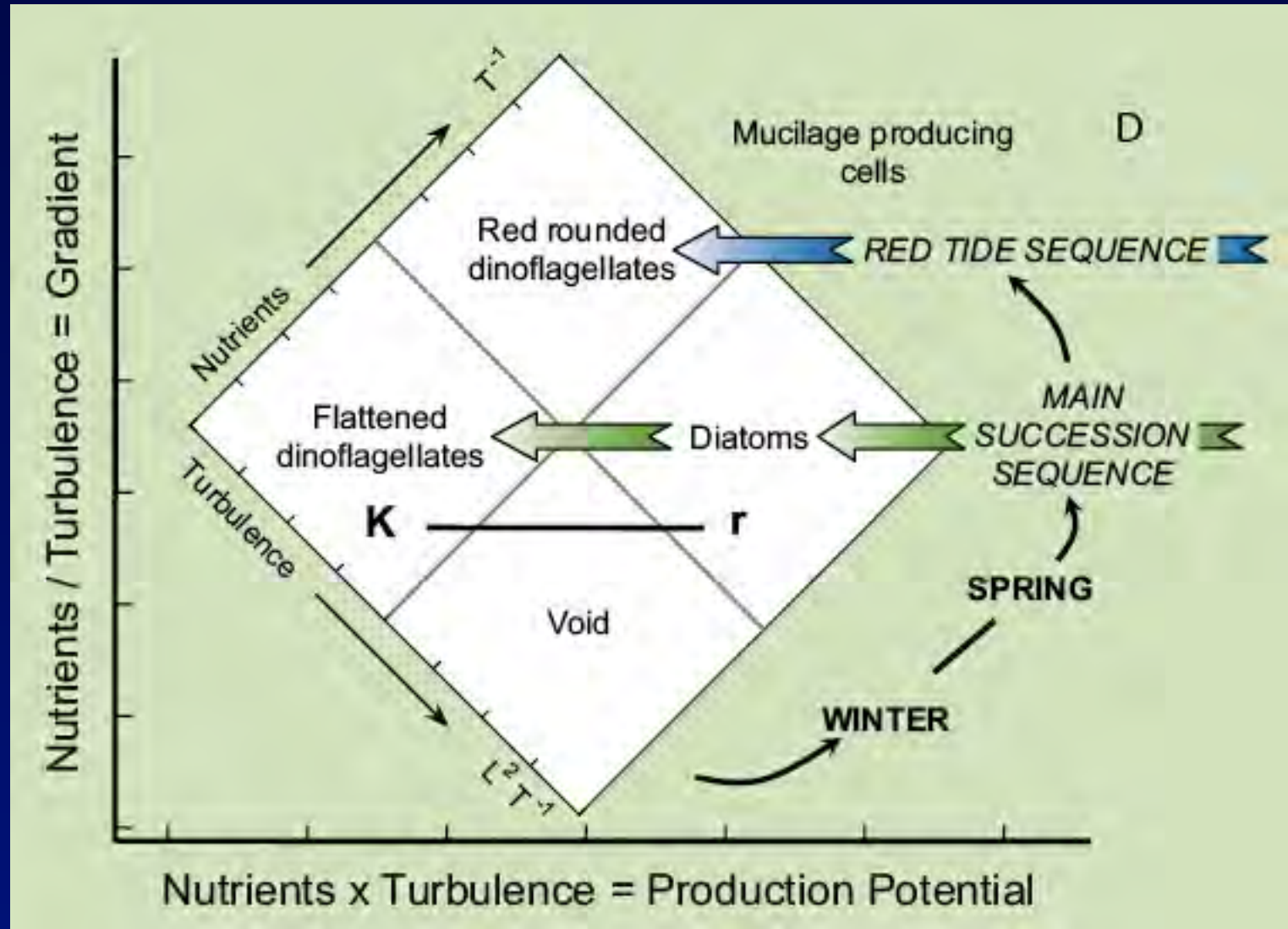
SCIENCE VOL 311 24 MARCH 2006 1737

## Niche Partitioning Among *Prochlorococcus* Ecotypes Along Ocean-Scale Environmental Gradients

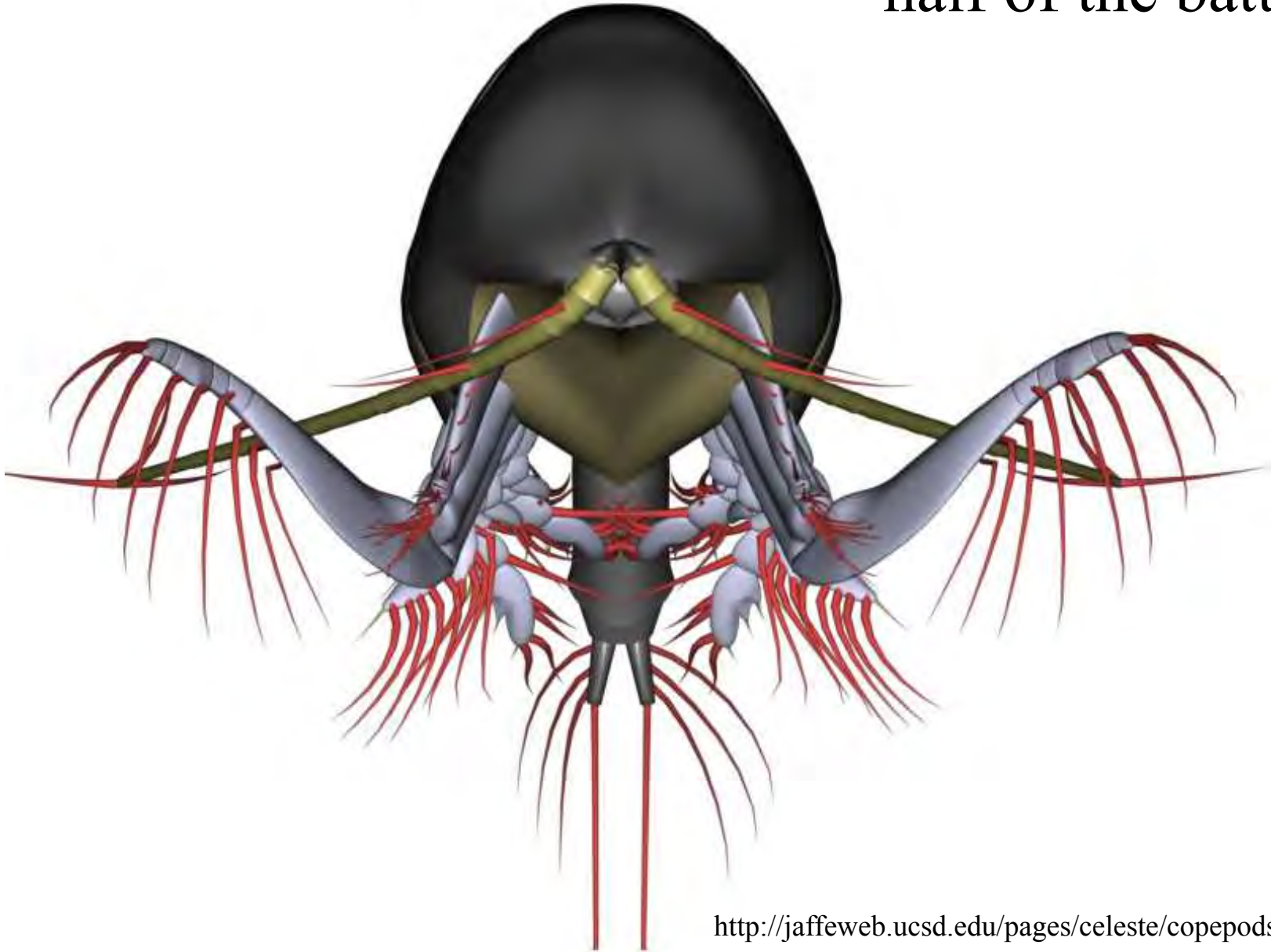
Zackary I. Johnson,<sup>1,2\*</sup> Erik R. Zinser,<sup>1,3\*</sup> Allison Coe,<sup>1</sup> Nathan P. McNulty,<sup>1</sup>  
E. Malcolm S. Woodward,<sup>4</sup> Sallie W. Chisholm<sup>1†</sup>



# Many other things define the growth-niche of marine phytoplankton



And cell division is only  
half of the battle!

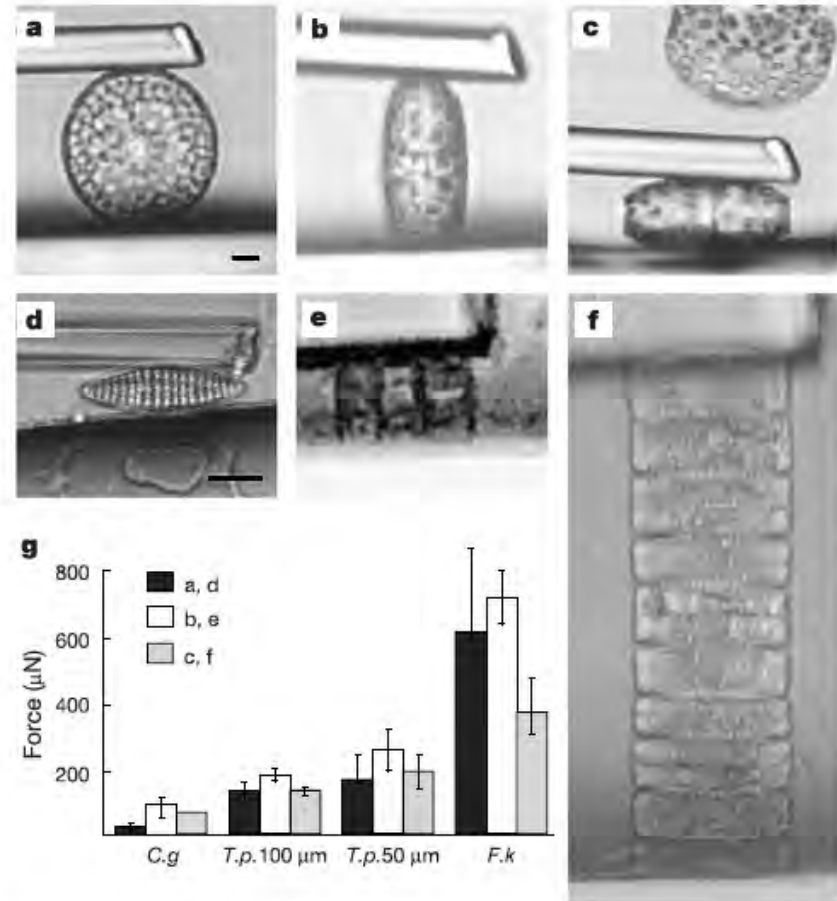
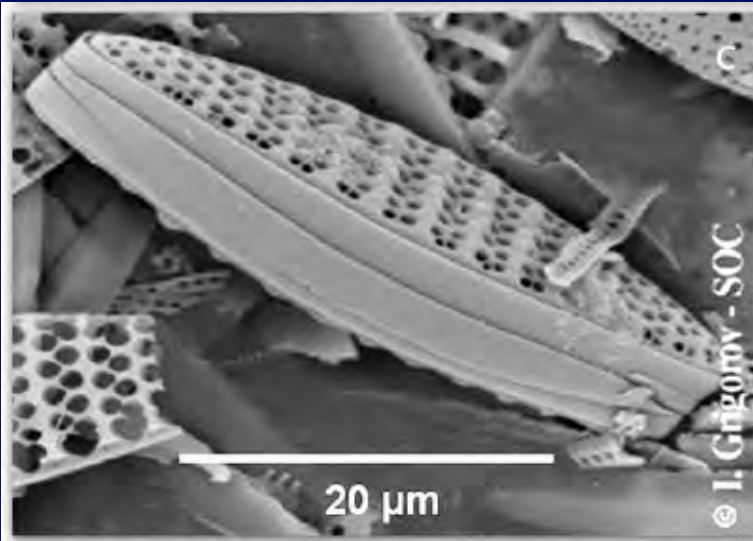


# Reduction of loss can be as good as an increase of growth rate

## Architecture and material properties of diatom shells provide effective mechanical protection

Christian E. Hamm\*, Rudolf Merkel†‡, Olaf Springer§, Piotr Jurkojc§, Christian Maier†, Kathrin Prechtel† & Victor Smetacek\*

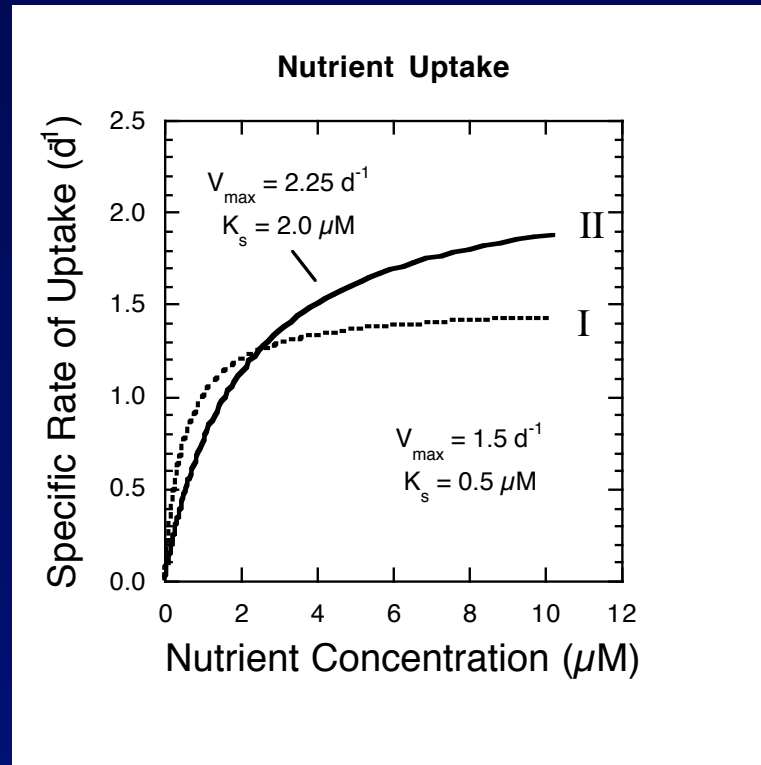
NATURE | VOL 421 | 20 FEBRUARY 2003 | www.nature.com/nature



**Figure 1** Glass needle tests: Live single cells of *T. punctigera* (a–c) and *F. kerguelensis* (d–f), in chains (e, f). Pressures applied along the girdle bands, (a, d), across the girdle bands (b, e), and across the centre of the valves (c, f). **g**, Forces necessary to break *Coscinodiscus granii* (*C.g.*), *Thalassiosira punctigera* (*T.p.*) with diameters of 100 and 50 μm, and *Fragilariopsis kerguelensis* (*F.k.*). *C. granii* has a geometry similar to that of *T. punctigera*. Scale bars, 10 μm.

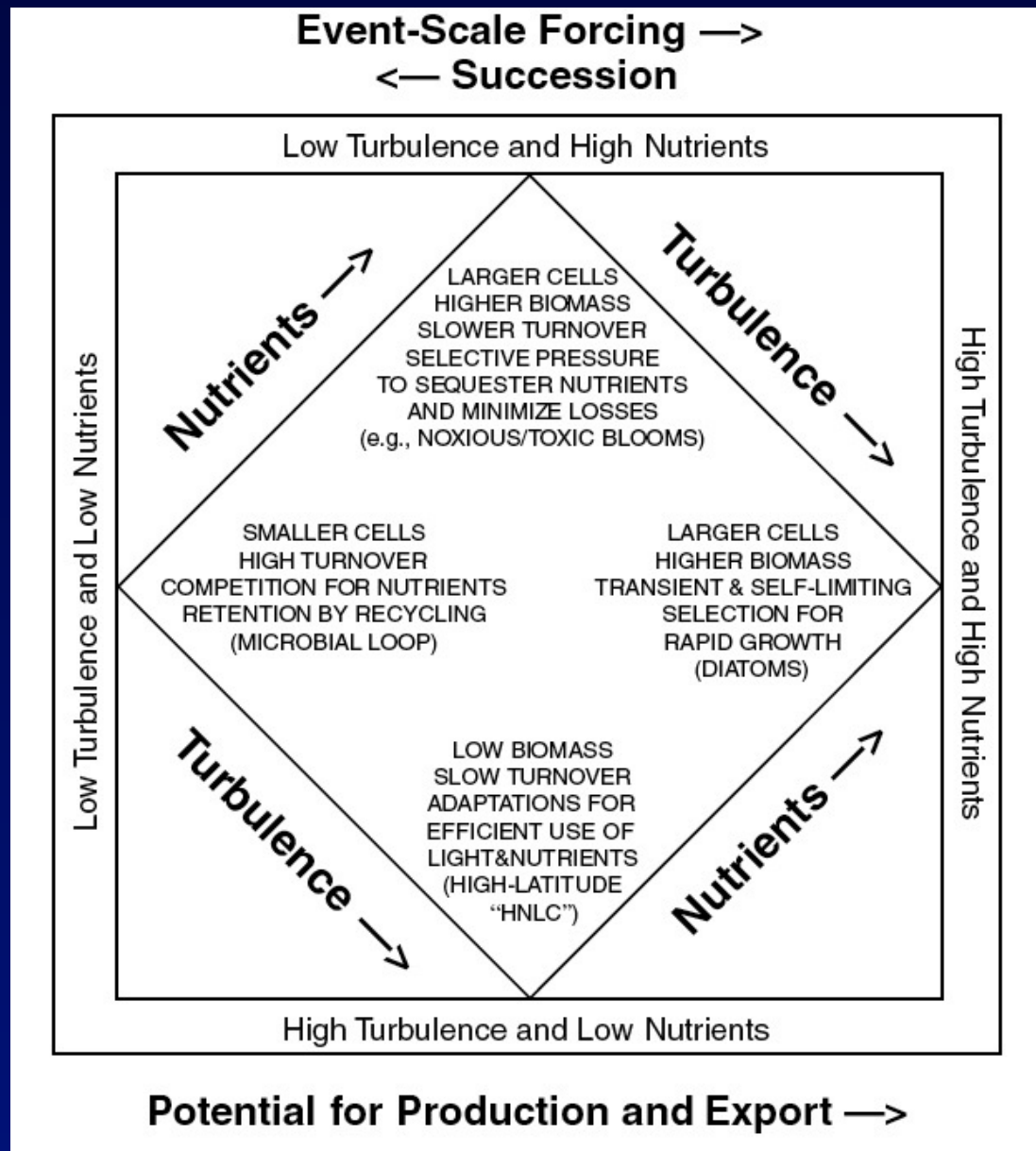
*It's the gene, stupid!*

So rapid growth is not the only strategy for survival / selection

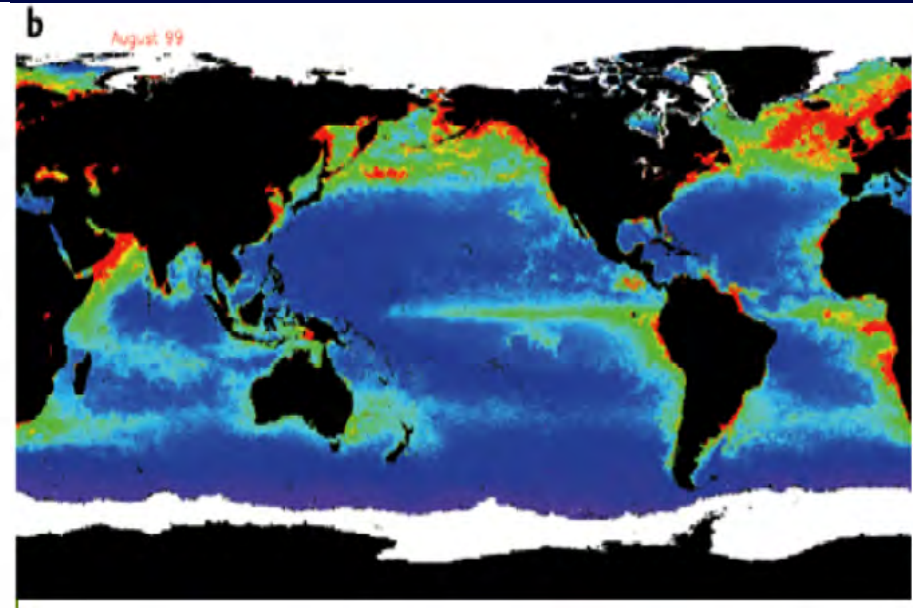
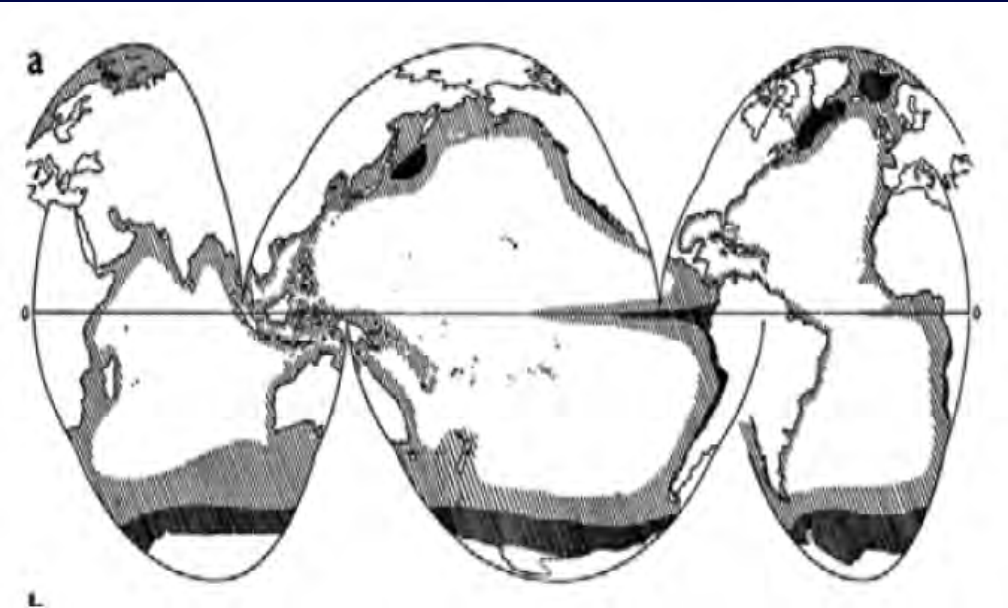


