

Nitrogen cycling in the ocean

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Outline:

1. The Redfield ratio
2. The nitrogen cycle
3. Liebig's Law of the Minimum
4. New & regenerated production
5. Is the N cycle in steady state?

Nitrogen Revolution mid-1990s-present



- Nitrification
- Denitrification/anammox
- N₂ fixation
- Anthropogenic N inputs

ON THE PROPORTIONS OF ORGANIC DERIVATIVES IN SEA WATER AND THEIR RELATION TO THE COMPOSITION OF PLANKTON¹

ALFRED C. REDFIELD

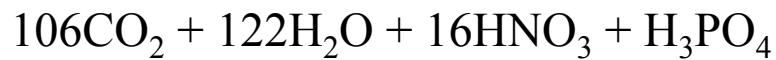
PROFESSOR OF PHYSIOLOGY, HARVARD UNIVERSITY, AND
SENIOR BIOLOGIST, WOODS HOLE OCEANOGRAPHIC INSTITUTION

(Received September 5, 1933)

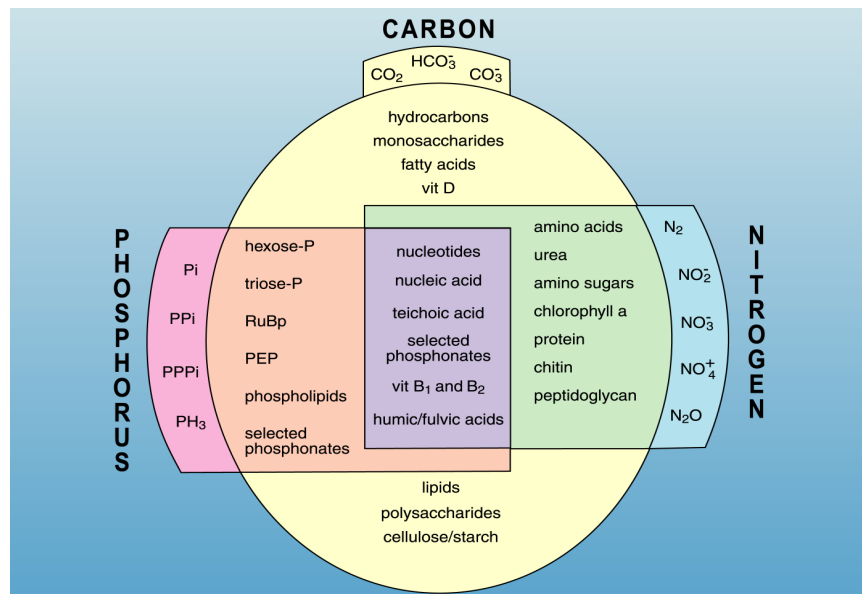
Redfield 1934 James Johnstone Memorial Volume

The Redfield Ratio

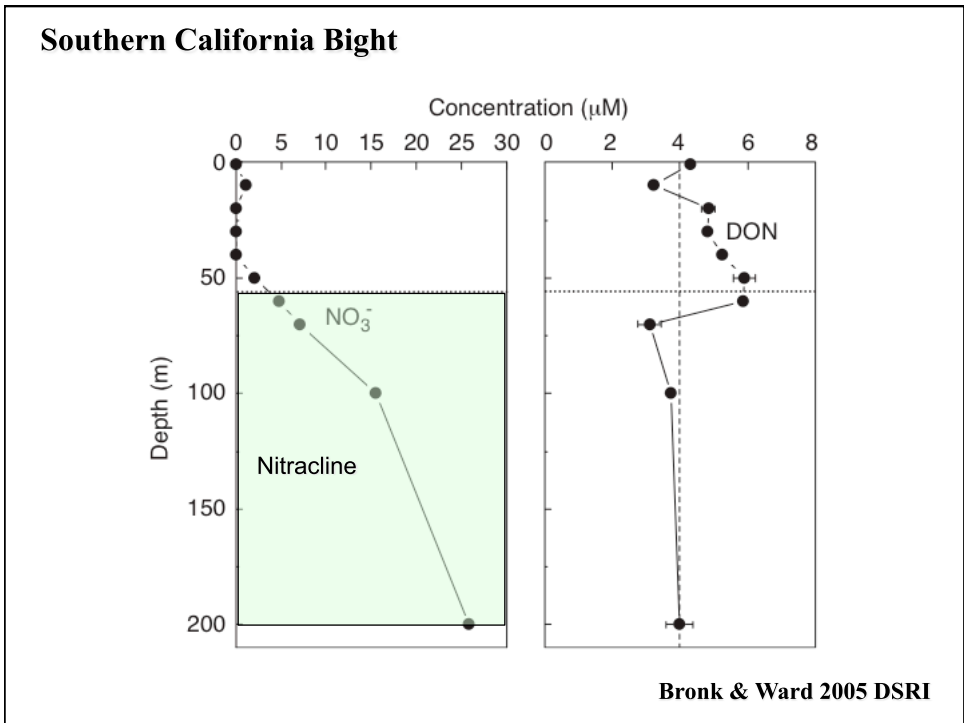
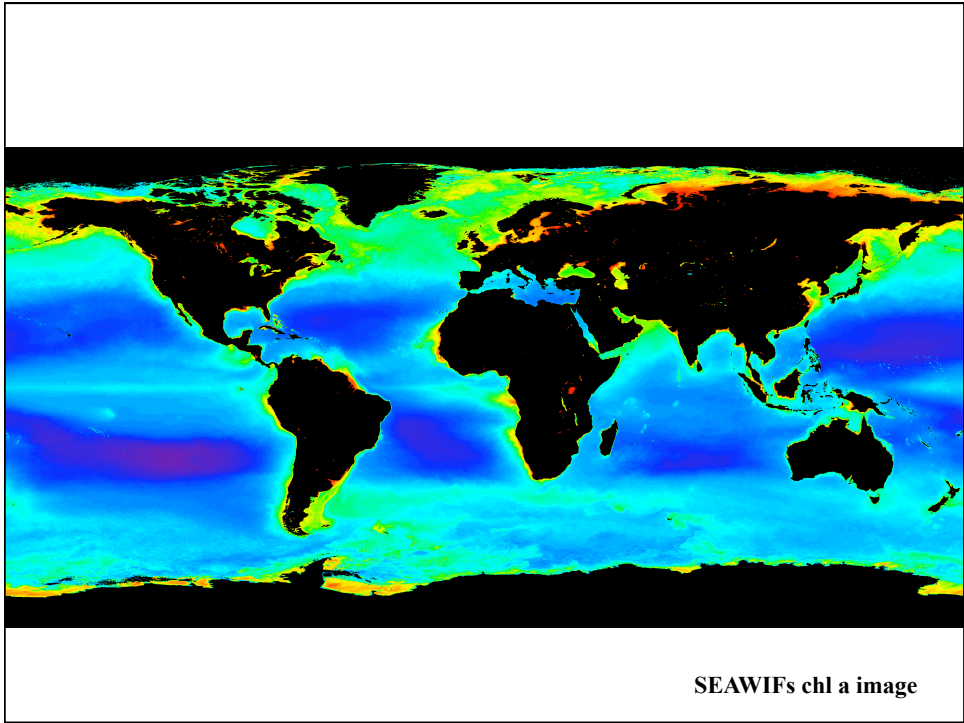
$$\text{C:N:P} = 106:16:1$$



detritus vs. phyto vs. bacteria?