[www.andyslocum.com/images/tortoise&hare.jpg](http://www.andyslocum.com/images/tortoise&hare.jpg)

# Same framework



# Top-down or bottom-up control?

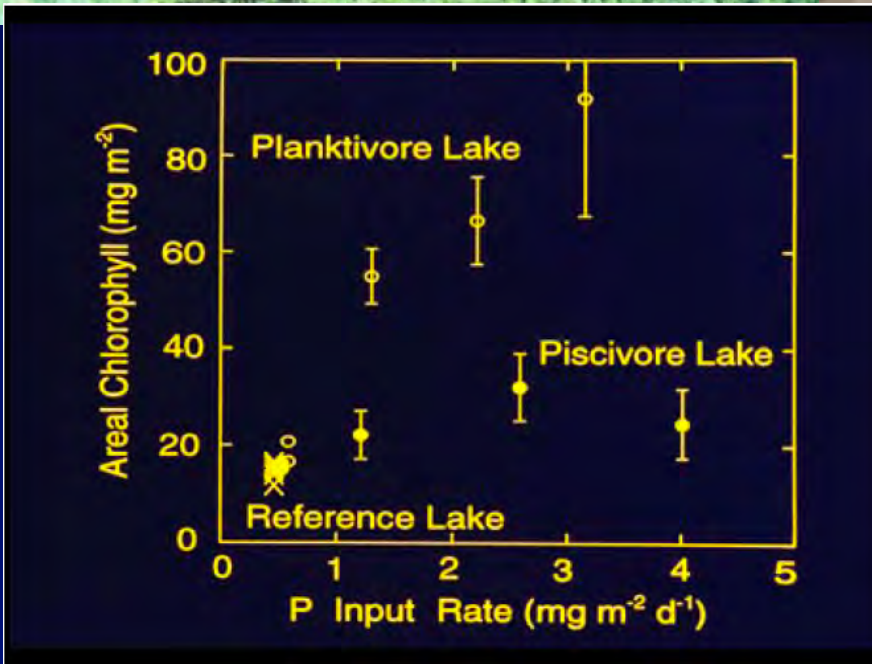
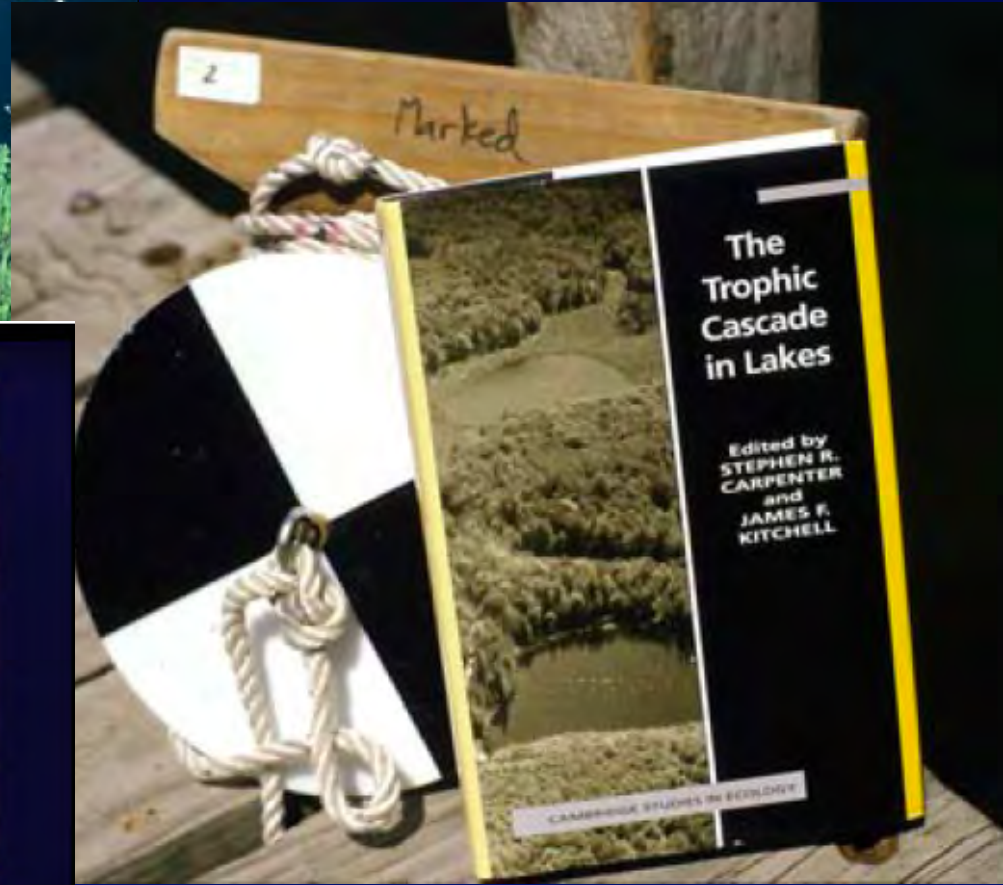


**Sverdrup's (1955) map of productivity based on vertical convection, upwelling and turbulent diffusion**

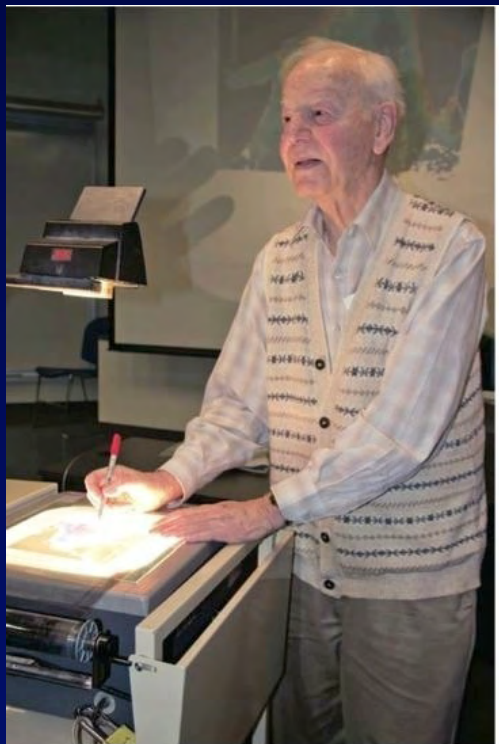
**Global productivity estimated from remote sensing (Falkowski et al. 1998)**



# Top-down control



# Top-down control



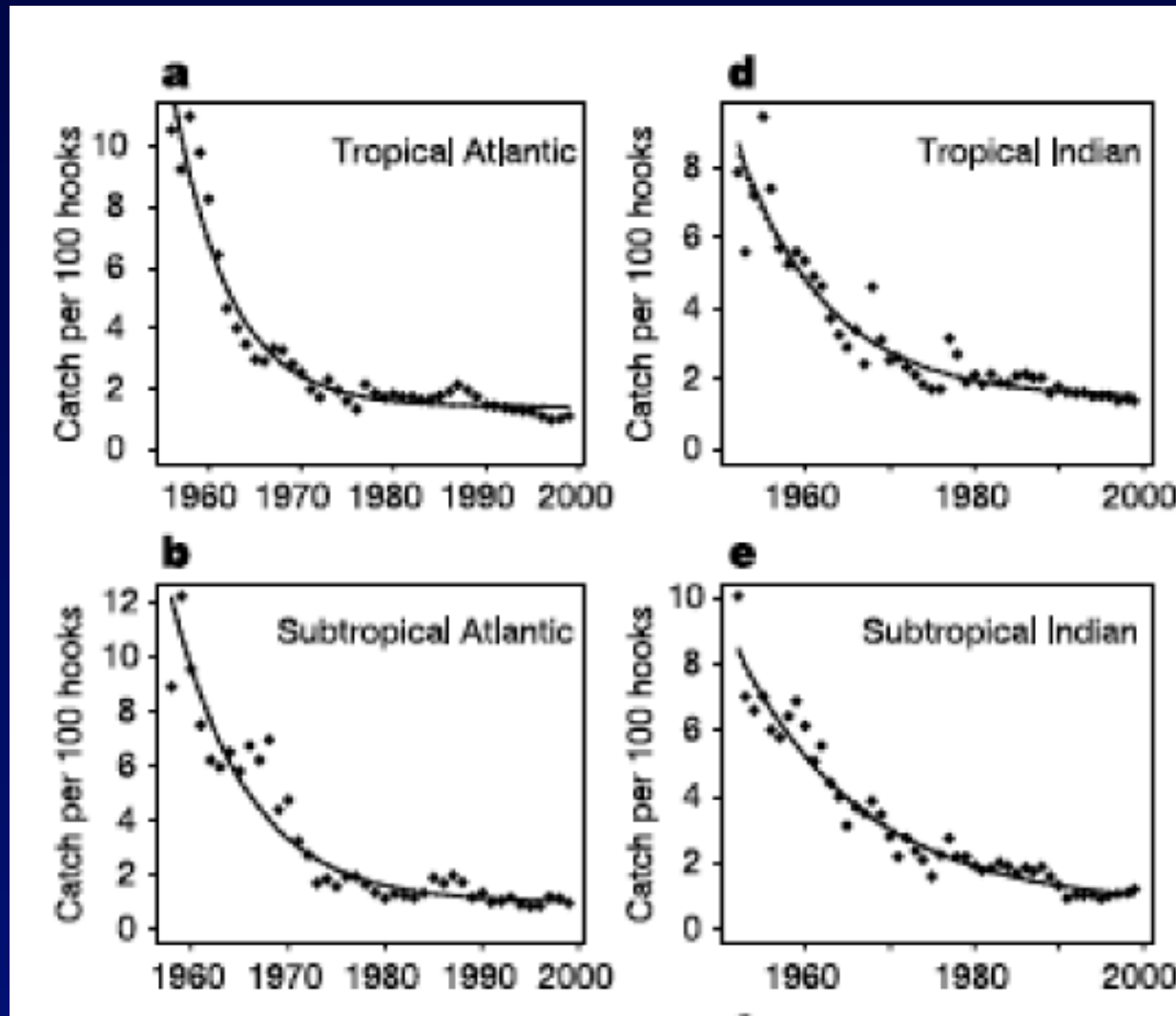
Karl Banse

## Reflections About Chance in My Career, and on the Top-Down Regulated World

Karl Banse

School of Oceanography, University of Washington, Seattle, Washington 98195-7940;  
email: banse@uw.edu

# Global test of the top-down hypothesis?



Decline of fish stocks since 1960

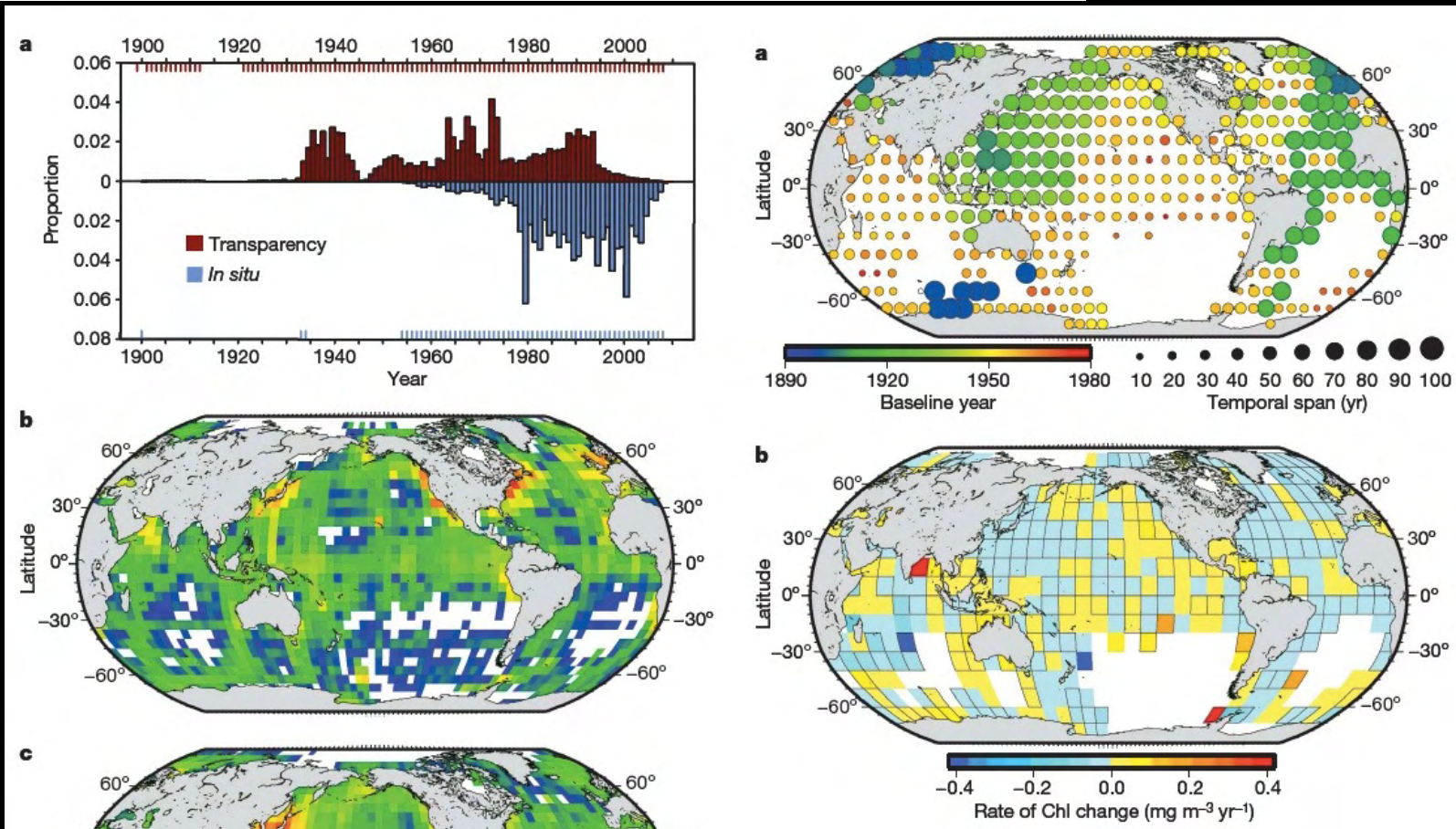
Myers and Worm Nature 2003

# Global phytoplankton decline over the past century

Daniel G. Boyce<sup>1</sup>, Marlon R. Lewis<sup>2</sup> & Boris Worm<sup>1</sup>

In the oceans, ubiquitous microscopic phototrophs (phytoplankton) account for approximately half the production of organic matter on Earth. Analyses of satellite-derived phytoplankton concentration (available since 1979) have suggested decadal-scale fluctuations linked to climate forcing, but the length of this record is insufficient to resolve longer-term trends. Here we combine available ocean transparency measurements and *in situ* chlorophyll observations to estimate the time

# Top-down effects?



# BRIEF COMMUNICATIONS ARISING

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## Does blending of chlorophyll data bias temporal trend?

David L. Mackas<sup>1</sup>

<sup>1</sup>Institute of Ocean Sciences, Fisheries and Oceans Canada, PO Box 6000, Sidney, British Columbia, V8L 4B2, Canada.

e-mail: Dave.Mackas@dfo-mpo.gc.ca

**ARISING FROM** D. G. Boyce, M. R. Lewis & B. Worm *Nature* **466**, 591–596 (2010)

## A measured look at ocean chlorophyll trends

Ryan R. Rykaczewski<sup>1,2</sup> & John P. Dunne<sup>2</sup>

<sup>1</sup>University Corporation for Atmospheric Research, Boulder, Colorado 80307-3000 USA.

## Is there a decline in marine phytoplankton?



Links to useful references

Abigail McQuatters-Gollop<sup>1</sup>, Philip C. Reid<sup>1</sup>, Martin Edwards<sup>1</sup>, Peter H. Burkill<sup>1</sup>, Claudia Castellani<sup>1</sup>, Sonia Batten<sup>1</sup>, Winfried Gieskes<sup>2</sup>, Doug Beare<sup>3</sup>, Robert R. Bidigare<sup>4</sup>, Erica Head<sup>5</sup>, Rod Johnson<sup>6</sup>, Mati Kahru<sup>7</sup>, J. Anthony Koslow<sup>7</sup> & Angelica Pena<sup>8</sup>

<sup>1</sup>Sir Alister Hardy Foundation for Ocean Science, The Laboratory, Citadel Hill, Plymouth, PL1 2PB, UK.

e-mail: abiqua@sahfos.ac.uk

<sup>2</sup>University of Groningen, ESRIG, Dept. Ocean Ecosystems, Nijenborgh 4,

# Many scientific controversies get sorted out in time

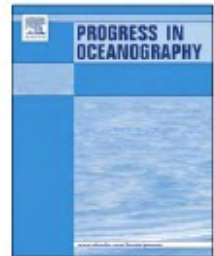
Progress in Oceanography 122 (2014) 163–173



Contents lists available at [ScienceDirect](#)

Progress in Oceanography

journal homepage: [www.elsevier.com/locate/pocean](http://www.elsevier.com/locate/pocean)



Estimating global chlorophyll changes over the past century

Daniel G. Boyce<sup>a,b,c,\*</sup>, Michael Dowd<sup>d</sup>, Marlon R. Lewis<sup>e</sup>, Boris Worm<sup>a</sup>



# Bottom-up processes

It can be argued that a similarly parsimonious set of factors determines the distribution of pelagic biomes, each with its characteristic flora and fauna... ***Copepods and whales do not determine which groups of plants will flourish; like the phytoplankton, they are themselves expressions of the regional physical oceanographic regime.***

(Alan Longhurst's section of Cullen et al., 2002, *The Sea* )

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Vol. 83: 291–303, 1992

MARINE ECOLOGY PROGRESS SERIES  
Mar. Ecol. Prog. Ser.