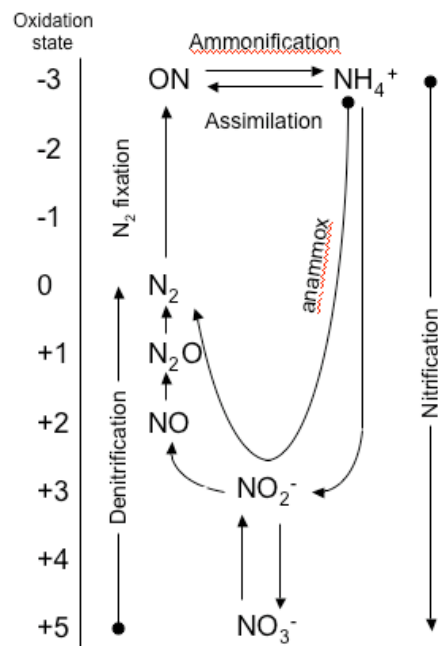


Karl & Björkman 2002 DOM book

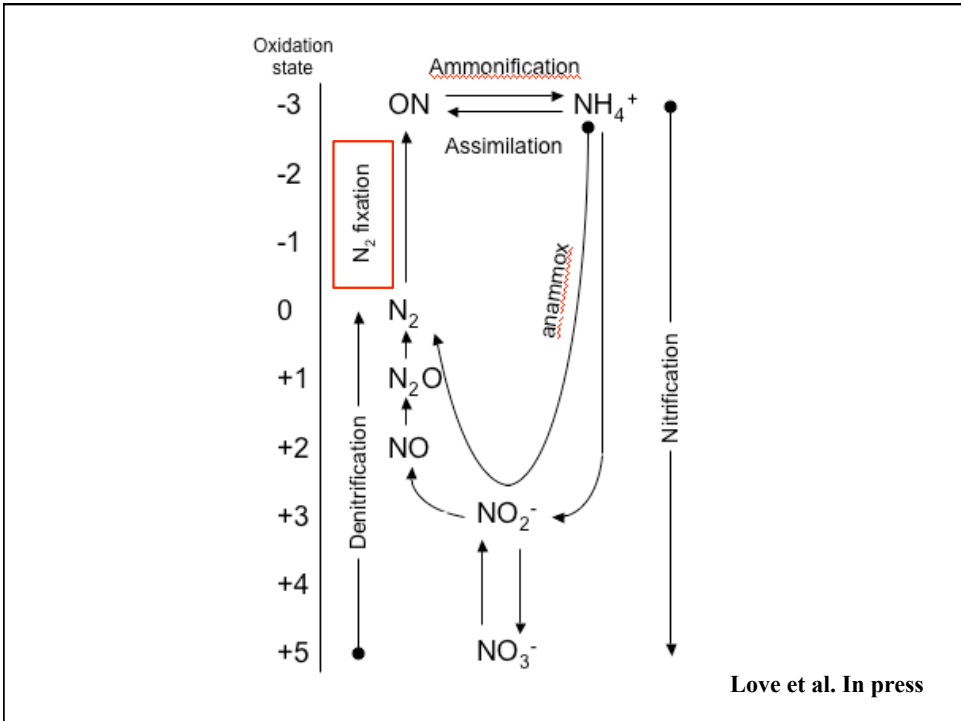
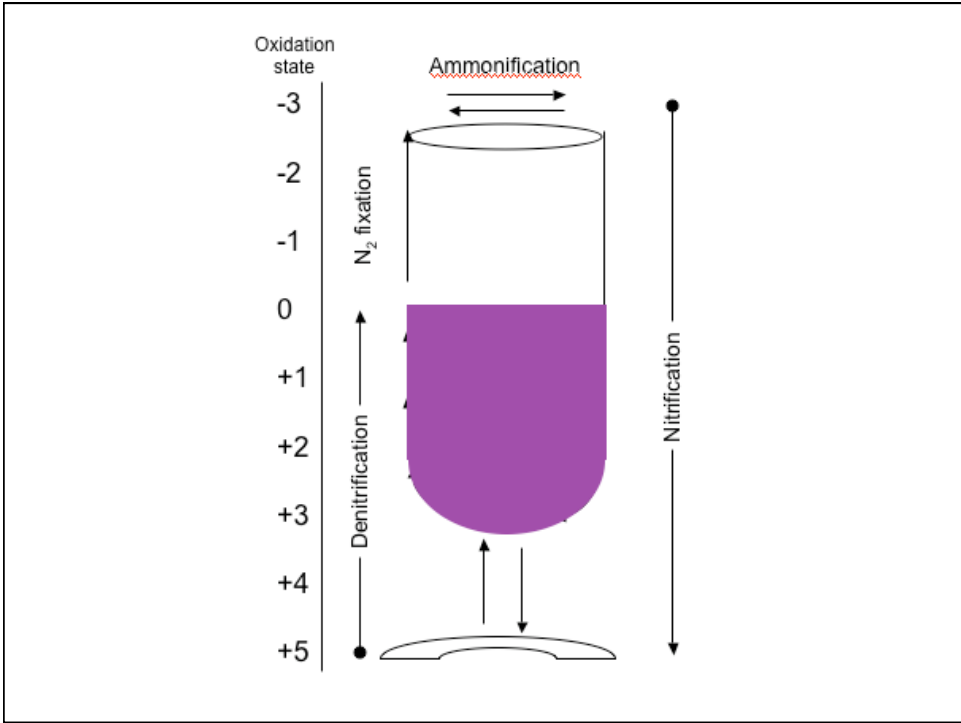


$$\text{TDN} - \text{DIN} = \text{DON}$$

$$\text{TDN} - (\text{NO}_3^- + \text{NO}_2^- + \text{NH}_4^+)$$

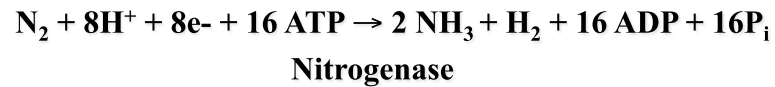


Love et al. In press



Love et al. In press

Nitrogen fixation



- “Fixing” broken N_2
- Energetically very expensive – N_2 has triple bond
- Nitrogenase is irreversibly inactivated by oxygen
- Most N_2 fixers form heterocysts
- Requires P, Fe, and trace metals (Mo, Co, V)

N_2 fixation



Trichodesmium



- Two morphological forms
- Colonial – 100s cells/trichome and 100s trichomes/colony
- Traditionally considered the dominant N fixer in the ocean
- Found in tropical and subtropical waters

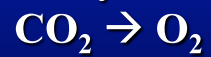
1987



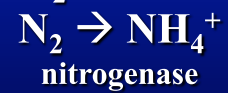
Trichodesmium mystery

Non-heterocystous

Photosynthesis



N_2 fixation



Temporal or spatial segregation?

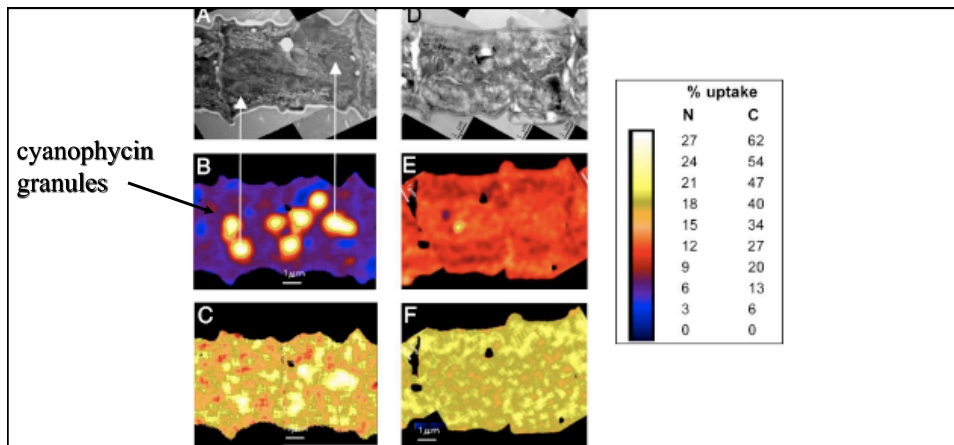


Fig. 3. TEM Images of ~2 cells from a *Trichodesmium* filament after (A) 8-h and (D) 24-h incubation with $H^{13}CO_3$ and $^{15}N_2$. Correlated NanoSIMS images demonstrate percentage fixed ^{15}N after (B) 8 h and (E) 24 h, percentage fixed ^{13}C after (C) 8 h and (F) 24 h. Arrows indicate correlation between cyanophycin granules identified by TEM and ^{15}N enriched hotspots evident in NanoSIMS image. (Scale bar, 1 μm .) Because NanoSIMS analysis is a destructive process, distinct cells were imaged for the 2 different time points.