Published July 16

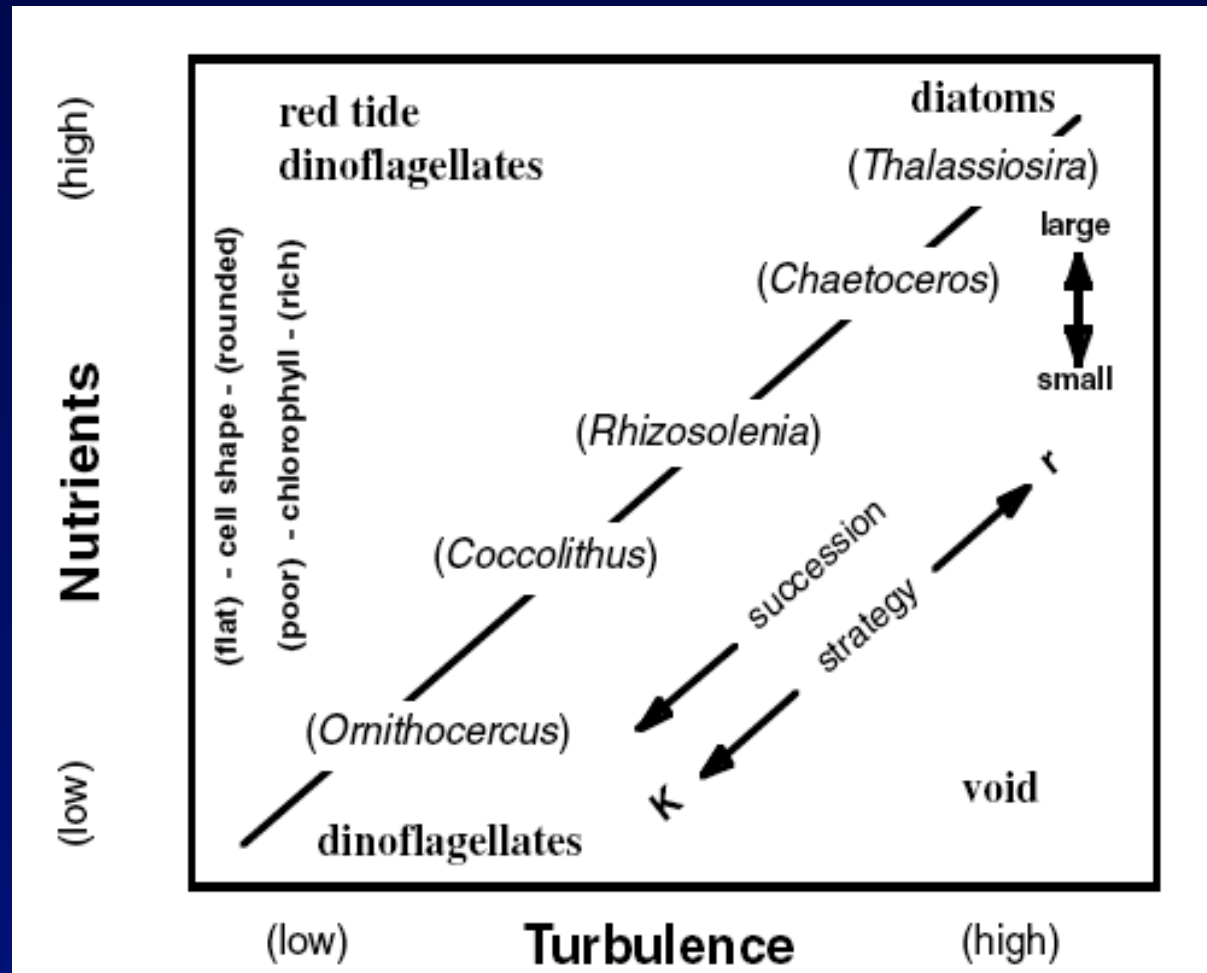
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**Grazing and iron limitation in the control of  
phytoplankton stock and nutrient concentration:  
a chemostat analogue of the Pacific equatorial  
upwelling zone**

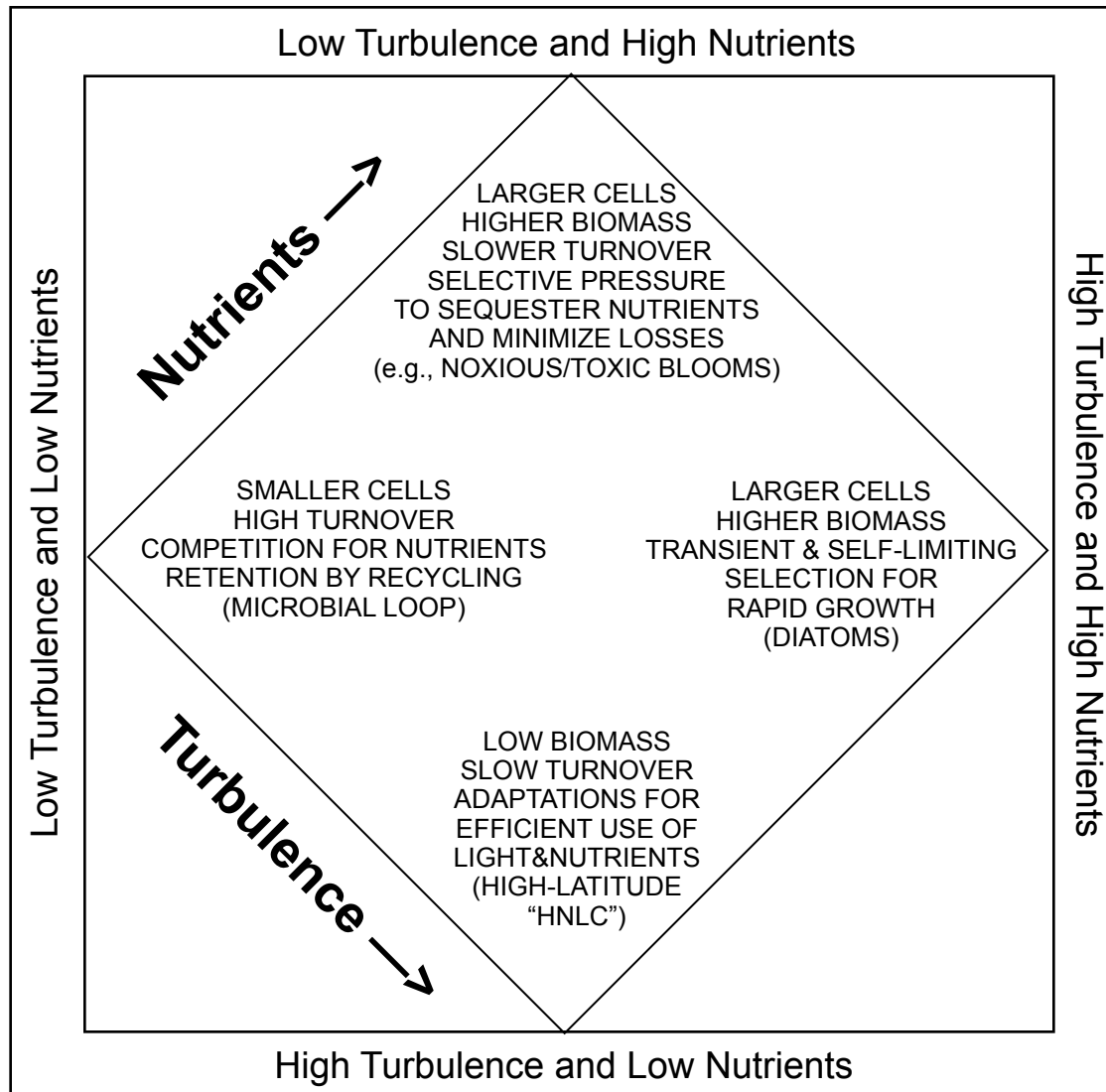
B. W. Frost, N. C. Franzen



# A Tool for Making Sense of Physically Forced Ecosystem Dynamics: Margalef's Mandala



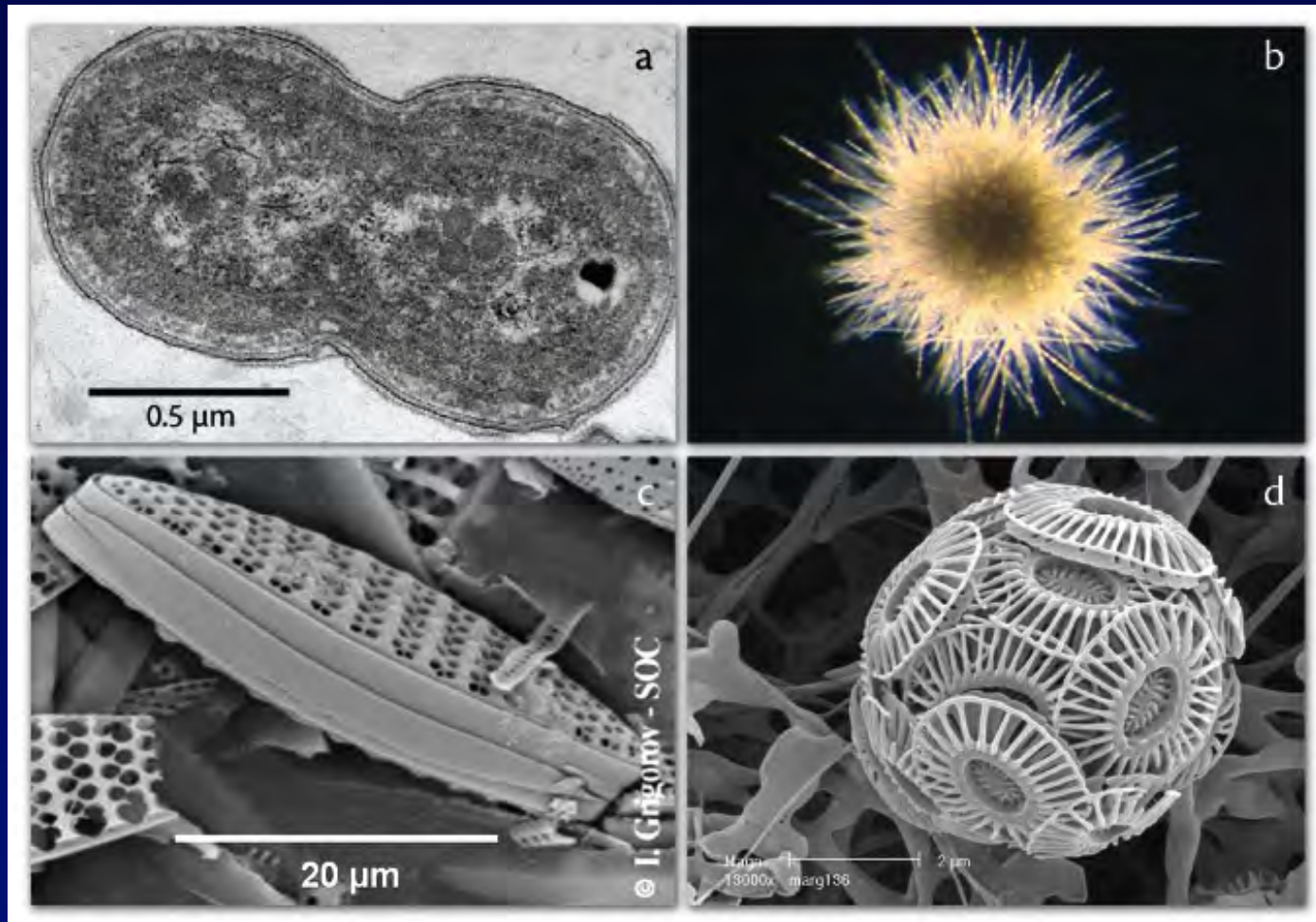
**Event-Scale Forcing →**  
**← Succession**



**Potential for Production and Export →**

**Cullen et  
al. 2002,  
The Sea**

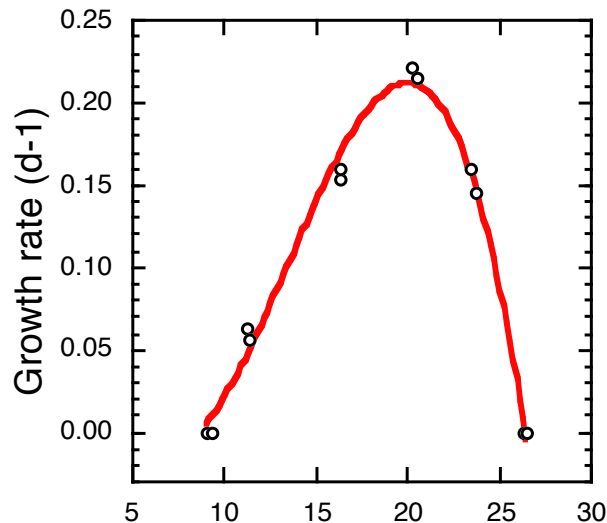
# The challenge: quantitative description of the niches of functional groups



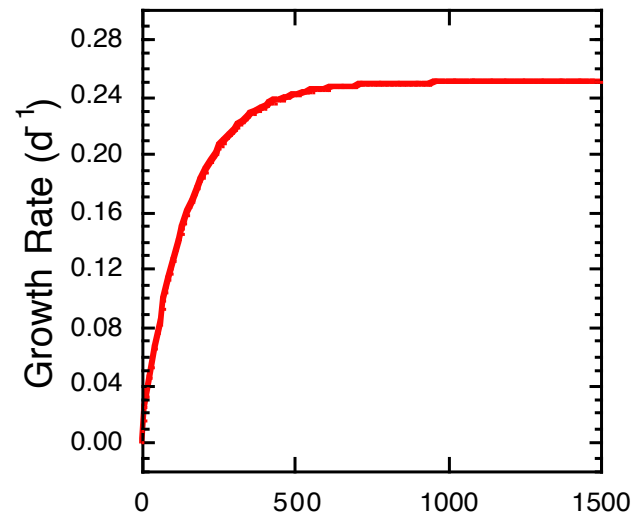
# Critical to know the Environmental Influences on the Growth of Phytoplankton:

Temperature  
Light  
Daylength  
Nutrients

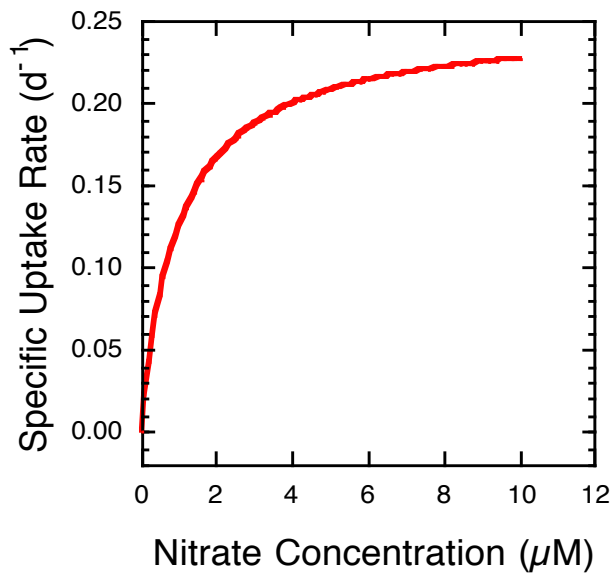
*Alexandrium ostenfeldii*



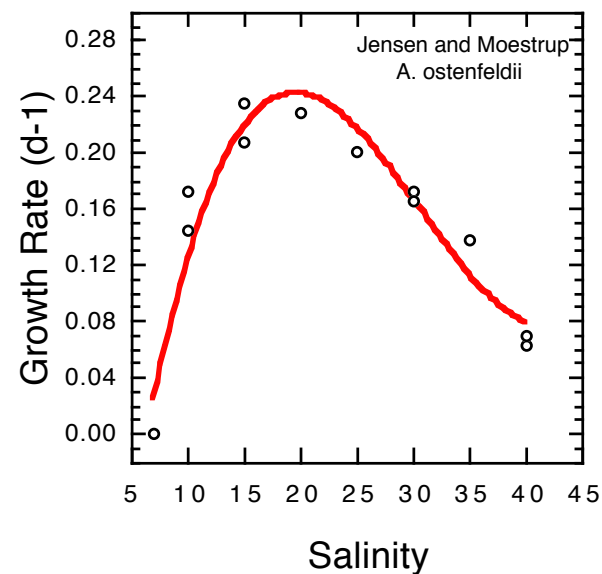
Growth vs Irradiance



Nutrient Uptake Kinetics



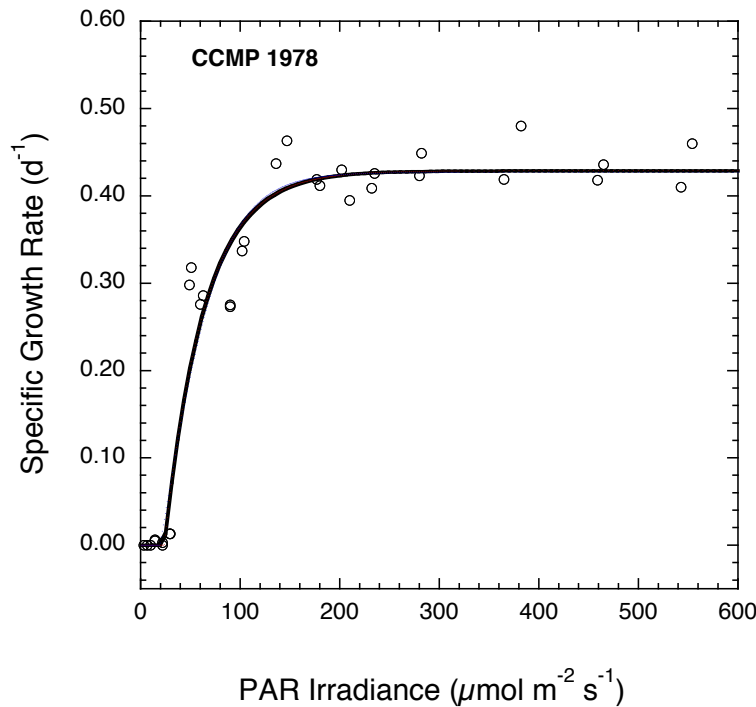
Effect of Salinity



Ecosystem modeling ground zero:

# Acclimated growth rate: genotypic

Cullen et al. in prep



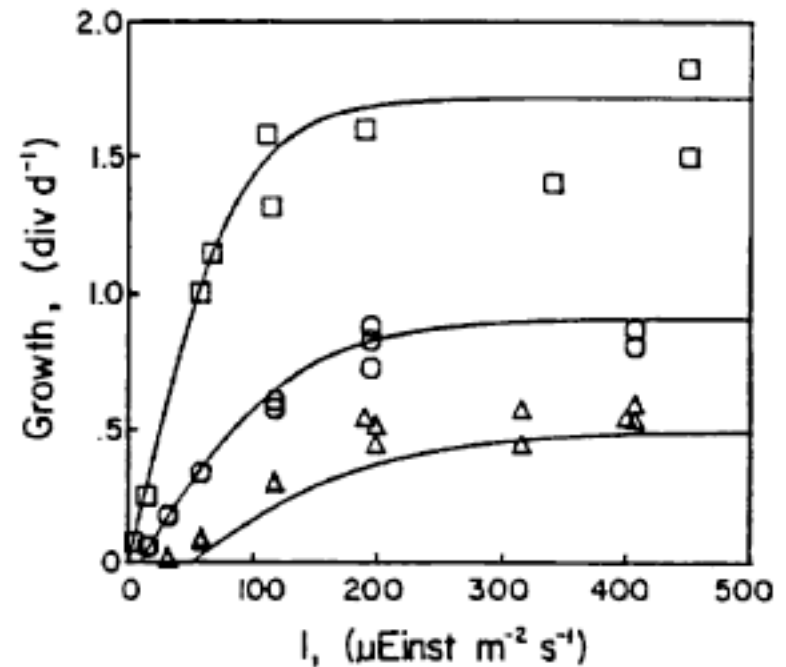
$$\mu(E) = (\mu_{\max}) \left(1 - e^{-\frac{(E - K_C)}{(K_E - K_C)}}\right) \quad \text{if } E > K_C$$
$$\mu(E) = 0 \quad \text{if } E \leq K_C$$