High resolution-secondary ion mass spectrometry (Nano SIMS)

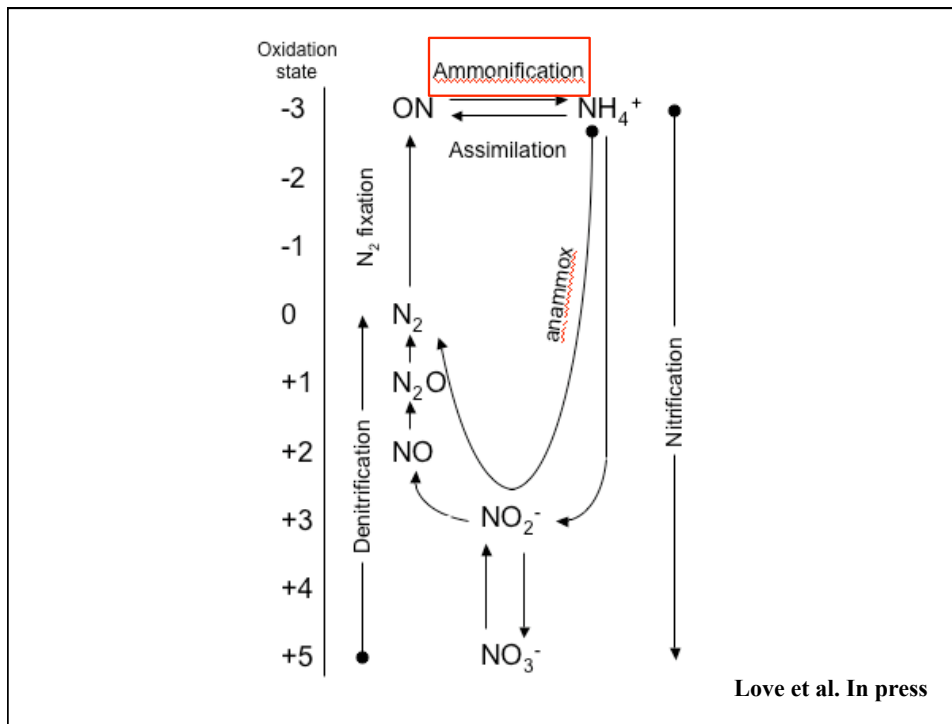
Supports temporal segregation!

Finzi-Hart et al. 2009 PNAS

N₂ fixation

Zehr et al. 2001 Nature
 Unicellular cyanobacteria that expressed nitrogenase at HOT

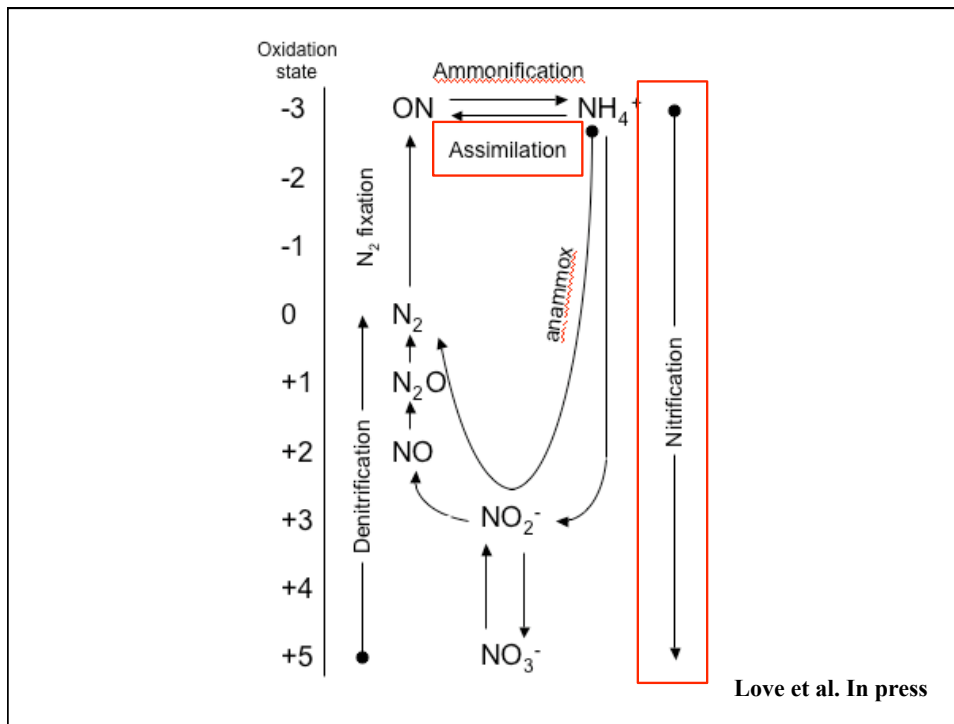
Montoya et al. 2004 Nature
 Rates of N fixation by the single cell forms can equal or exceed rates by *Trichodesmium*



Ammonification -
the conversion of DON or PON to NH_4^+

Two types:

1. Bacterial -
 - a. Traditional view of bacterial decomposition
 - b. Was considered primary source of NH_4^+
(Now primary source if believed to be grazers)
2. Photochemical (abiotic!)
 - a. More recently recognized source
 - b. Importance of process is still debated