Species evolve different functions through **adaptation** and selection

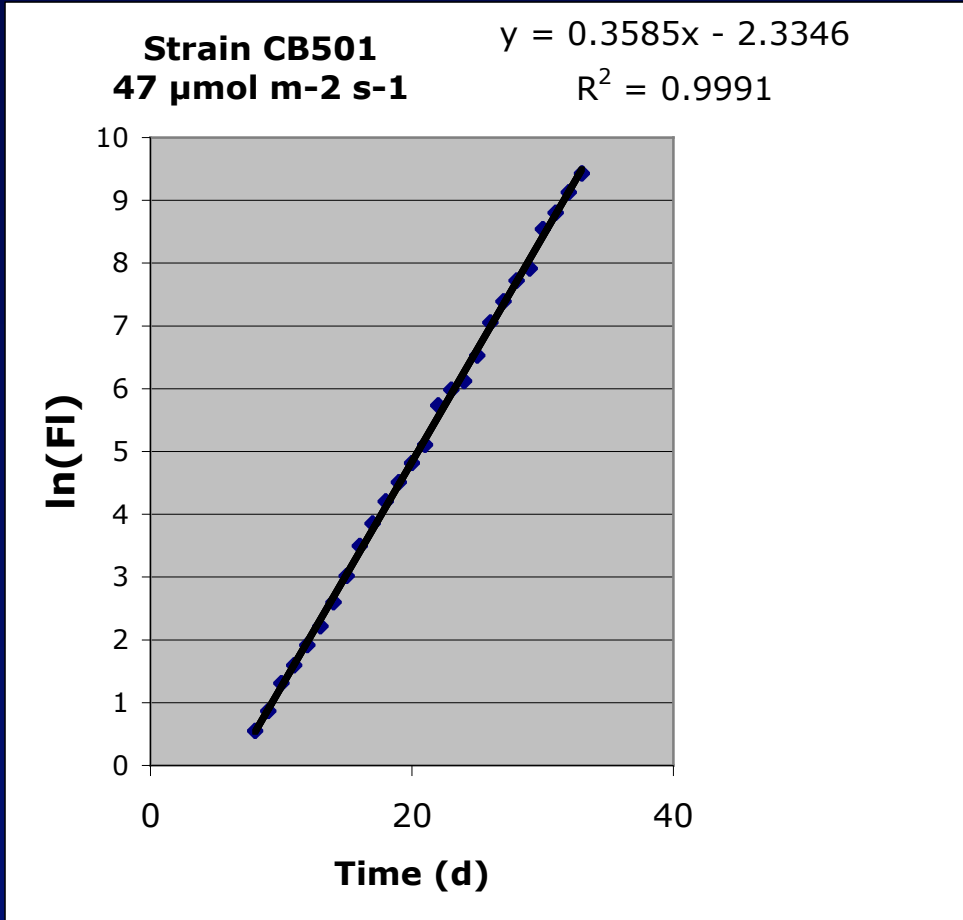
Journal of Plankton Research Vol.9 no.3 pp.459-482, 1987

On the causes of interspecific differences in the growth-irradiance relationship for phytoplankton. Part I. A comparative study of the growth-irradiance relationship of three marine phytoplankton species: *Skeletonema costatum*, *Olisthodiscus luteus* and *Gonyaulax tamarensis*

Christopher Langdon<sup>1</sup>



# $\mu$ as a function of $T$ and $E$ may not be relevant for describing growth rate in dynamic environments



## *Alexandrium fundyense*

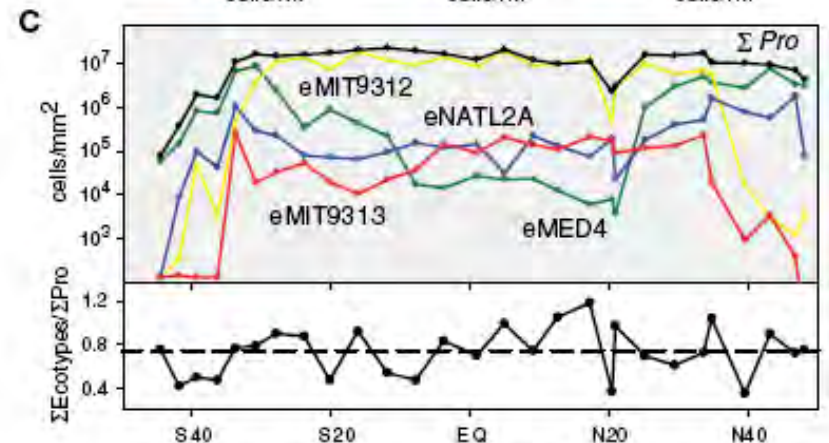
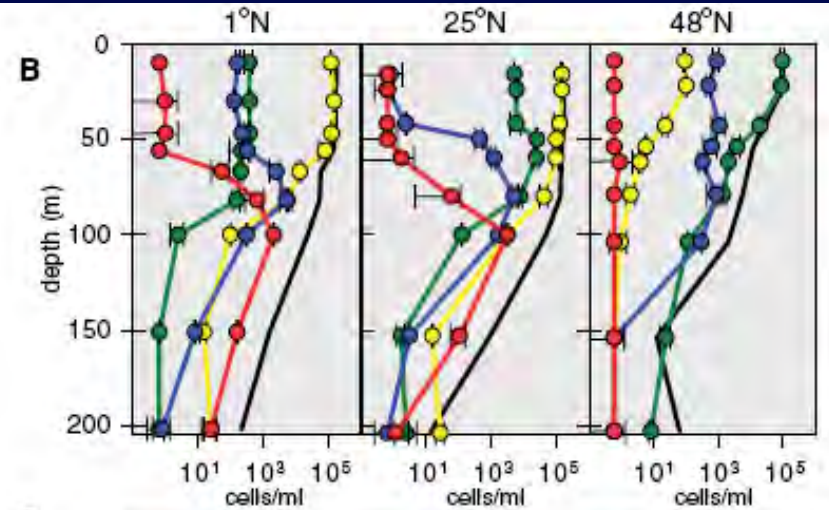
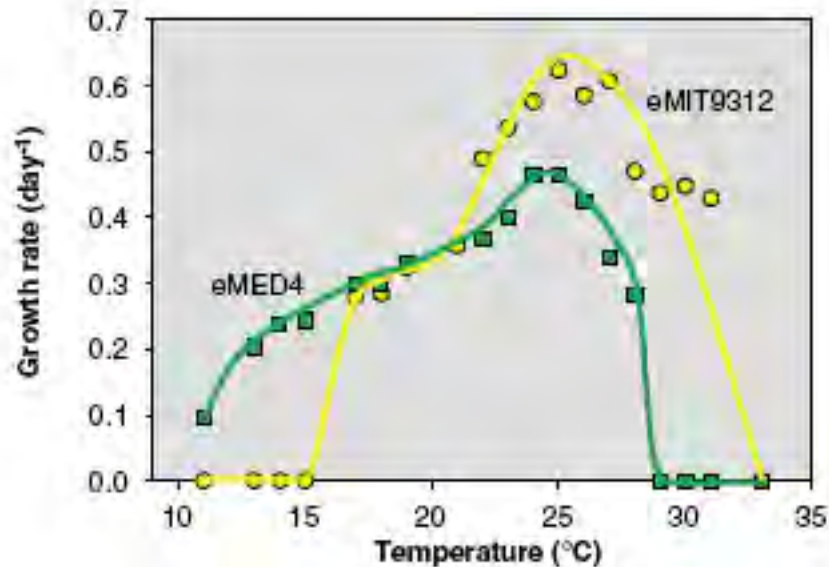
Growth determined using the method of Brand and Guillard

Brand, L. E. and Guillard, R. R. L. (1981). A method for the rapid and precise determination of acclimated phytoplankton reproduction rates. *J. Plankton Res.* 3: 191-201.

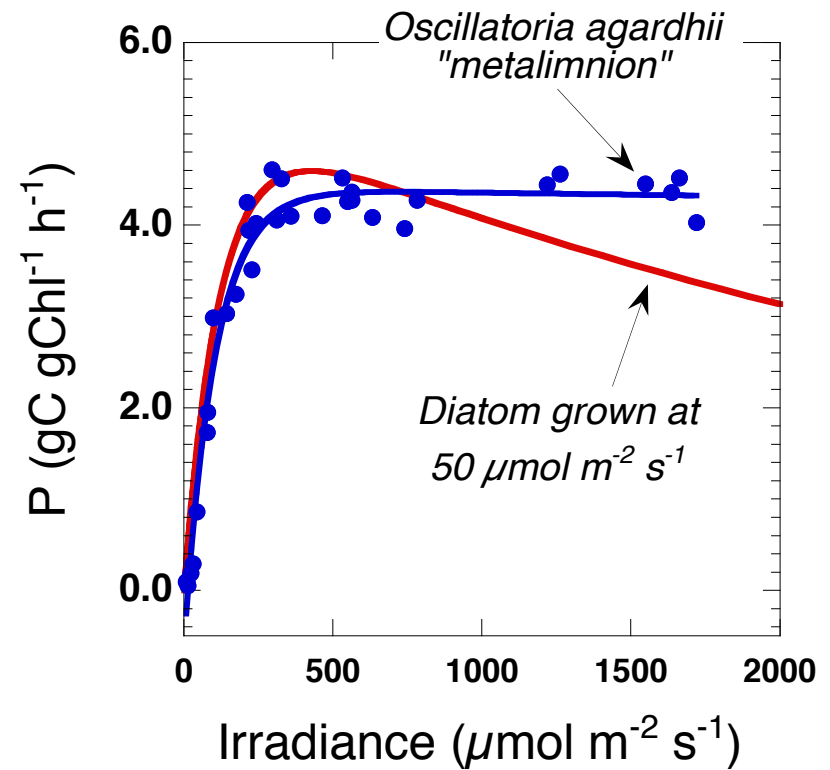
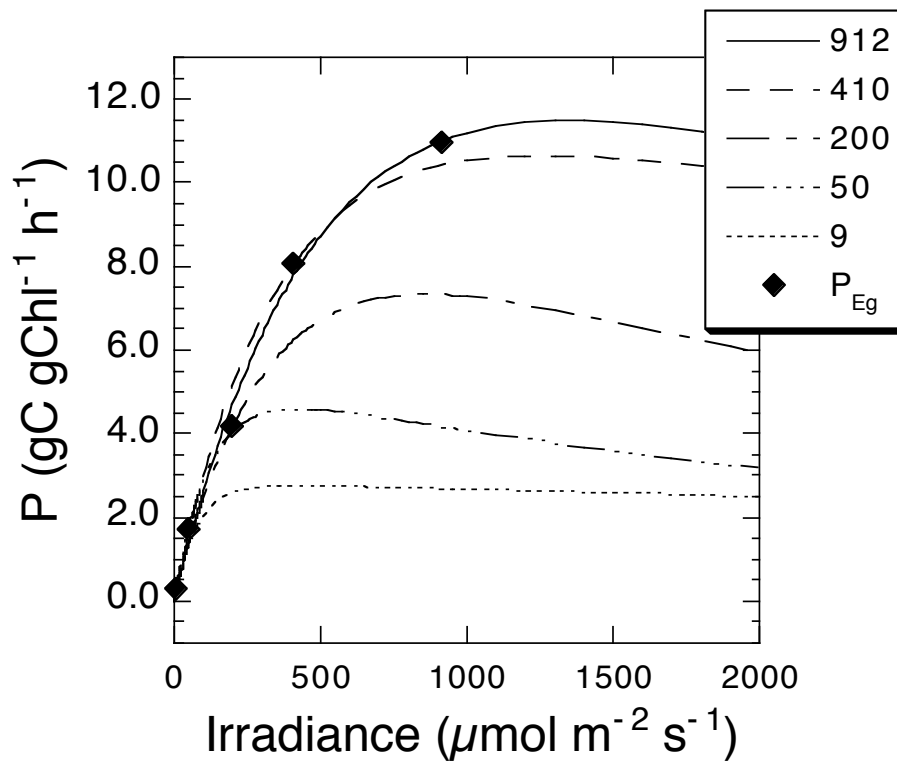
# But it is an excellent start for identifying niches

## Niche Partitioning Among *Prochlorococcus* Ecotypes Along Ocean-Scale Environmental Gradients

Zackary I. Johnson,<sup>1,2\*</sup> Erik R. Zinser,<sup>1,3\*</sup> Allison Coe,<sup>1</sup> Nathan P. McNulty,<sup>1</sup>  
E. Malcolm S. Woodward,<sup>4</sup> Sallie W. Chisholm<sup>1†</sup>



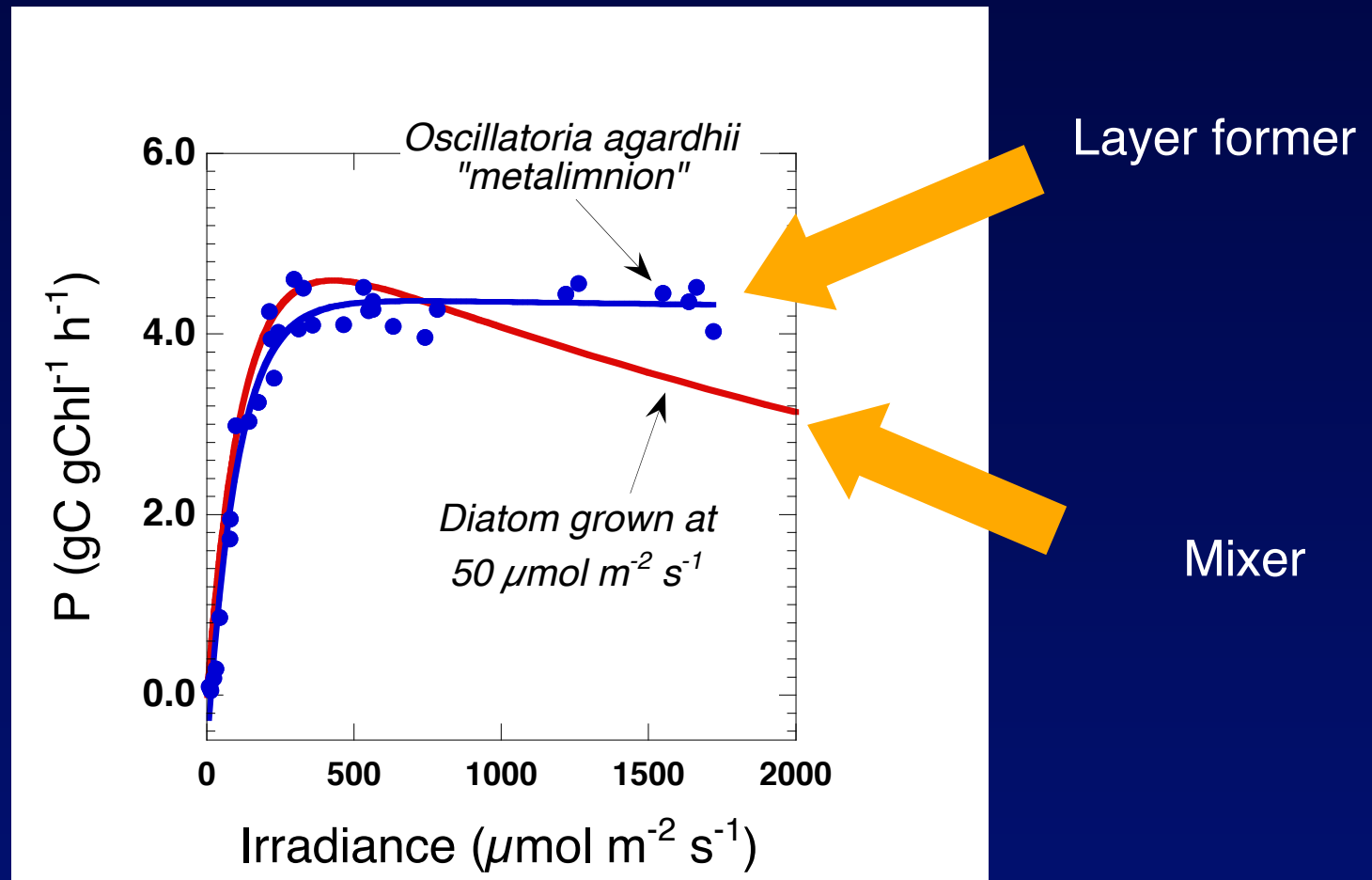
# Photosynthesis vs Irradiance: phenotypic



Species develop different functions through **acclimation**



# Photosynthesis vs Irradiance: phenotypic



Strategies to respond to environmental variability are adaptations

# Adaptations to oceanic vs coastal conditions

Vol. 41: 275–282, 1987

MARINE ECOLOGY – PROGRESS SERIES  
Mar. Ecol. Prog. Ser.

Published December 14

## *Thalassiosira oceanica* and *T. pseudonana*: two different photoadaptational responses

E. Sakshaug<sup>1</sup>, S. Demers<sup>2</sup>, C. M. Yentsch<sup>3</sup>

## Photosynthetic architecture differs in coastal and oceanic diatoms

Robert F. Strzepek<sup>1+</sup> & Paul J. Harrison<sup>2+</sup>

NATURE | VOL 431 | 7 OCTOBER 2004 | www.nature.com/nature

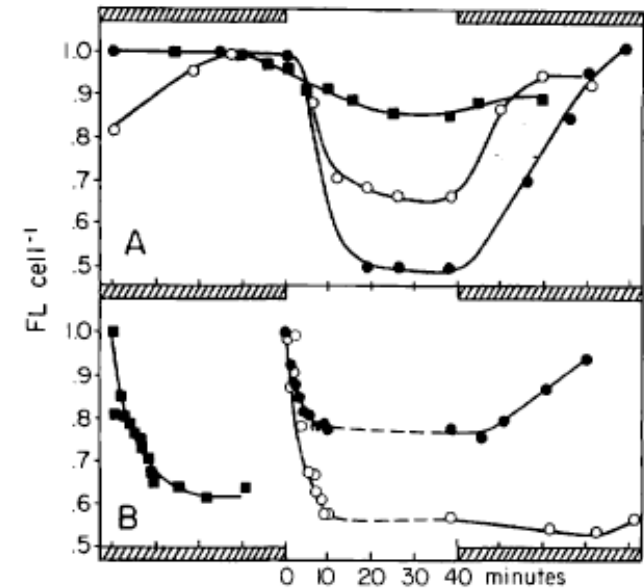


Fig. 5. *Thalassiosira pseudonana* and *T. oceanica*. Short-term changes of *in vivo* fluorescence per cell normalized to the highest observed value in each experiment. (A) *T. pseudonana* at 600 (○) or 800  $\mu\text{mol m}^{-2} \text{s}^{-1}$  (●) exposed to 2800, and then to 600 or 800  $\mu\text{mol m}^{-2} \text{s}^{-1}$  and *T. oceanica* (■) at 800  $\mu\text{mol m}^{-2} \text{s}^{-1}$  exposed to 2800 and then to 800  $\mu\text{mol m}^{-2} \text{s}^{-1}$ . (B) Changes in *in vivo* fluorescence per cell of *T. pseudonana* after addition of gramicidin; (■) cultures at 800  $\mu\text{mol m}^{-2} \text{s}^{-1}$ ; (○) cultures at 2800  $\mu\text{mol m}^{-2} \text{s}^{-1}$  followed by darkness; (●) control (no gramicidin), cultures at 800  $\mu\text{mol m}^{-2} \text{s}^{-1}$  followed by darkness. Horizontal bars: period of low light or darkness according to descriptions above