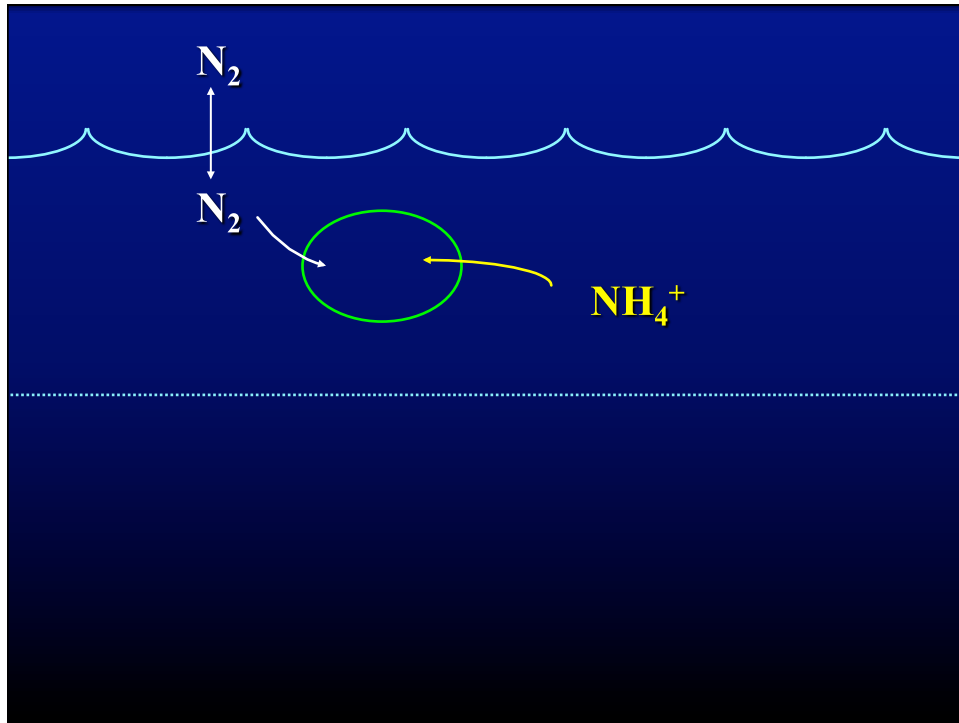
The Ocean

Euphotic zone

light - ~little N

Aphotic zone

no light - lots N



Nitrification

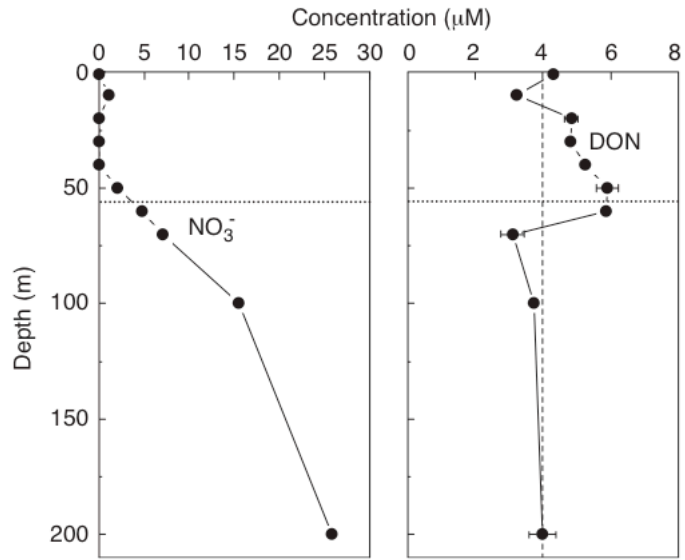


Ammonium oxidizers:
very slow growing
sensitive to light

Nitrite oxidizers:
faster growing
more sensitive to light

- Nitrifiers are chemolithoautotrophs.
- Maximum rates occur near the base of the euphotic zone.

Southern California Bight



Bronk & Ward 2005 DSRI

Nitrification



Nitrifying bacteria - or is it??

**Karner et al.
2001 Nature**

**39% of the
picoplankton in
the mesopelagic
at HOT are
archaea.**

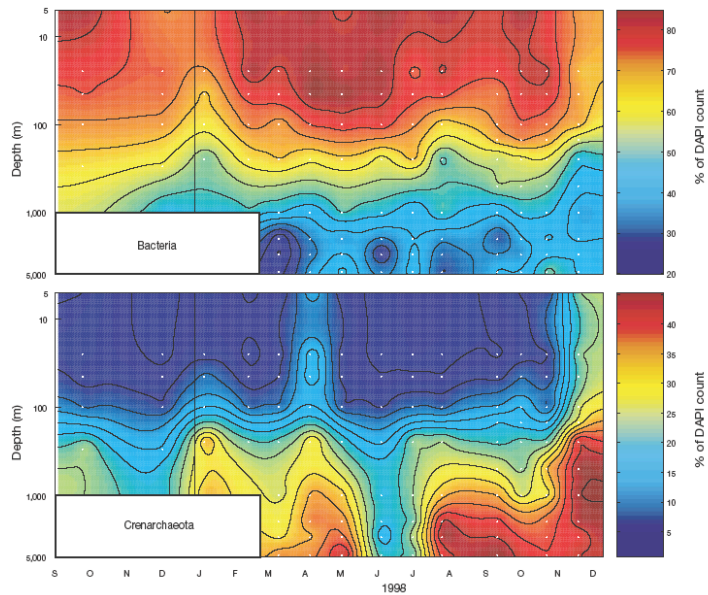


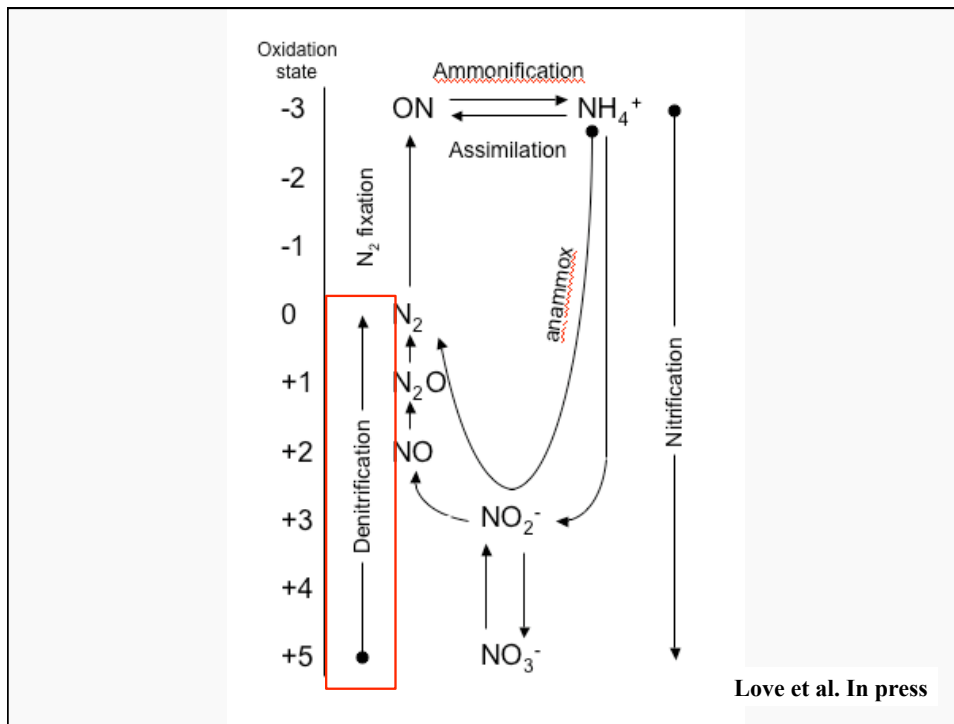
Figure 1 Contour plots of relative abundances with depth of bacteria and pelagic crenarchaeota during a 1-yr sampling effort at the Hawaii Ocean Time-series station, ALDH, in the North Pacific subtropical gyre. White dots indicate dates and depths where samples were collected. Contour lines are percentages of bacteria and pelagic crenarchaeota as compared with total microbial abundance at each depth. Total cell abundance was assessed using the DAPI nucleic acid stain. Bacteria and archaea were enumerated using whole-cell rRNA targeted fluorescent *in situ* hybridization with fluorescein-labelled polynucleotide probes. See also Supplementary Information.

Könneke et al. 2005 Nature

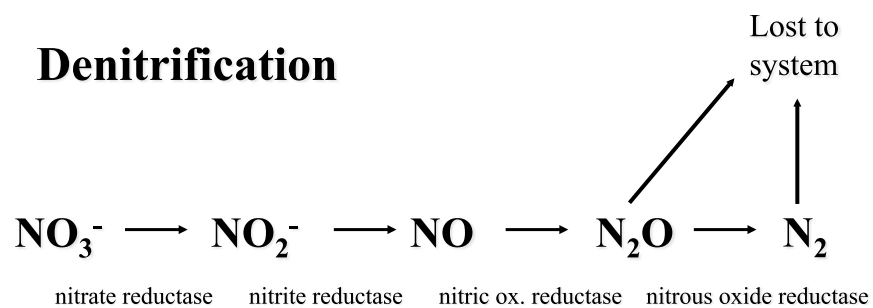
**Isolated a marine crenarchaeota that can
grow by aerobically oxidizing NH_4^+ to NO_2^- .**