# Oceanic species has much reduced capability for regulation

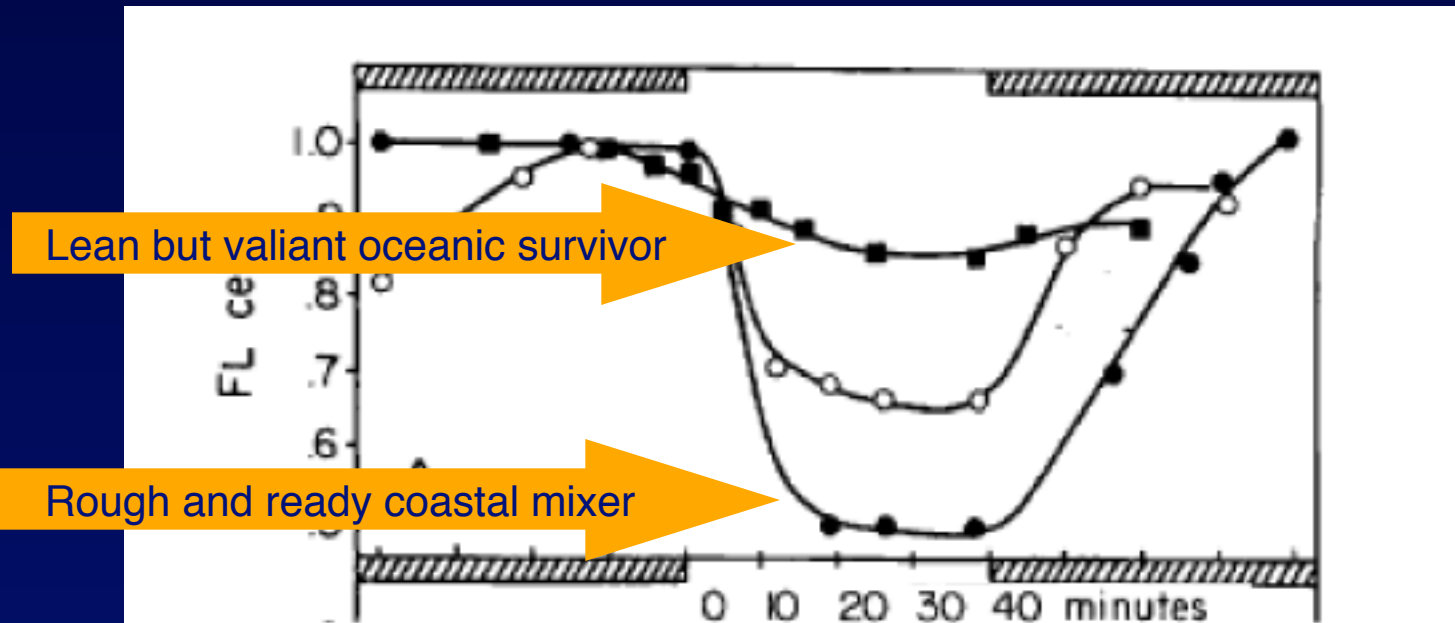
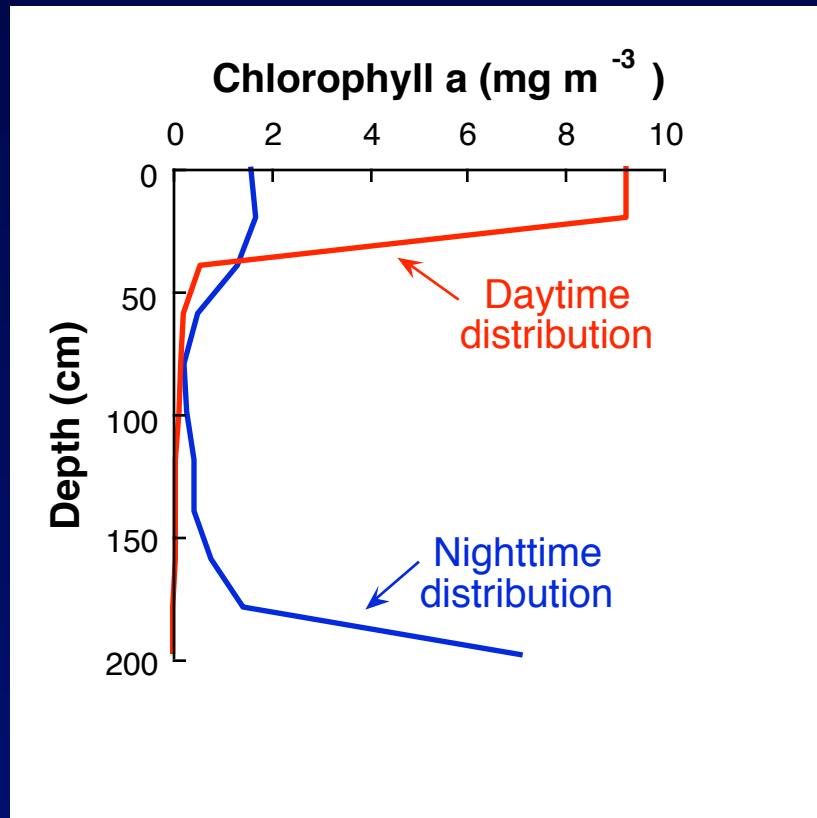


Fig. 5. *Thalassiosira pseudonana* and *T. oceanica*. Short-term changes of *in vivo* fluorescence per cell normalized to the highest observed value in each experiment. (A) *T. pseudonana* at 600 (○) or 800  $\mu\text{mol m}^{-2} \text{s}^{-1}$  (●) exposed to 2800, and then to 600 or 800  $\mu\text{mol m}^{-2} \text{s}^{-1}$  and *T. oceanica* (■) at 800  $\mu\text{mol m}^{-2} \text{s}^{-1}$  exposed to 2800 and then to 800  $\mu\text{mol m}^{-2} \text{s}^{-1}$ .

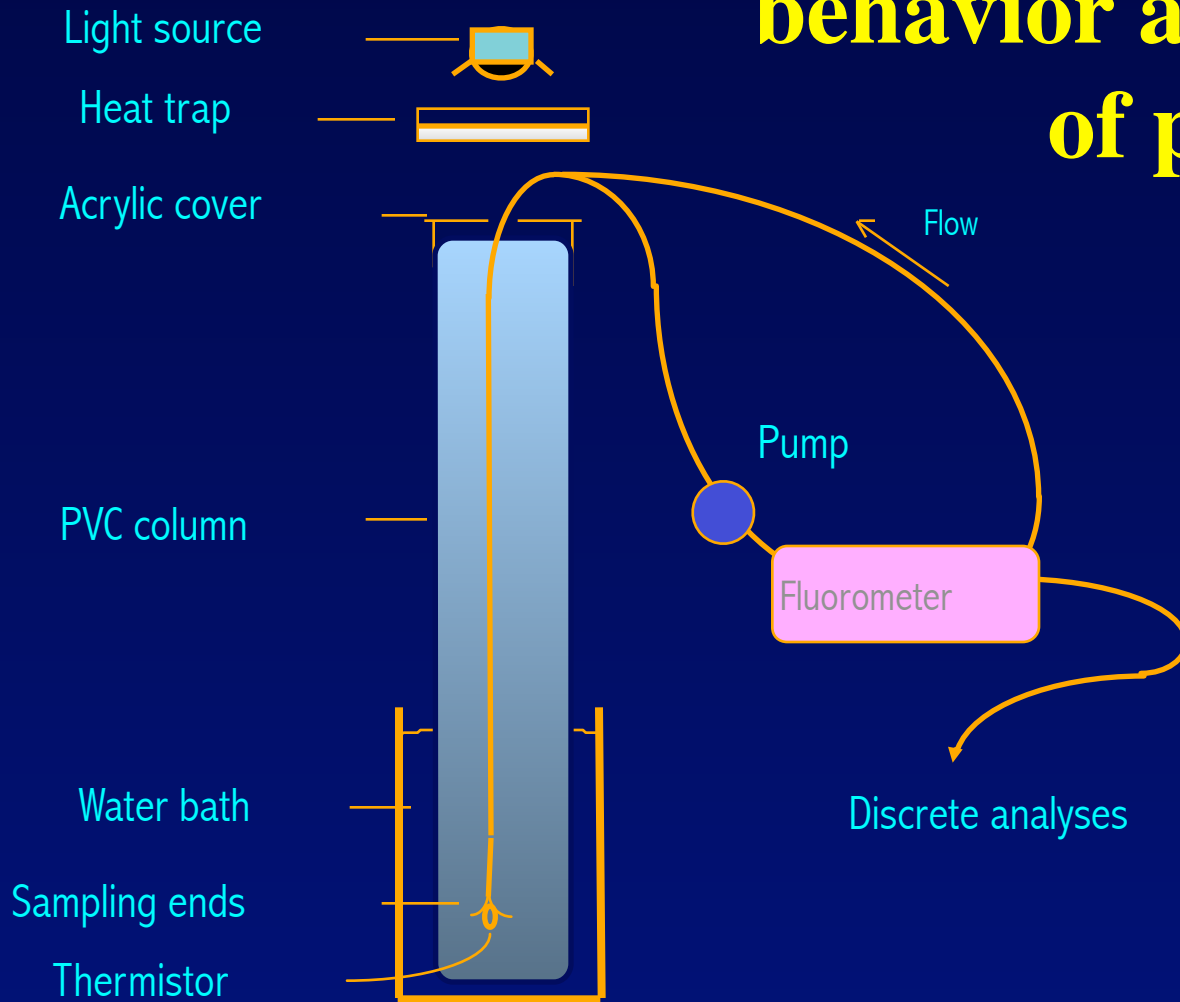
# And what about vertical migrators?



Nutrient concentration?

Temperature?

# “Cheap Tank” — A useful tool for studying the behavior and physiology of phytoplankton



Other mesocosms have been used in a few labs (e.g., M. Estrada)

# Migration studies

Excellent tool for describing behavior in natural gradients of environmental factors

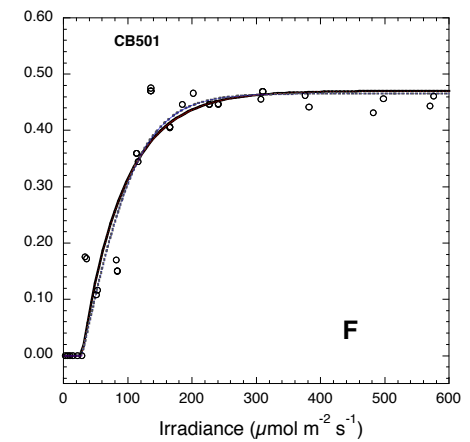
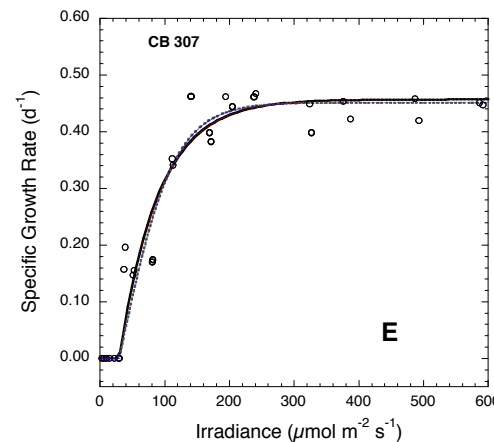
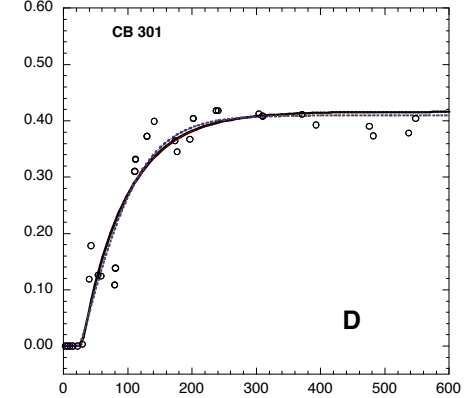
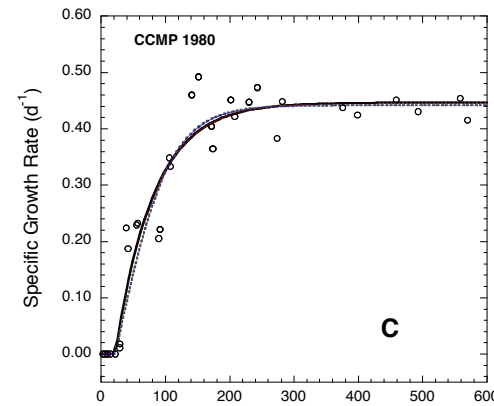
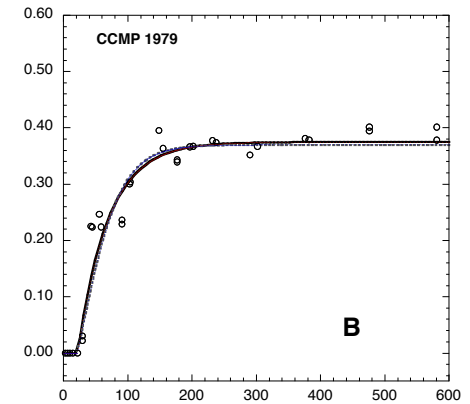
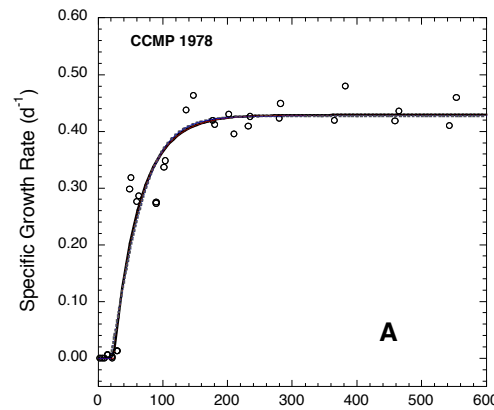
Difficult to specify growth rate in nature

Good for examining interspecific and intraspecific variability in ecologically important traits

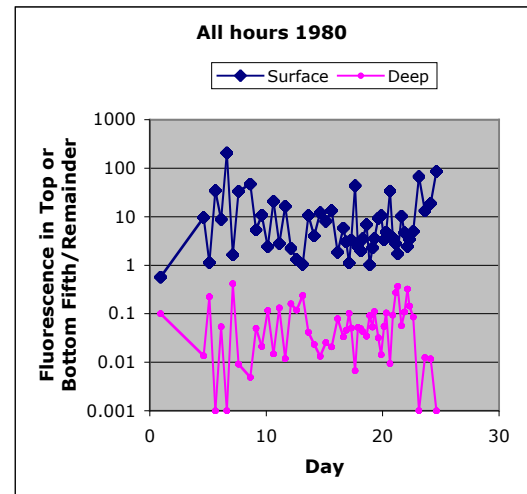
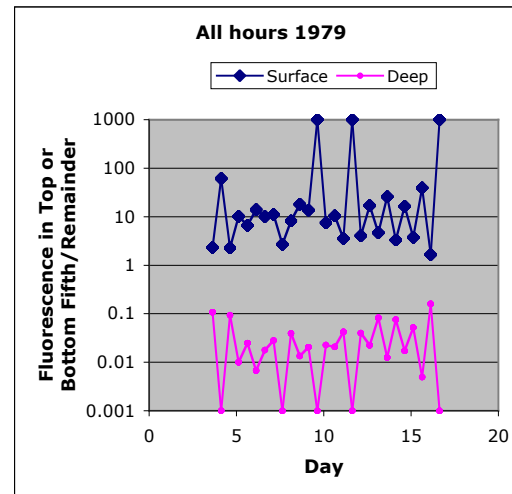
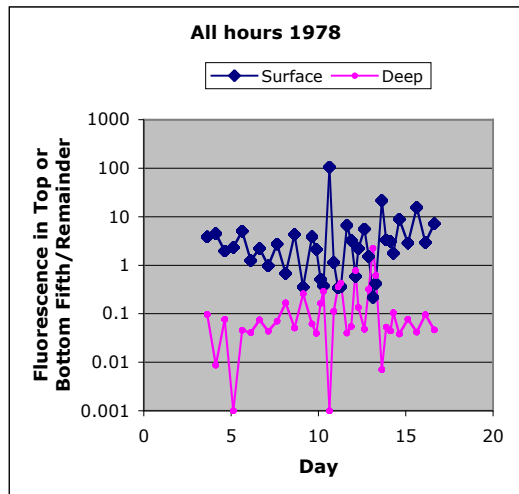
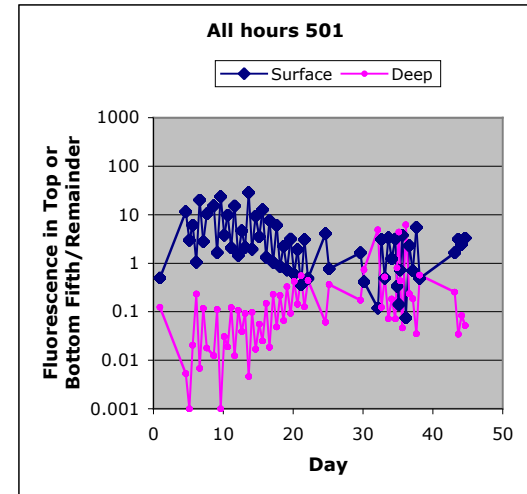
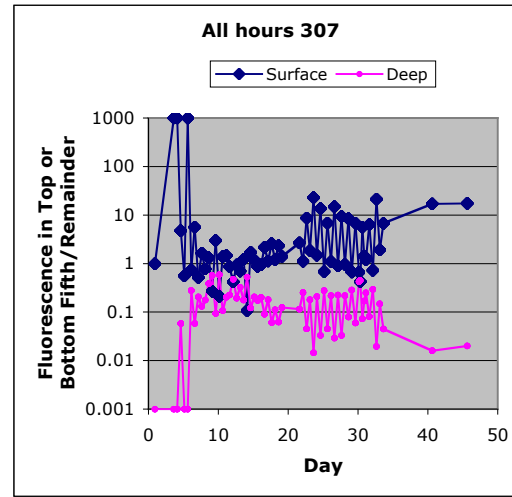
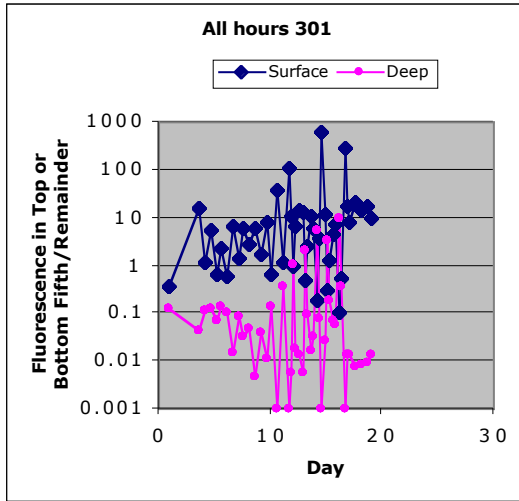
# Alexandrium fundyense

## Results for 6 strains:

## Growth Rate vs Irradiance



# Behavior showed significant variability: Aggregation at top vs bottom all hours



| Top Layer < 5  $\mu\text{M}$

..... Bottom Layer < 5  $\mu\text{M}$

| Nitrate Restored



# Bouncing around Redfield

NATO ASI Series, Vol. G 41  
 Physiological Ecology of Harmful Algal Blooms  
 Edited by D.M. Anderson, A.D. Cembella, and G. M. Hallegraeff  
 © Springer-Verlag, Berlin, Heidelberg 1998

## Behavior, physiology and the niche of depth-regulating phytoplankton

John J. Cullen and J. Geoffrey MacIntyre

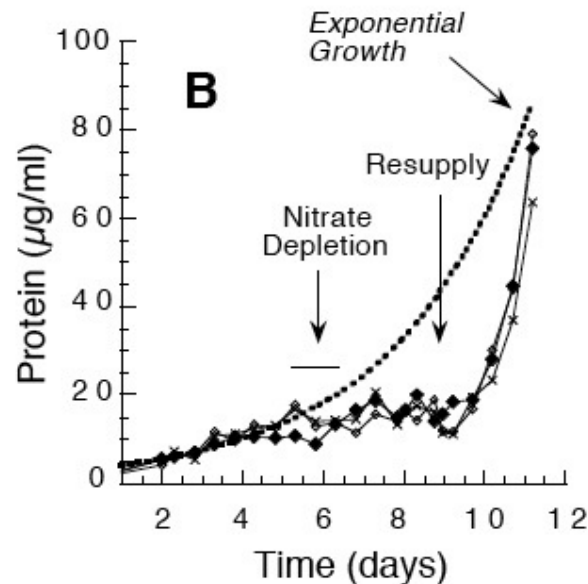
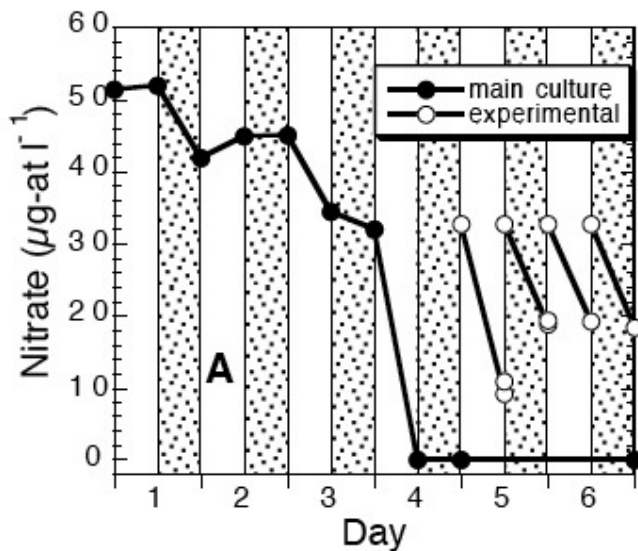
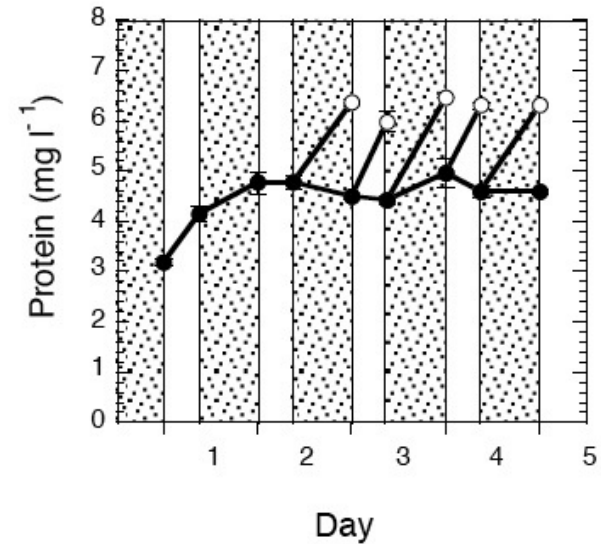


Fig. 4. Uptake and assimilation of nitrate after N-starvation. A. The diatom *Skeletonema*

# Bouncing around Redfield

Vol. 87: 123-134, 1992

MARINE ECOLOGY PROGRESS SERIES  
Mar. Ecol. Prog. Ser.