Ingalls et al. 2006 PNAS

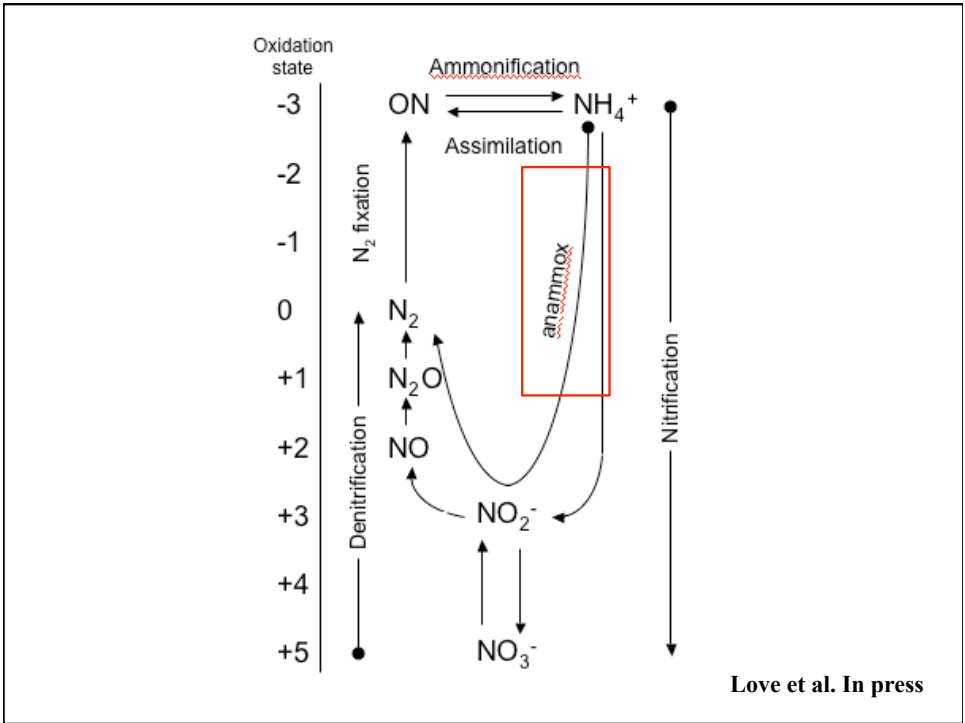
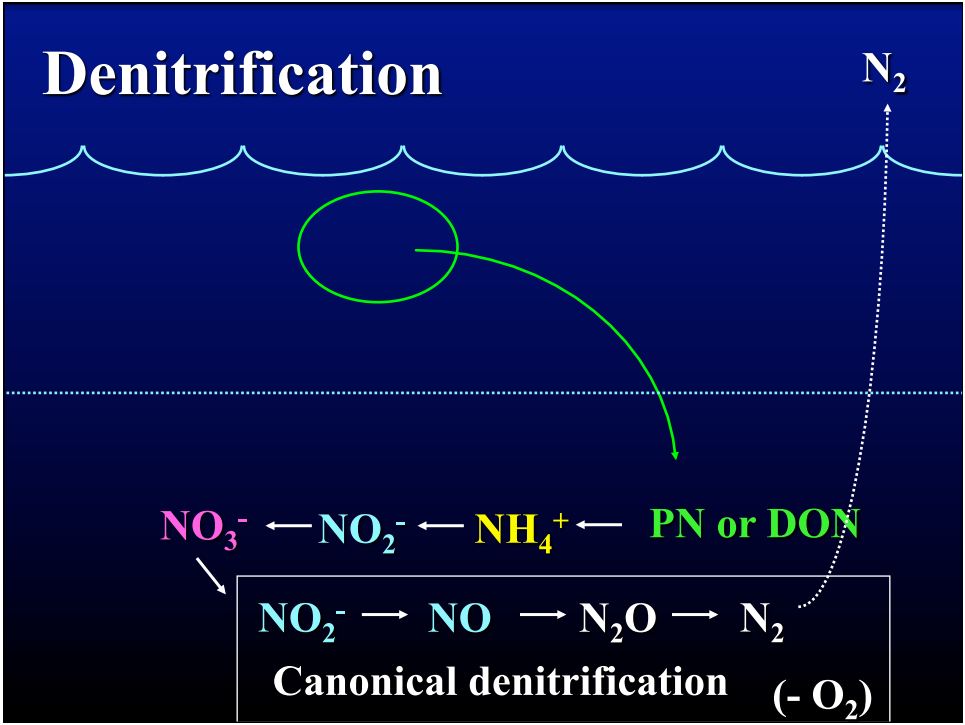
**An isotopic mass balance of radiocarbon
signatures of archaeal membrane lipids
indicates that 83% of their carbon is obtained
autotrophically at depth.**



Denitrification

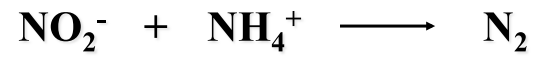


- N is used as an electron acceptor, not as a N source
- Lots of organisms can reduce NO_3^-
- Fewer can reduce NO_2^-
- All the enzymes are induced by anoxia
- NO is very labile and does not accumulate

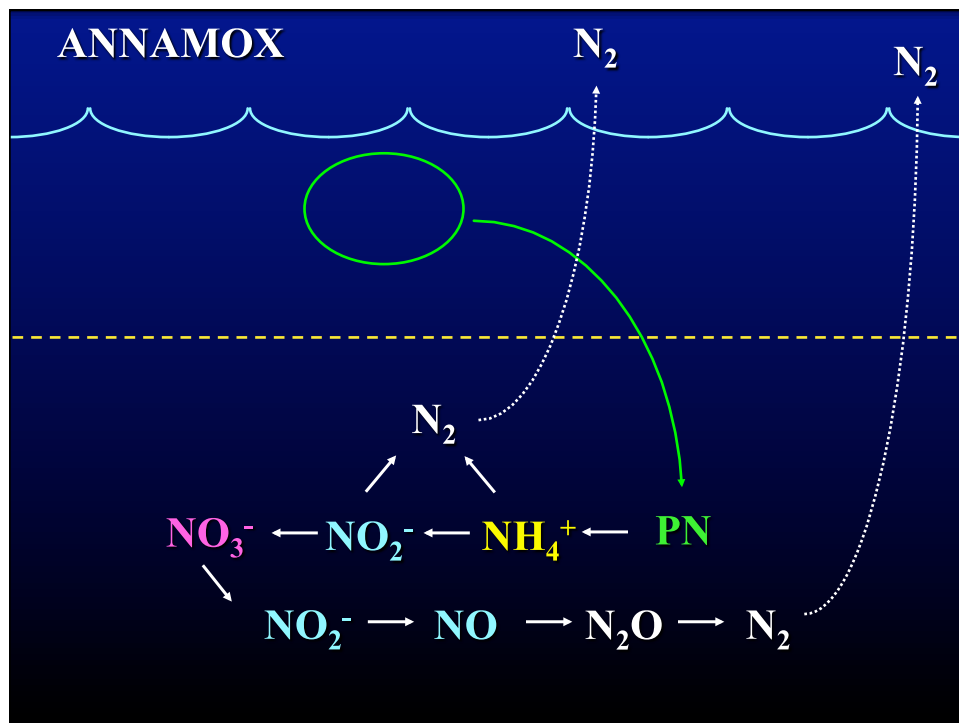


ANAMMOX

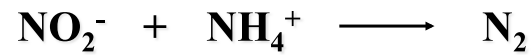
ANaerobic AMMonium OXidation



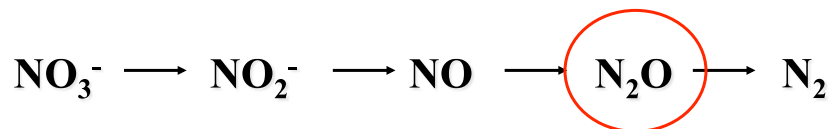
- First described in a wastewater treatment plant in the Netherlands in 1995.
- Oxygen inhibition is reversible.
- Thandrup & Dalsgaard (2002) published the first marine data (sediment study)



Canonical denitrification? OR Anammox??



obligate anaerobic autotrophs

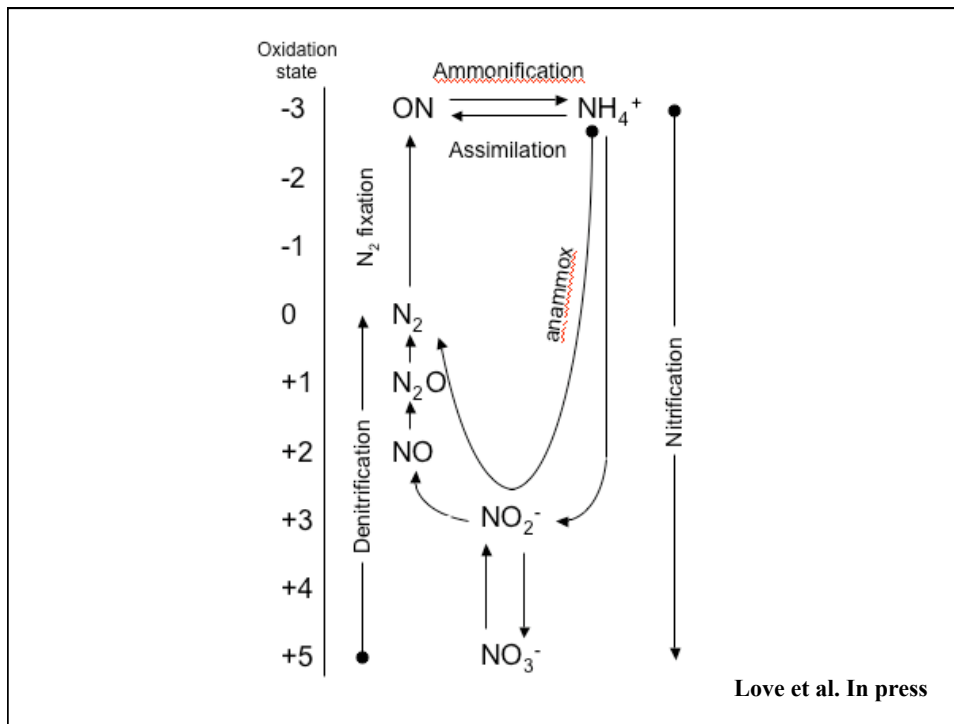


facultative anaerobic heterotrophs

Greenhouse Gases

The most abundant greenhouse gases
(in order of relative abundance)

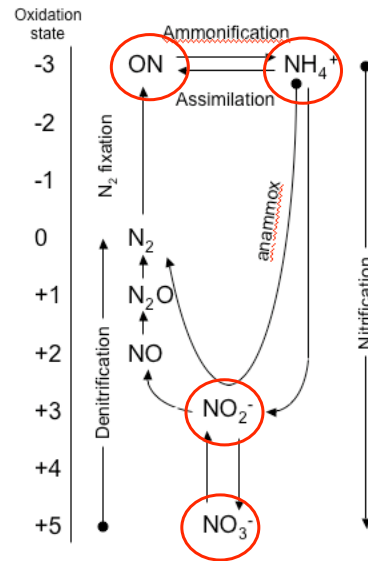
- water vapor
- CO₂
- methane
- N₂O
- ozone
- CFCs



What's the best way to make a living in a given place?

Reaction	Energy yield (kcal)
Aerobic respiration	686
Denitrification	545 (-O₂)
Nitrification NH₄⁺ oxidation	66
NO₂⁻ oxidation	17
N₂ fixation	-147

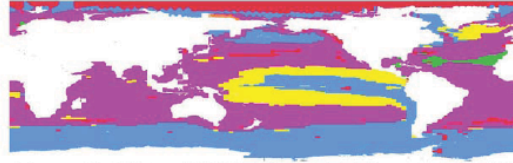
Uptake and regeneration in the surface ocean



Liebig's Law of the Minimum (1840) - the resource in smallest supply relative to what the organism needs is the limiting factor.

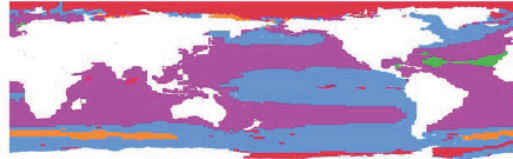
Growth versus biomass

A) Diatom Growth Limitation

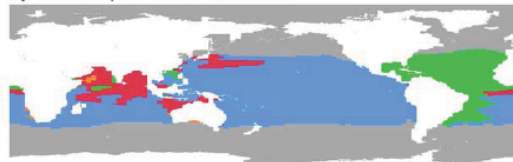


■ Nitrogen ■ Iron ■ Phosphorus ■ Silicon
■ Light ■ Temperature ■ Replete

B) Small Phytoplankton Growth Limitation



C) Diazotroph Growth Limitation



Factor limiting
growth rates
during summer

Moore et al. 2004 GBC

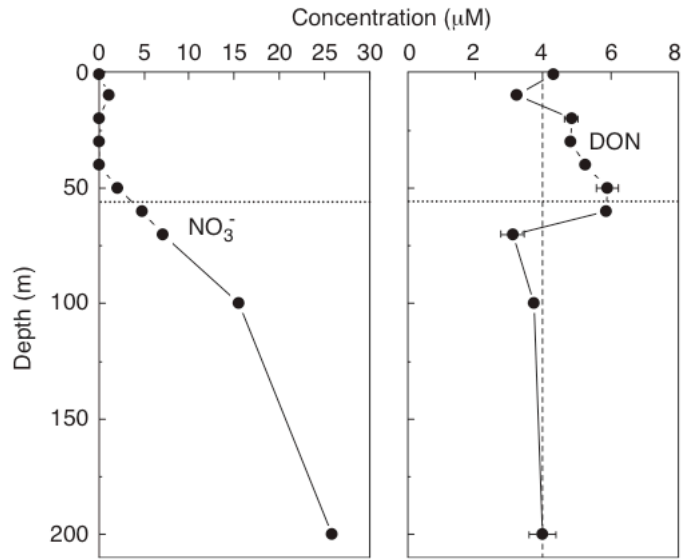
UPTAKE OF NEW AND REGENERATED FORMS OF NITROGEN IN PRIMARY PRODUCTIVITY¹

R. C. Dugdale and J. J. Goering
Institute of Marine Science, University of Alaska, College 99735

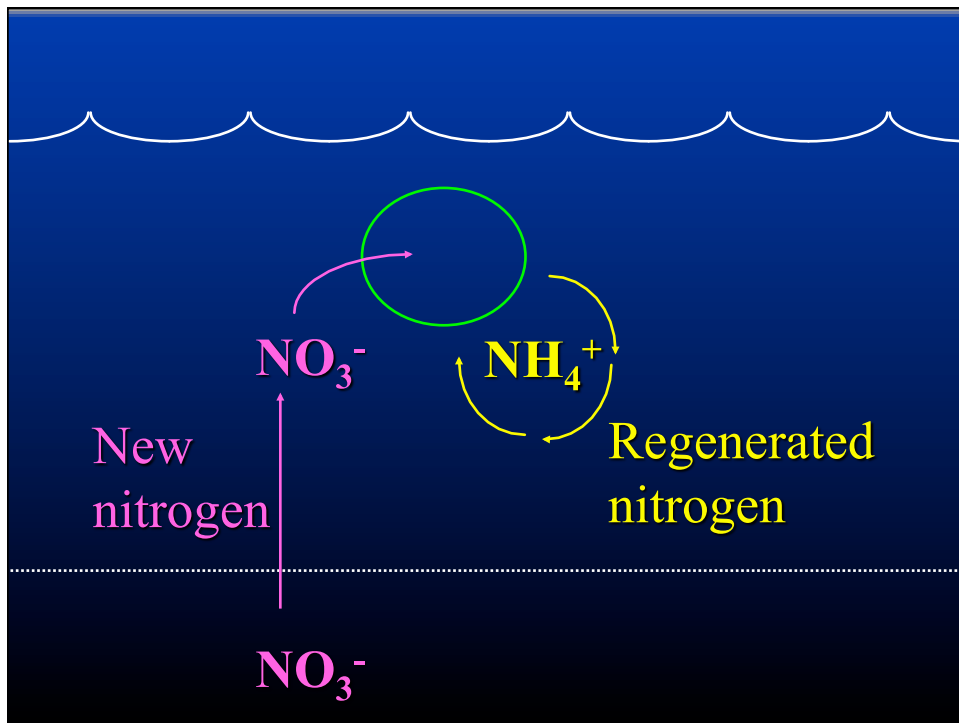
**Production can be defined as new or
regenerated based on the source of the
nitrogen that fueled it.**