Published October 8

## Stoichiometric variations of N, P, C and O<sub>2</sub> during a *Gymnodinium catenatum* red tide and their interpretation

F. Fraga, F. F. Pérez, F. G. Figueiras, A. F. Ríos

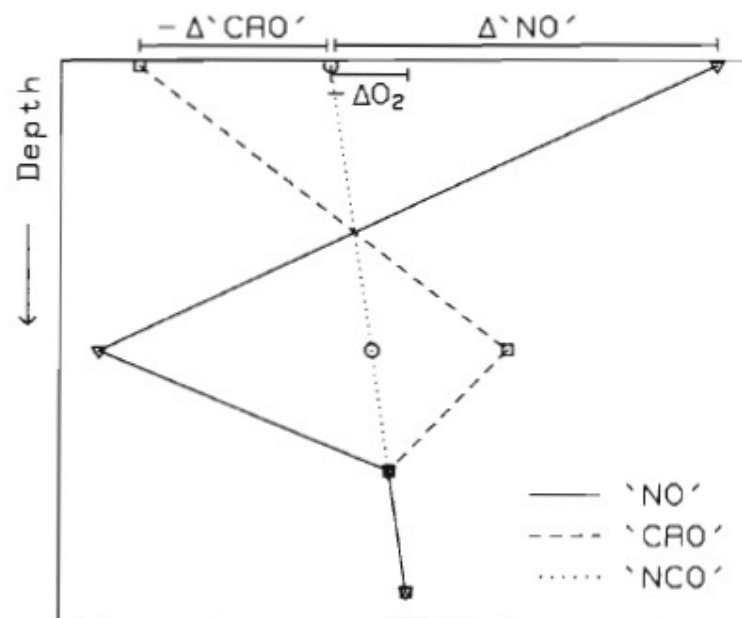


Fig. 5. Theoretical scheme of vertical variation of the chemical parameters 'NO' or 'PO', 'CAO' and 'NCO' or 'PCO' according to the data given in Table 3. The loss of oxygen ( $-\Delta O_2$ ) to the atmosphere is also shown and decreases linearly with depth

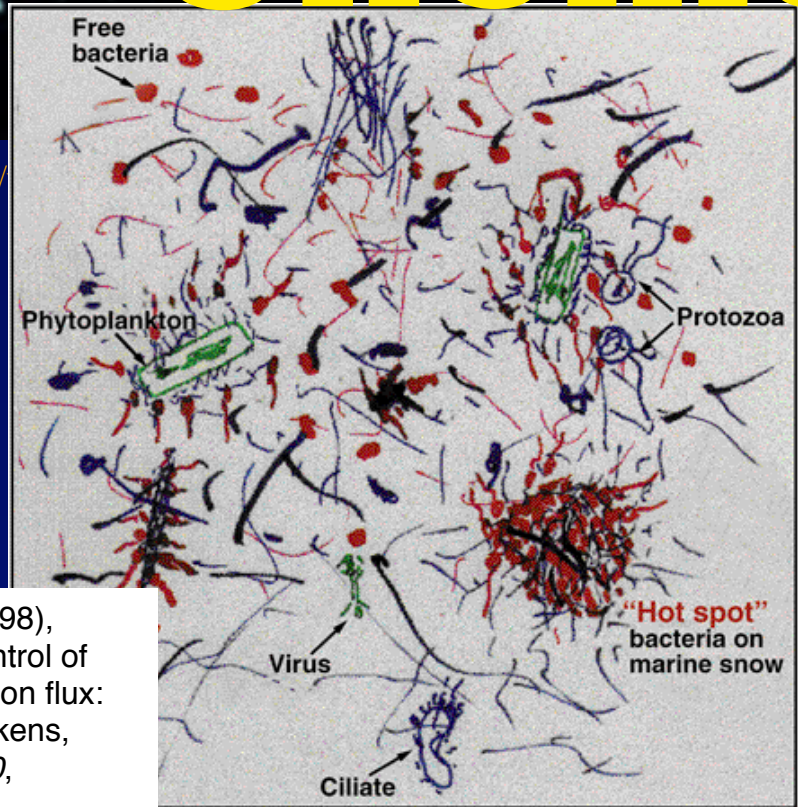
# Rapid chemotactic response enables marine bacteria to exploit ephemeral microscale nutrient patches

Roman Stocker\*†, Justin R. Seymour\*, Azadeh Samadani‡, Dana E. Hunt\*, and Martin F. Polz\*

\*Department of Civil and Environmental Engineering, Massachusetts Institute of Technology, 77 Massachusetts Avenue, Cambridge, MA 02139; and ‡Department of Physics, Brandeis University, 415 South Street, Waltham, MA 02454

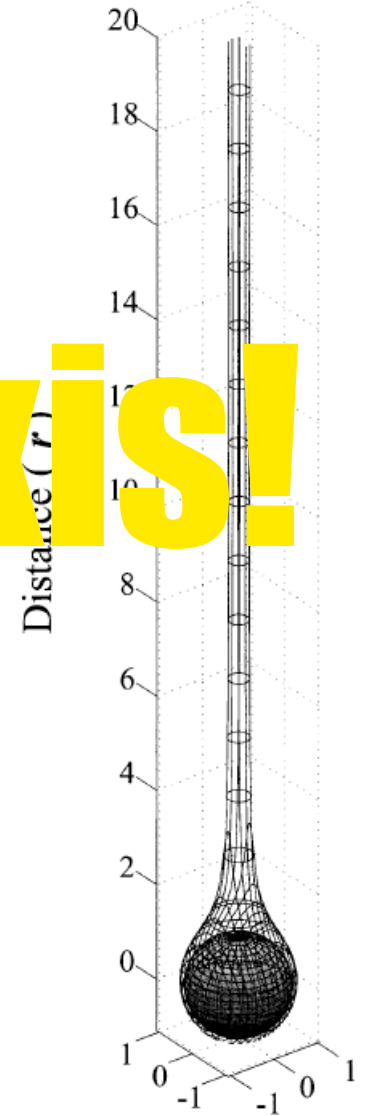
Edited by David M. Karl, University of Hawaii, Honolulu, HI, and approved January 22, 2008 (received for review October 14, 2007)

# Chemotaxis!



http://

Azam, F. (1998), Microbial control of oceanic carbon flux: The plot thickens, *Science*, 280, 694-696.



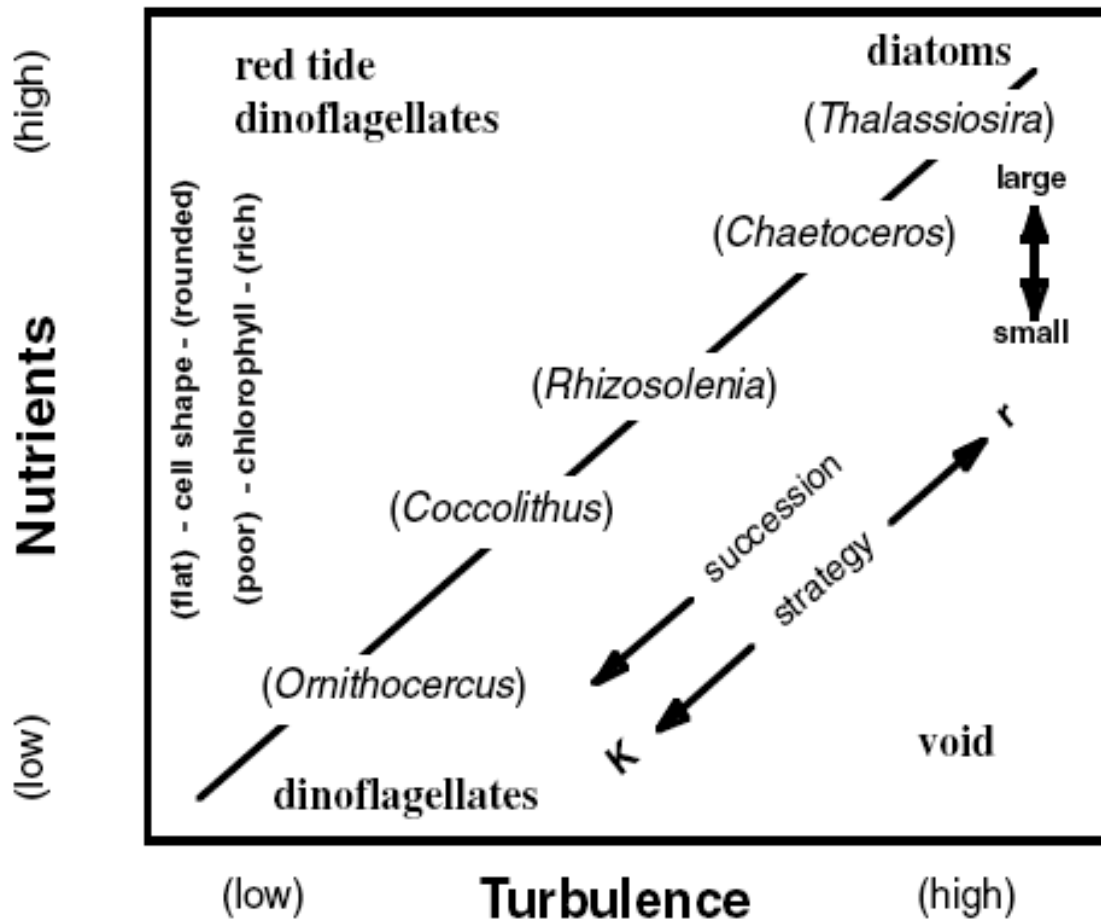
*Limnol. Oceanogr.*, 46(6), 2001, 1309-1318  
© 2001, by the American Society of Limnology and Oceanography, Inc.

Marine snow, organic solute plumes, and optimal chemosensory behavior of bacteria

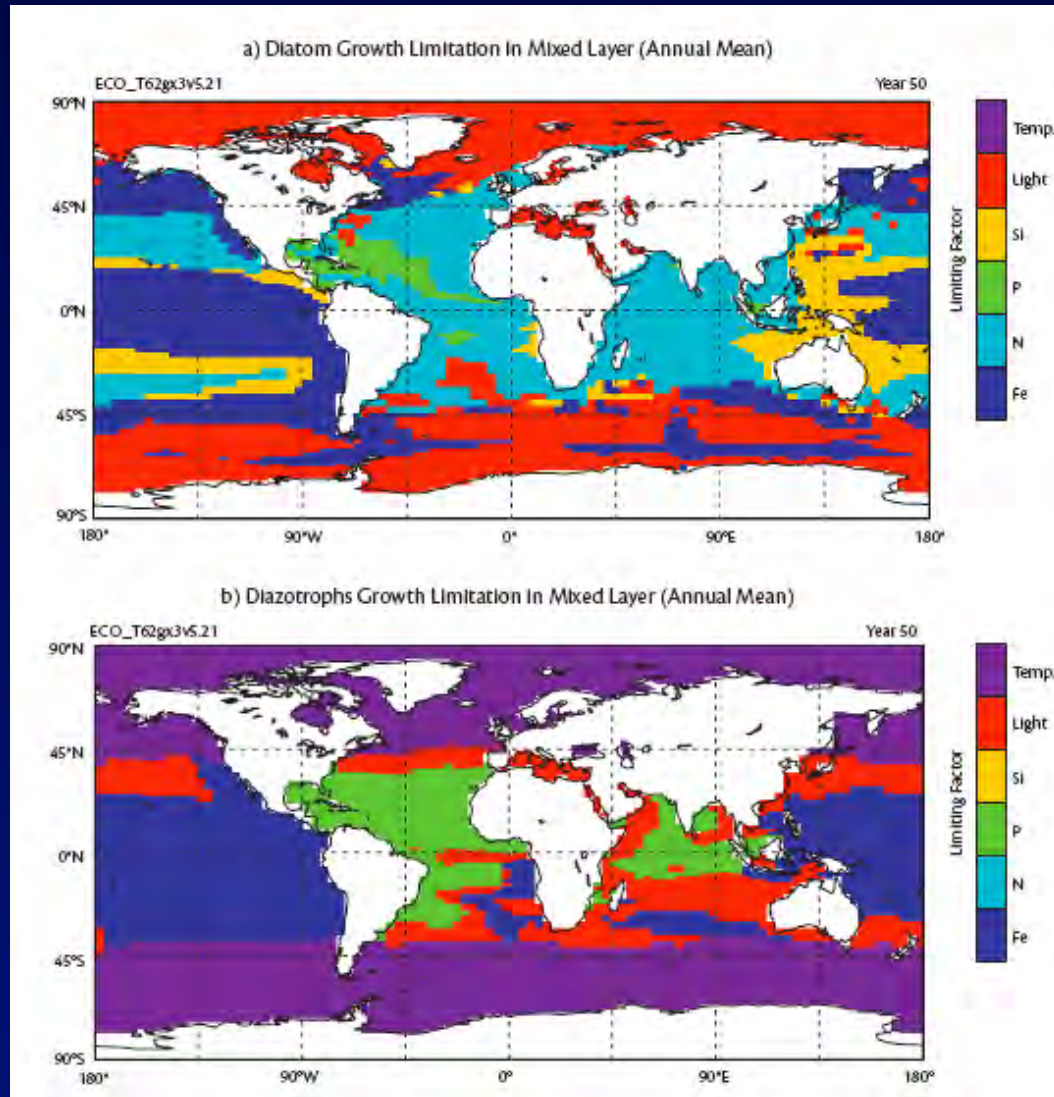
Thomas Kjørboe  
Danish Institute for Fisheries Research, Kavalergaarden 6, DK-2920 Charlottenlund, Denmark<sup>1</sup>

George A. Jackson  
Department of Oceanography, Texas A&M University, College Station, Texas 77843

# Niches abound!

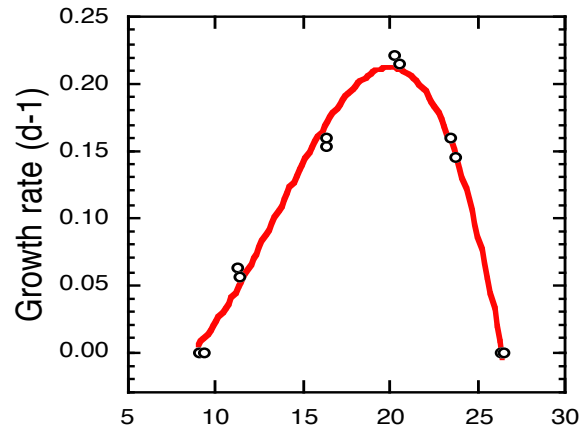


# But what about quantitative prediction?

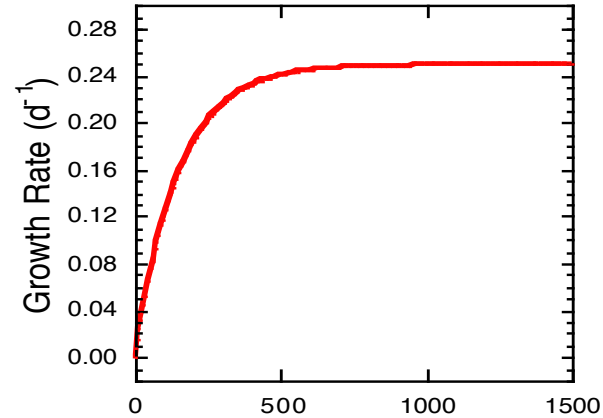


# Growth must be described as a function of environmental conditions

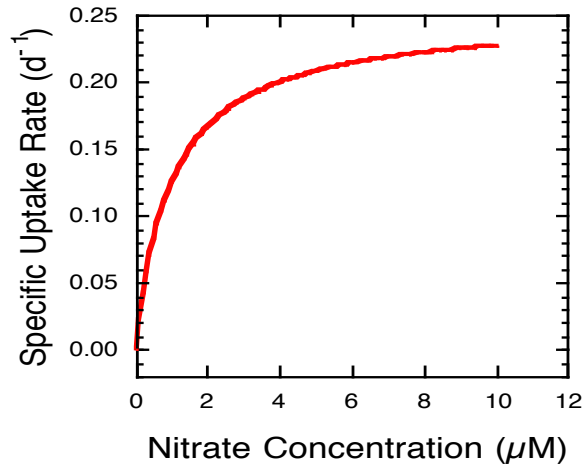
*Alexandrium ostenfeldii*



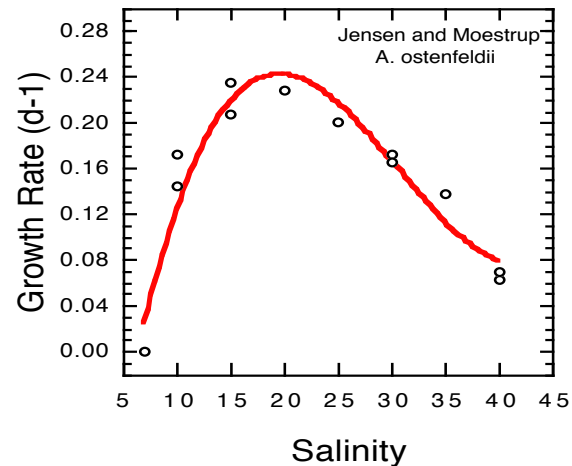
Growth vs Irradiance



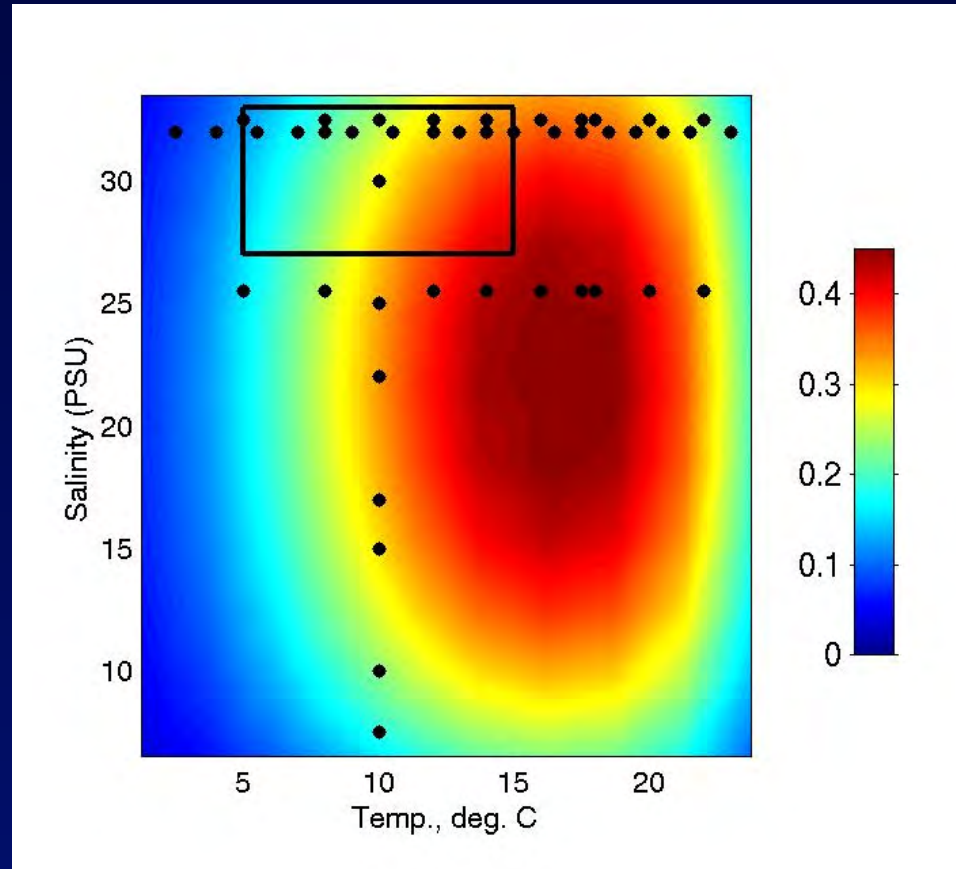
Nutrient Uptake Kinetics



Effect of Salinity



# Functions can be Developed



McGillicuddy,  
Stock, Anderson,  
and Signell  
WHOI

$$\mu = f(D, E, N, T)$$

(Daylength, Irradiance, Temperature, Nutrients)

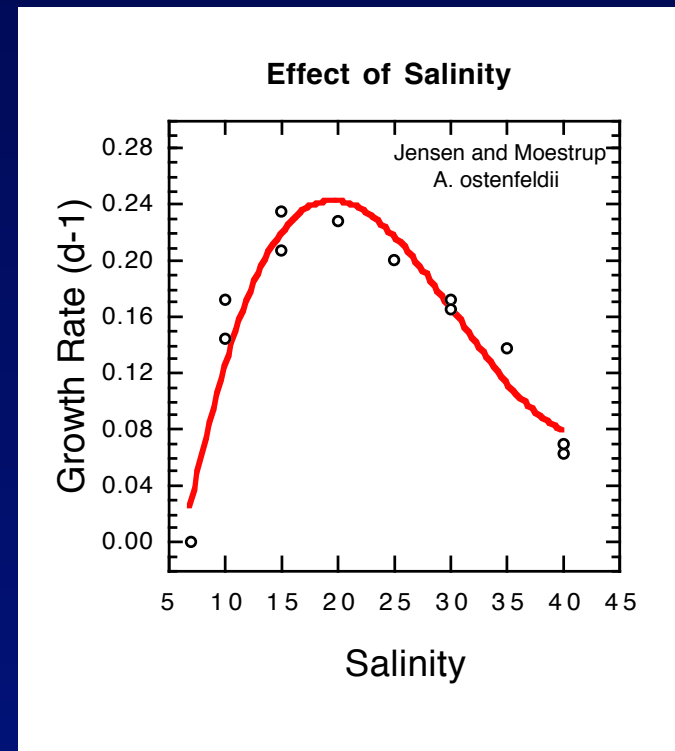
...but

- Requires a huge amount of work with cultures
- Algae should be acclimated to each set of conditions
  - This can require several weeks
- Conditions in nature are almost never so stable
  - Phytoplankton are subject to vertical mixing
  - Vertical migration
- All combinations of *Daylength*, *Irradiance*, *Nutrients* and *Temperature* are essentially impossible to test



# Summary: $\mu$ as a function of environmental conditions

Good for identifying environmental ranges and optima



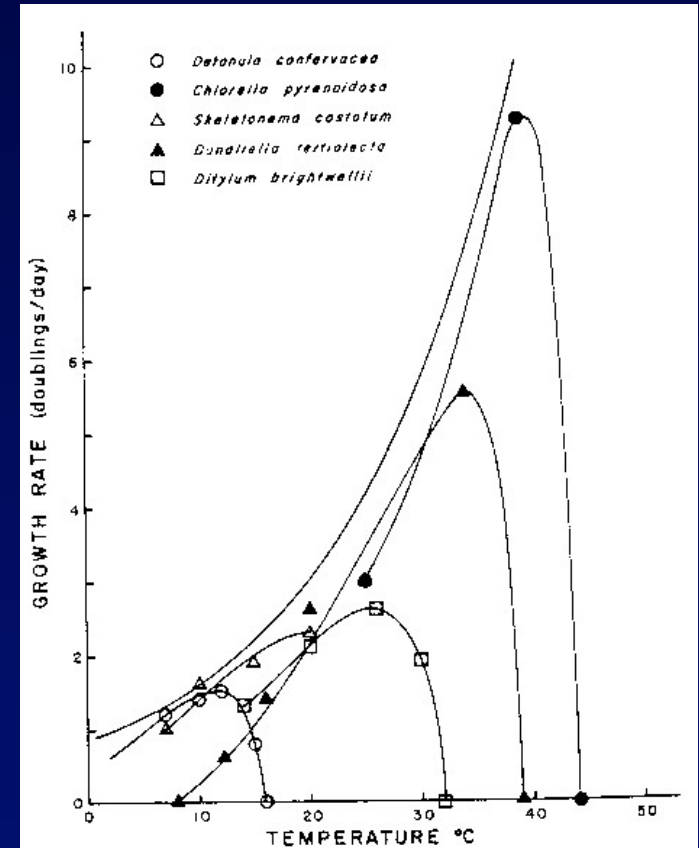
# Summary: $\mu$ as a function of environmental conditions

Excellent for describing differences between species

(and variations among strains of the same species)

*Ancient example:*

Gallagher, J. C. (1982). Physiological variation and electrophoretic banding patterns of genetically different seasonal populations of *Skeletonema costatum* (Bacillariophyceae). *J. Phycol.* 18: 148-162.



*Even more ancient:*

Eppley, R. W. 1972. Temperature and phytoplankton growth in the sea. *Fish. Bull.* 70: 1063-1085.

# Can we assume that adaptation provides all the raw genetic material for aggregate super-bugs?

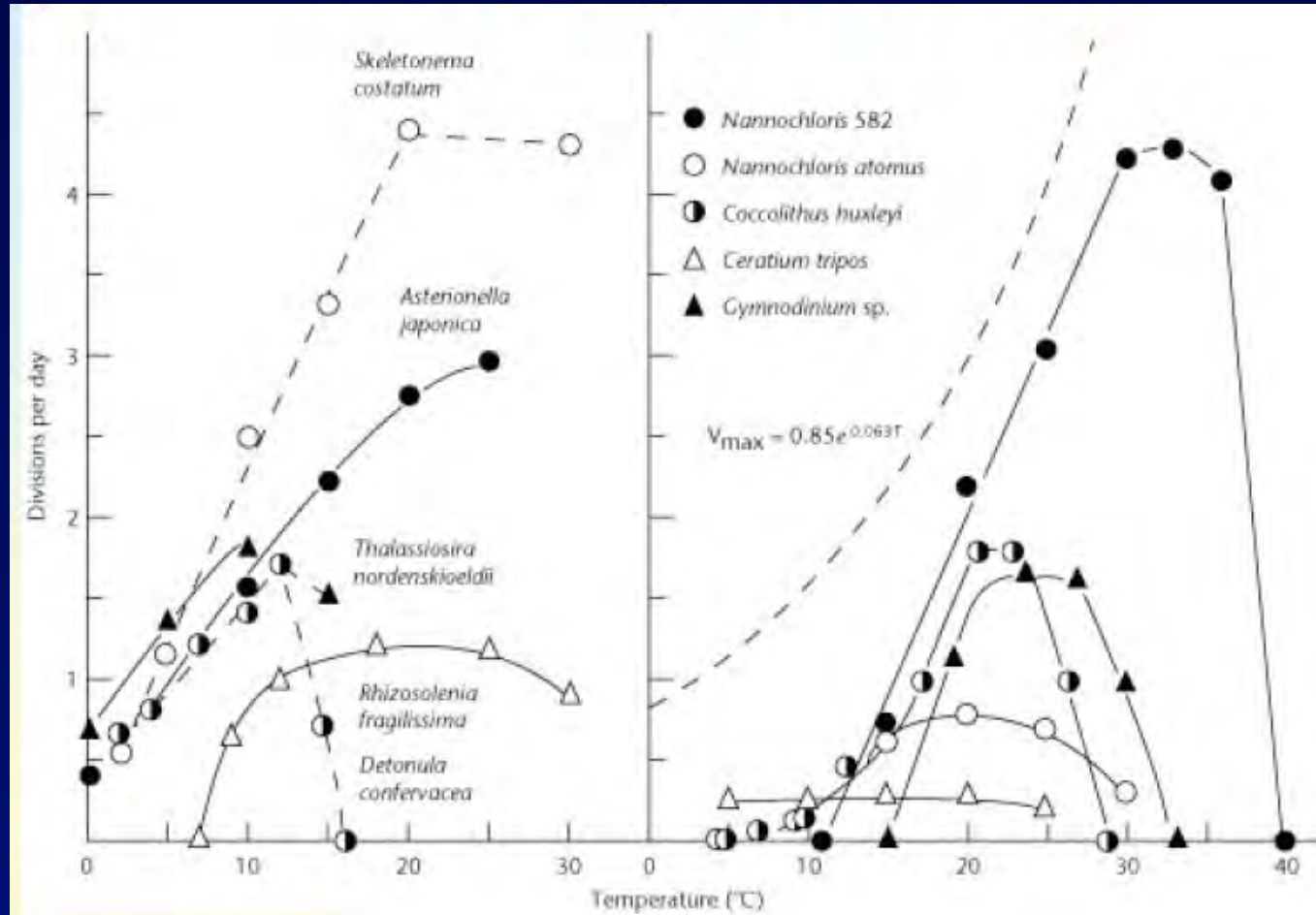


Figure from C.B. Miller, "Biological Oceanography" after Smayda, 1976

see Eppley, R. W. 1972. Temperature and phytoplankton growth in the sea. Fish. Bull. 70: 1063-1085.

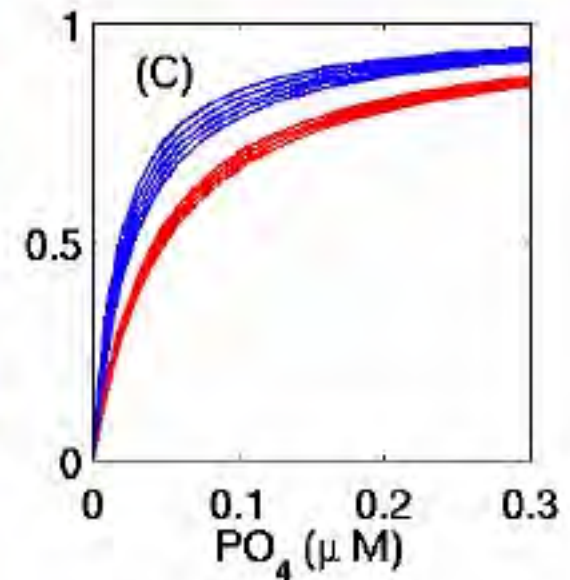
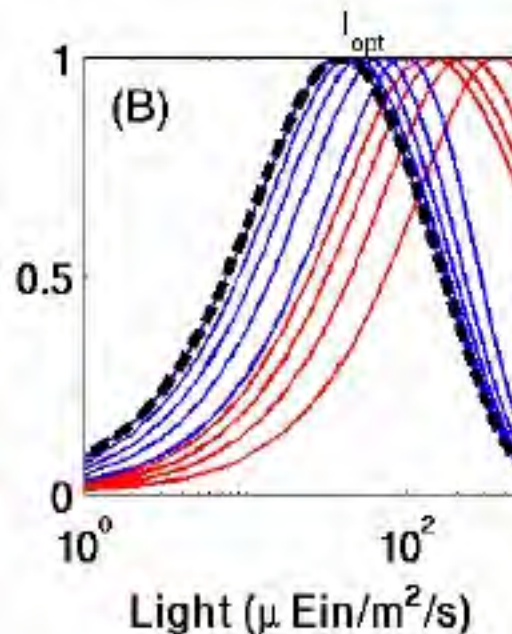
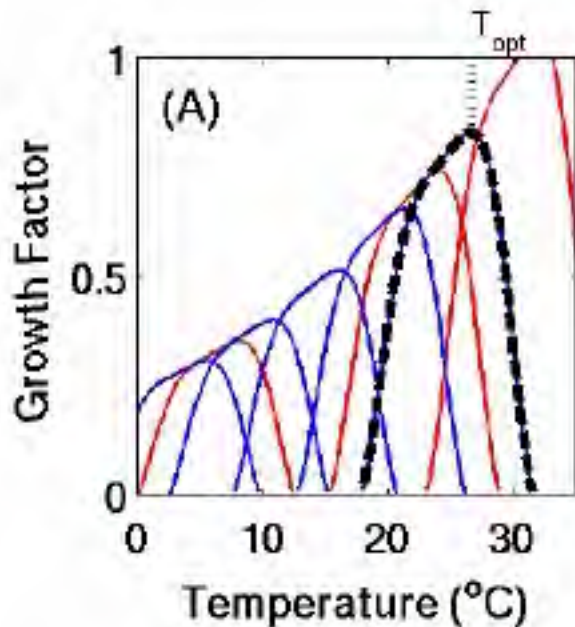
# A novel approach

## Emergent Biogeography of Microbial Communities in a Model Ocean

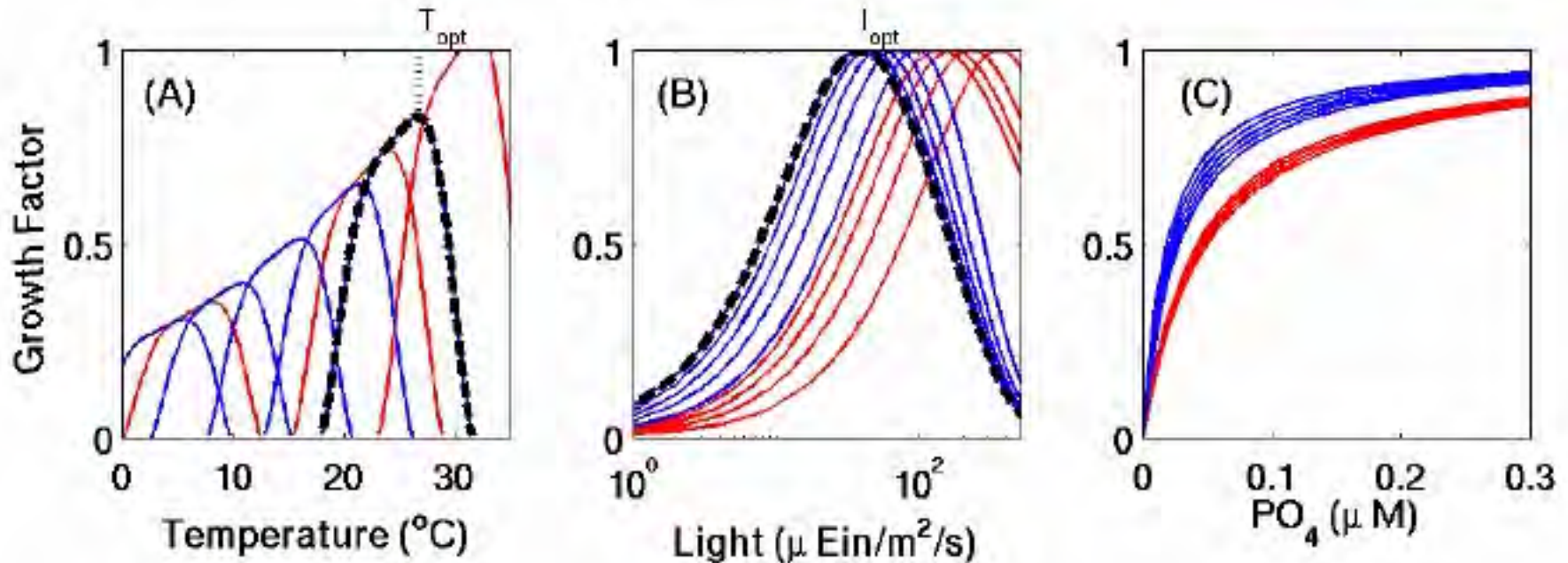
Michael J. Follows,<sup>1\*</sup> Stephanie Dutkiewicz,<sup>1</sup> Scott Grant,<sup>1,2</sup> Sallie W. Chisholm<sup>3</sup>