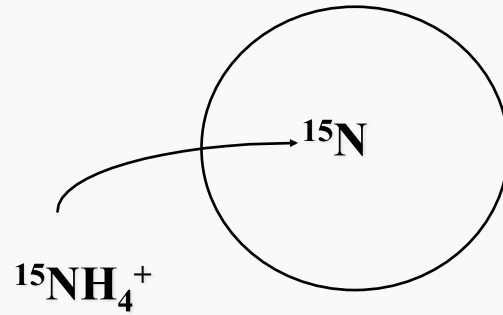
Dugdale & Goering 1967 L&O

Southern California Bight

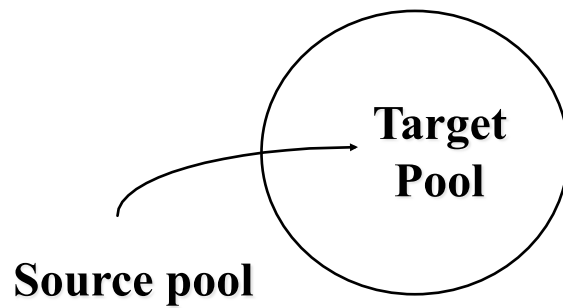


Bronk & Ward 2005 DSRI

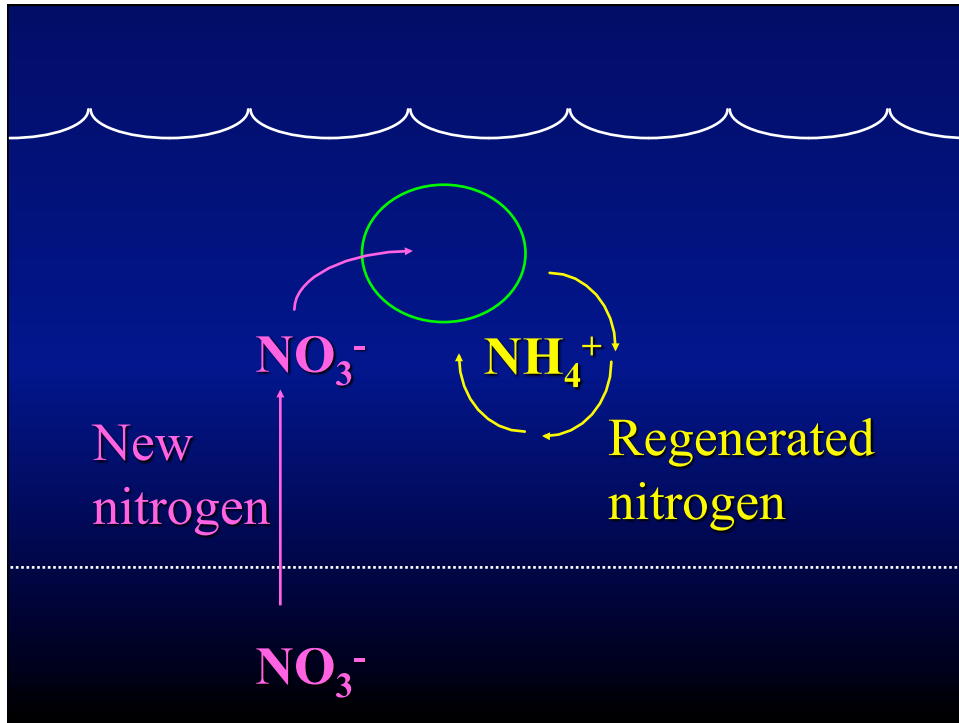




$$\text{Net Uptake Rate} = \frac{\text{atom\% PN}}{\text{atom \% NH}_4^+ \times \text{Time}} \times [\text{PN}]$$



$$\text{Rate} = \frac{\text{atom \% of target}}{\text{atom \% of source} \times \text{Time}} \times [\text{target}]$$



Particulate organic matter flux and planktonic new production in the deep ocean

Richard W. Eppley

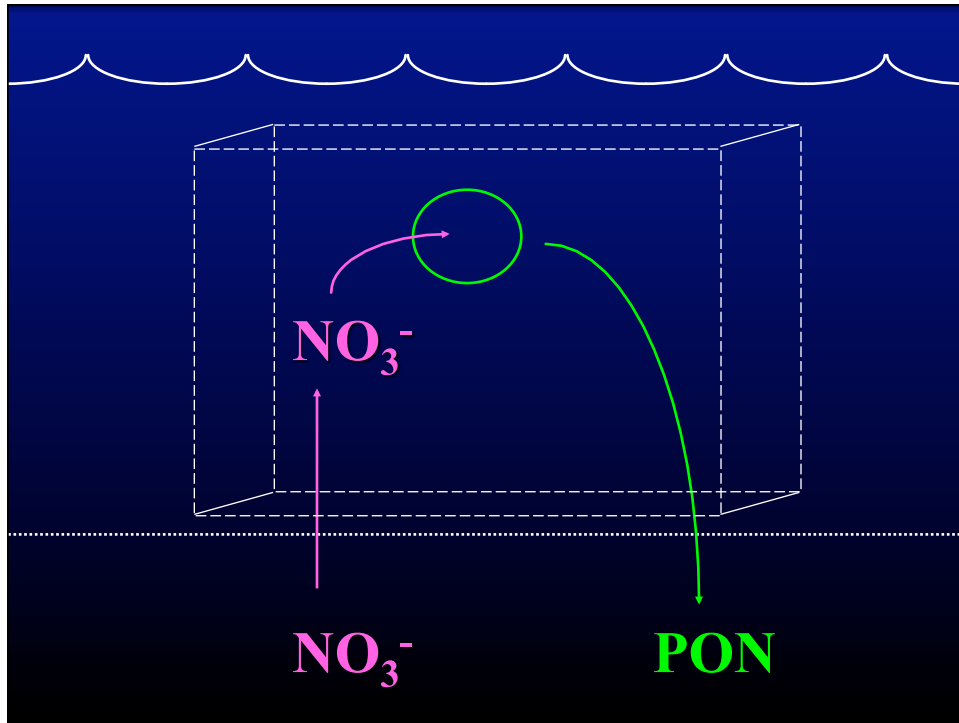
Institute of Marine Resources, A-018, Scripps Institution of Oceanography, University of California, San Diego, La Jolla, California 92093

Bruce J. Peterson

Ecosystems Center, Marine Biological Laboratory, Woods Hole, Massachusetts 02543

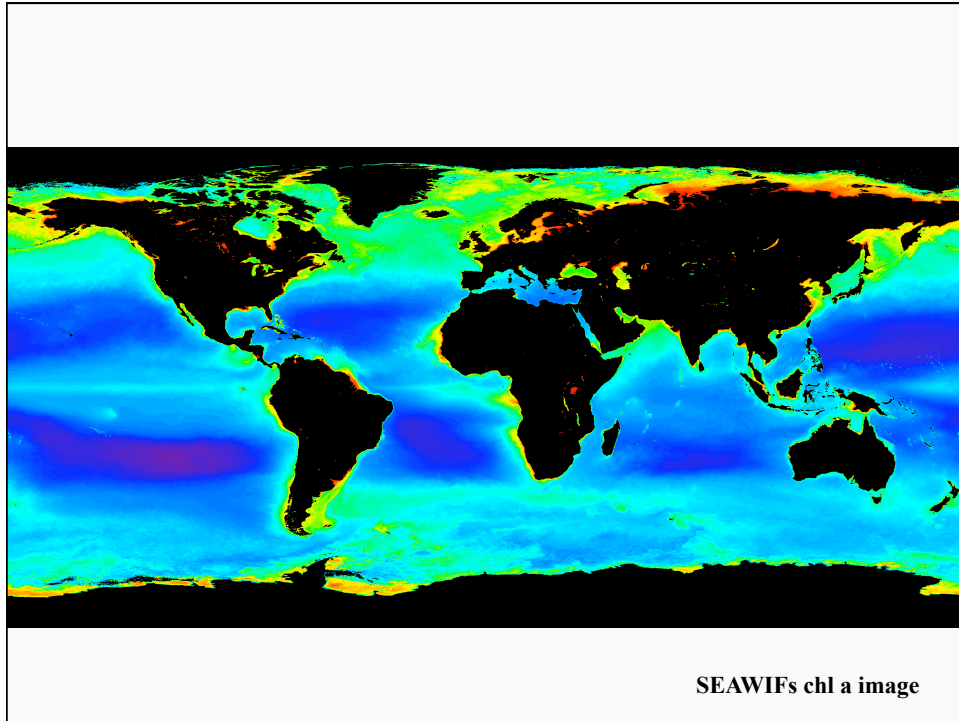
**New production over appropriate spatial
and temporal scales equals export flux.**