SCIENCE VOL 315 30 MARCH 2007

1843



# Everything (well, a lot of things) is everywhere and the environment selects



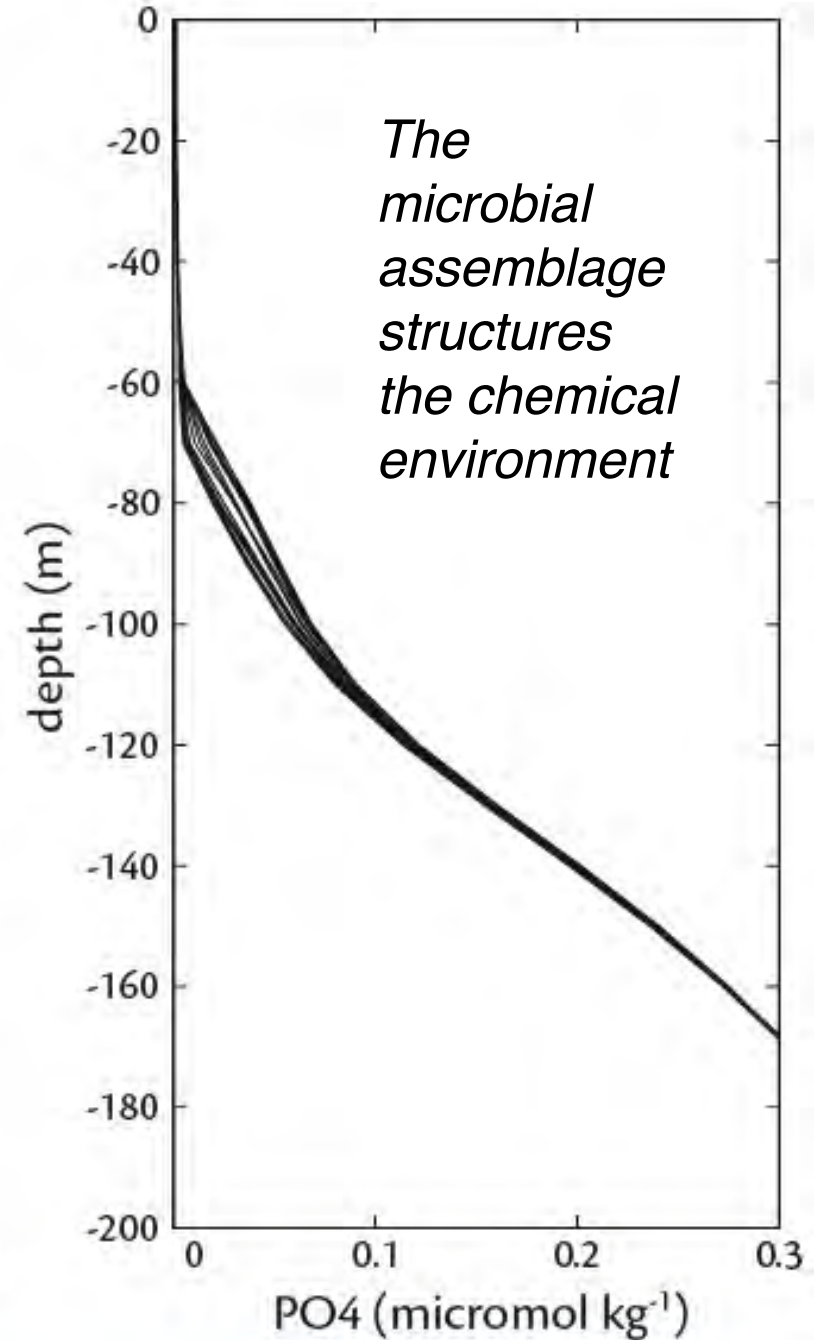
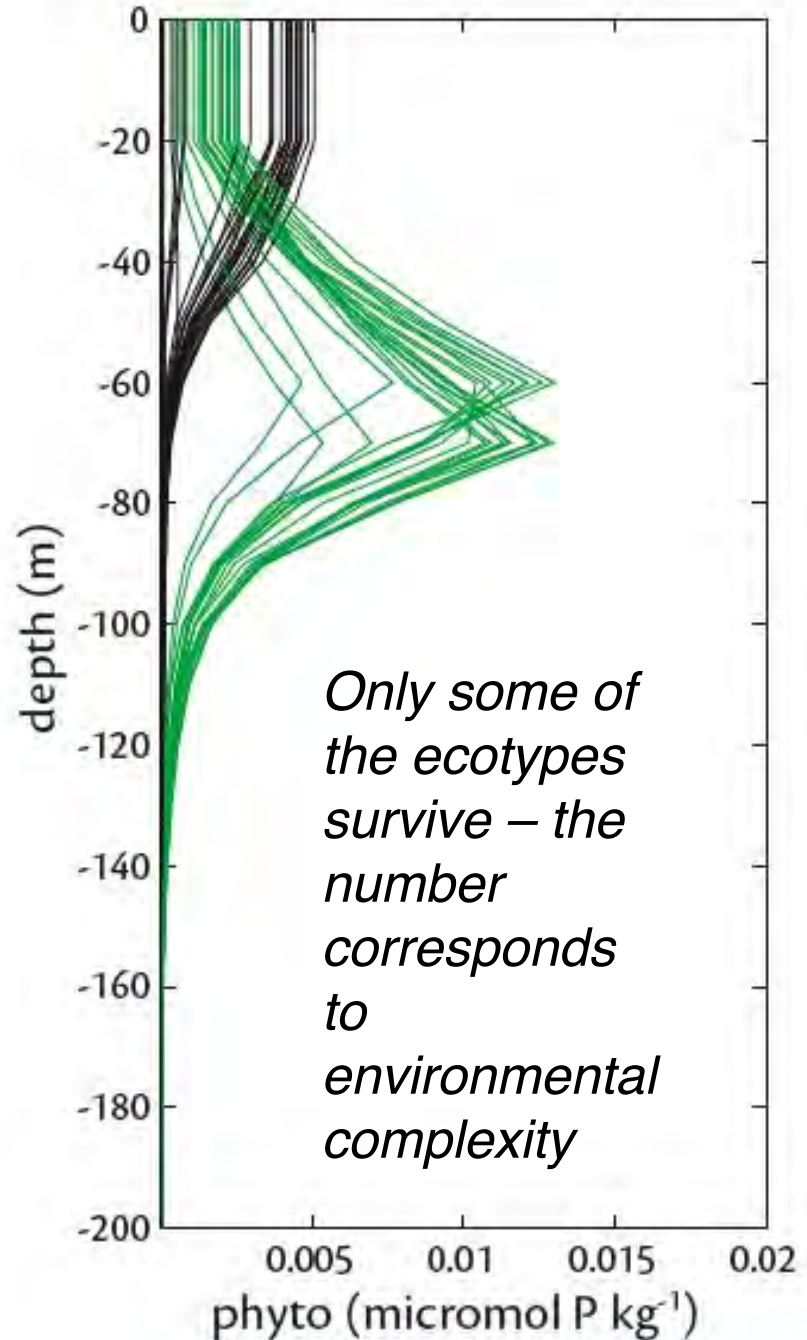
## Emergent Biogeography of Microbial Communities in a Model Ocean

Michael J. Follows,<sup>1\*</sup> Stephanie Dutkiewicz,<sup>1</sup> Scott Grant,<sup>1,2</sup> Sallie W. Chisholm<sup>3</sup>

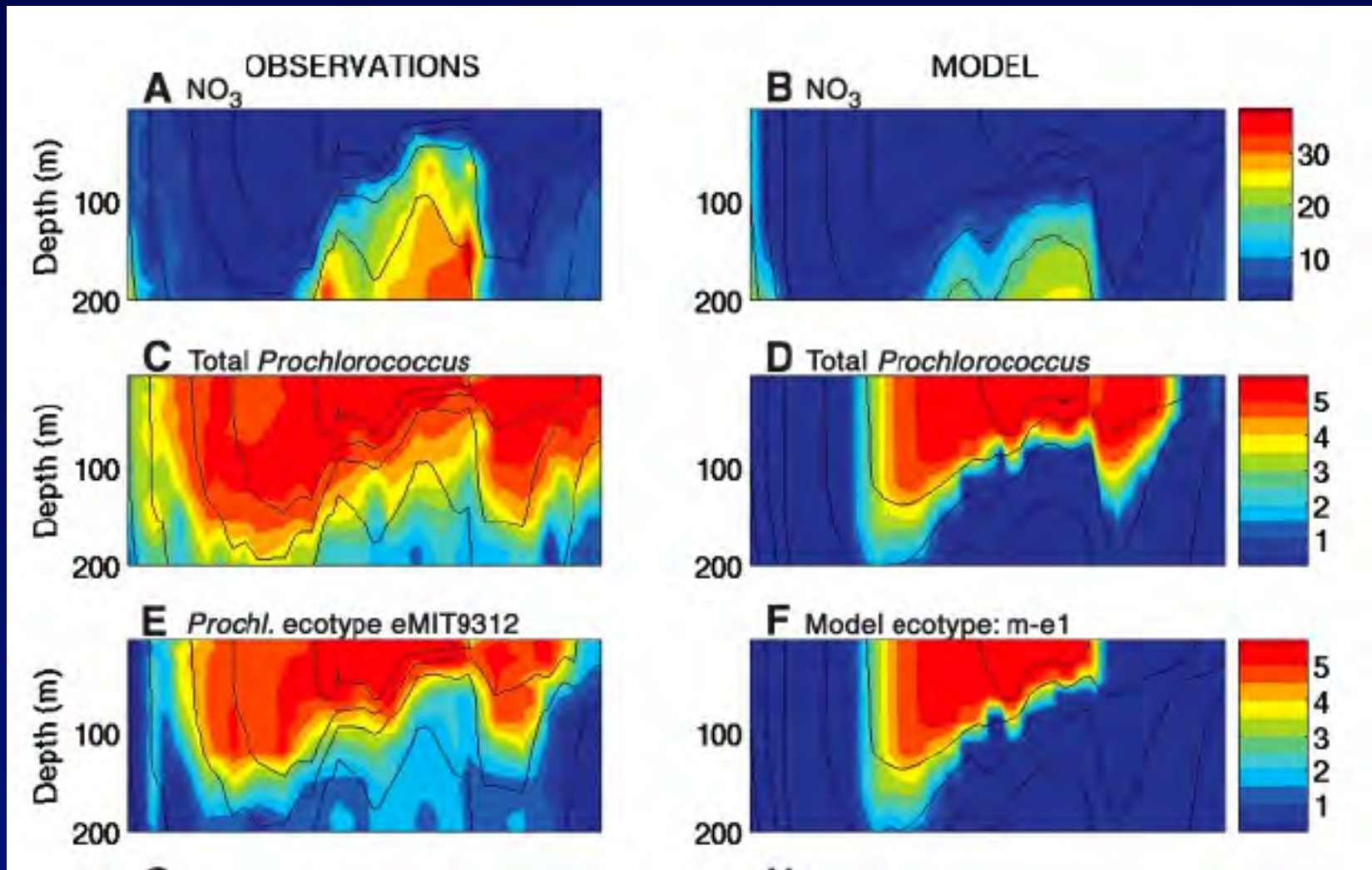
SCIENCE VOL 315 30 MARCH 2007

1843

Natural selection *in silico*

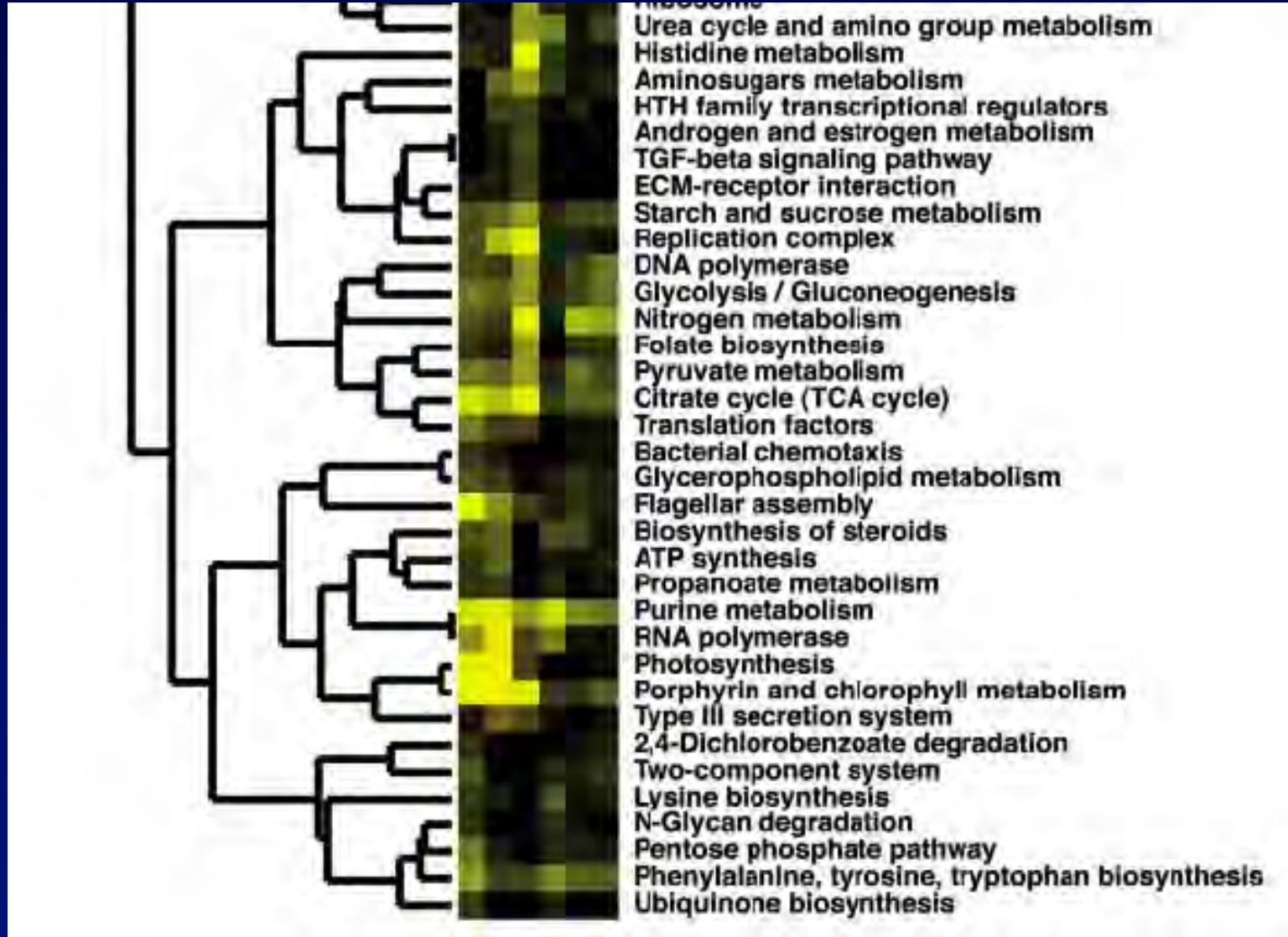


# A test of our understanding of what structures marine ecosystems (LPG)



Future steps:

Assembly and natural selection of microbial communities guided by metagenomics



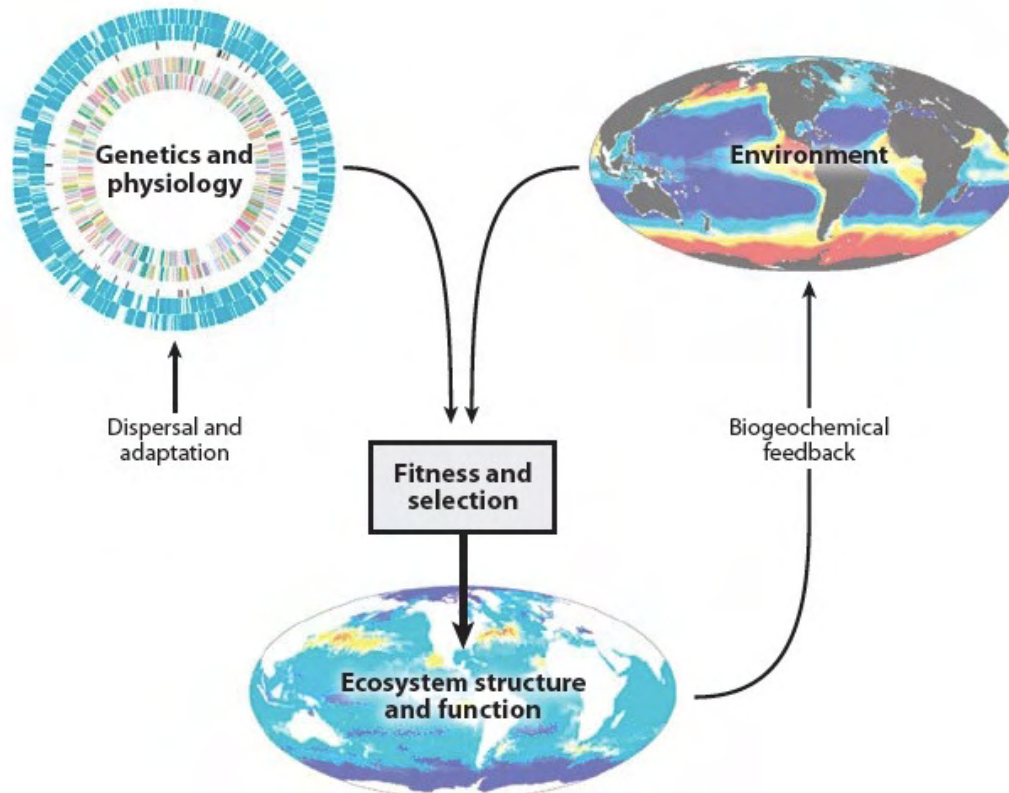


A big part of  
the answer is  
here

# Modeling Diverse Communities of Marine Microbes

Michael J. Follows and Stephanie Dutkiewicz

Earth, Atmospheric, and Planetary Sciences, Massachusetts Institute of Technology,  
Cambridge, Massachusetts 02139; email: mick@ocean.mit.edu, stephd@mit.edu



Annu. Rev. Mar. Sci. 2011. 3:427–51

The *Annual Review of Marine Science* is online at  
[marine.annualreviews.org](http://marine.annualreviews.org)

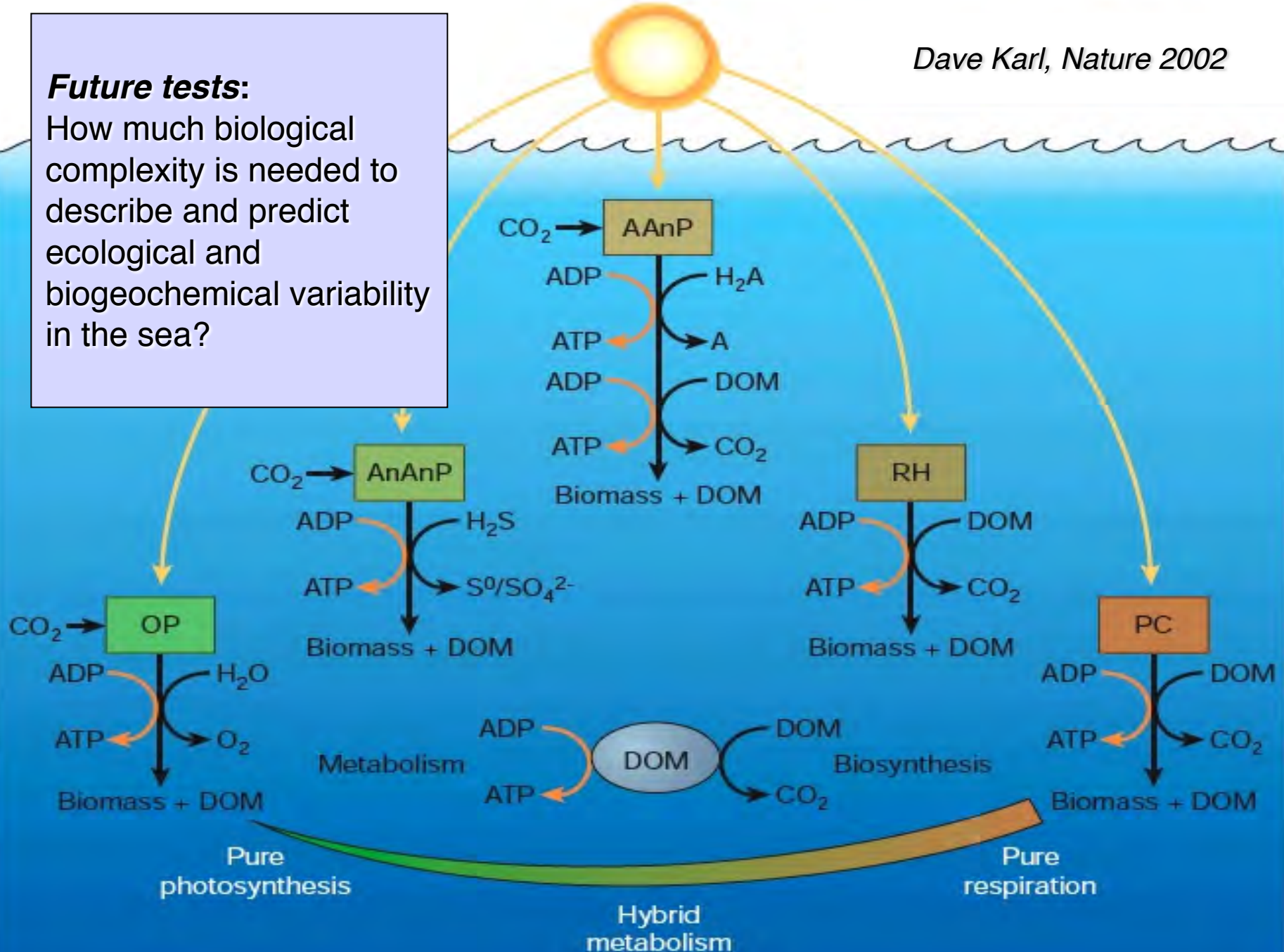
This article's doi:  
10.1146/annurev-marine-120709-142848

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All rights reserved

1941-1405/11/0115-0427\$20.00

**Future tests:**

How much biological complexity is needed to describe and predict ecological and biogeochemical variability in the sea?



# Conclusions

Relationships between environmental variability and microbial diversity must be described and ultimately predicted to understand the ecology and biogeochemistry of the sea

## Niches abound:

wide range of environmental tolerances

specialized life-styles (nutrient requirements, alternate modes of photosynthesis)

physiological plasticity vs specialization for stable environments

Complexity will never be fully described with numerical models

The degree of model complexity can and should be related to the ecological/biogeochemical question and its scale.

**This can be done — but models must be verified by measurements!**

*Mahalo!*