Eppley & Peterson 1979 Nature



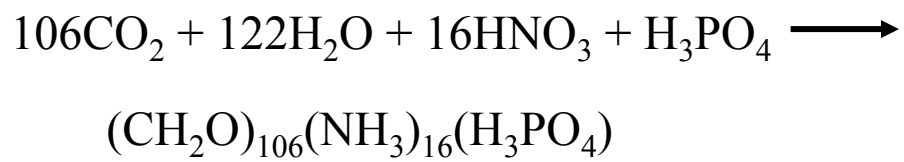
$$\text{f-ratio} = \frac{\text{New production}}{\text{New} + \text{Regenerated Production}}$$

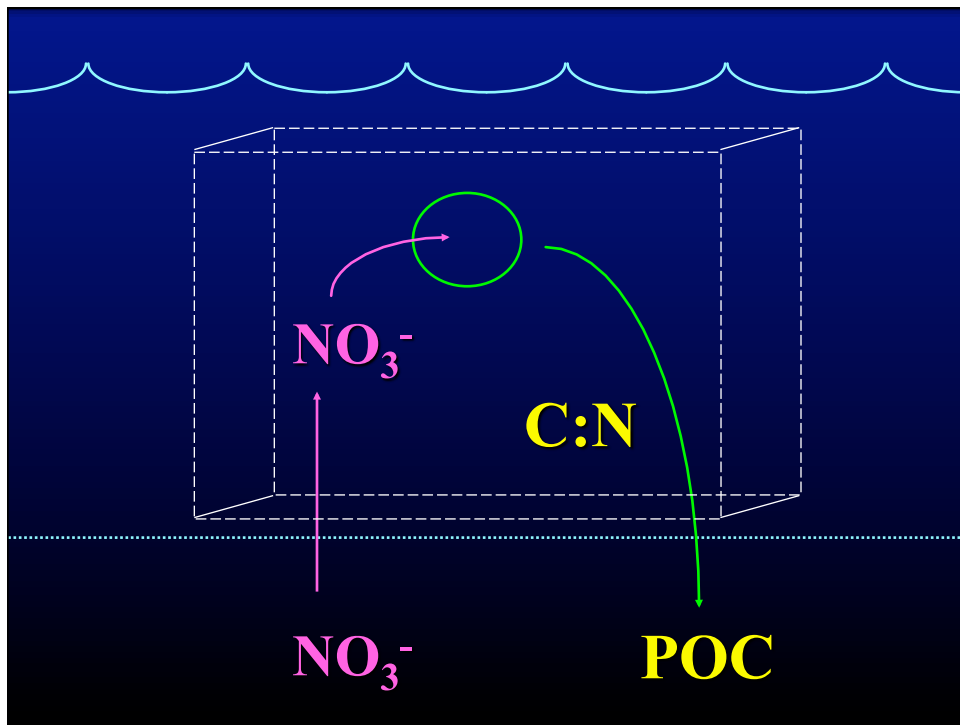
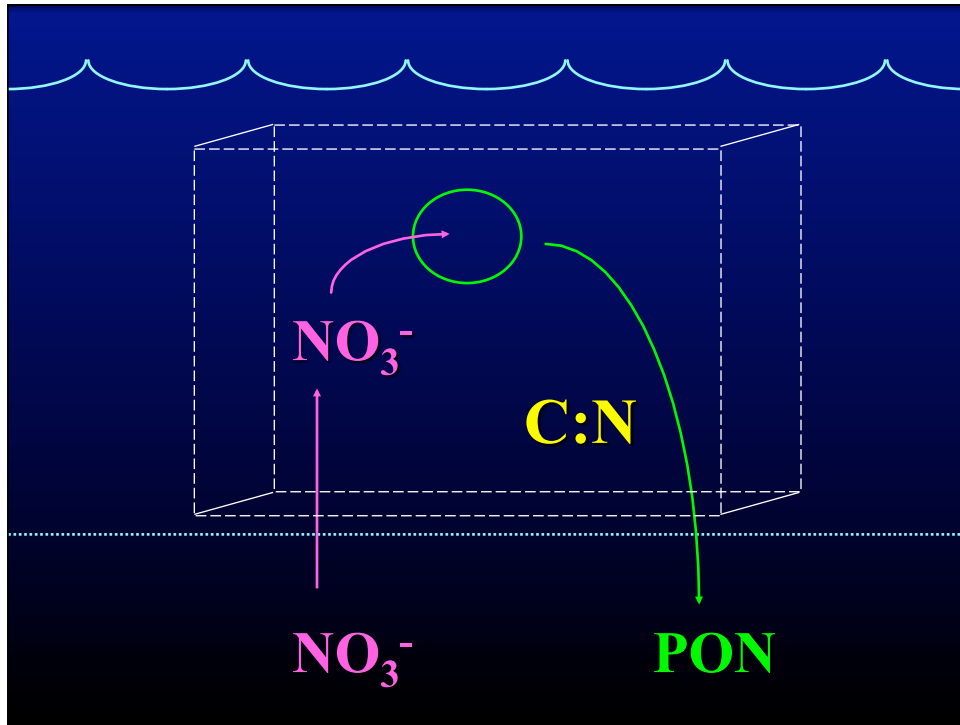
$$\text{f-ratio} = \frac{\text{NO}_3^- \text{ uptake}}{\text{NH}_4^+ + \text{NO}_3^- \text{ uptake}}$$

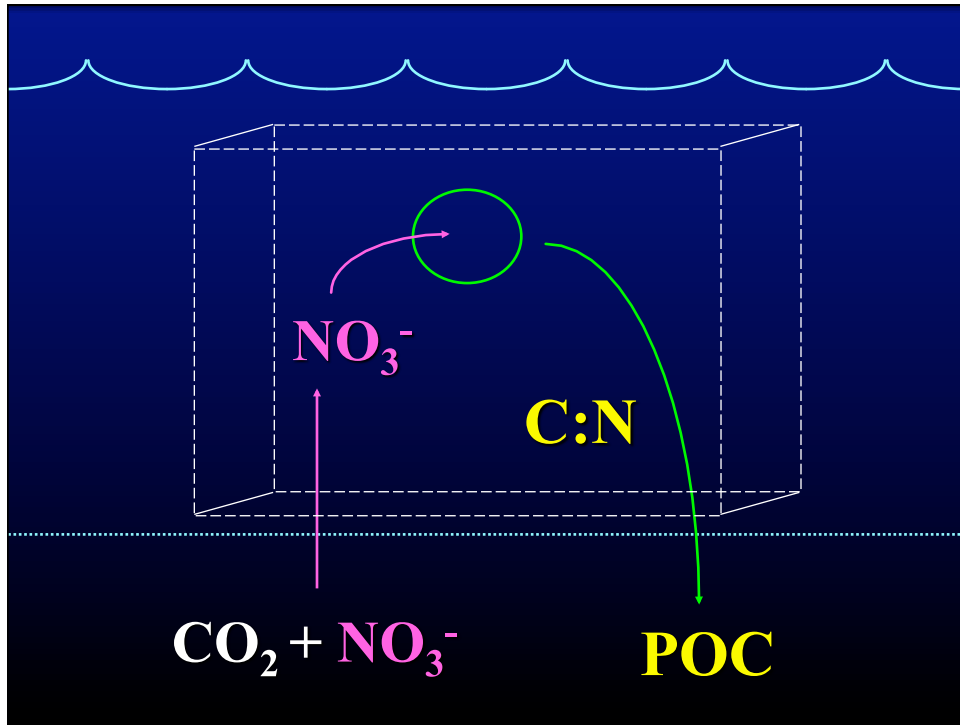


The Redfield Ratio

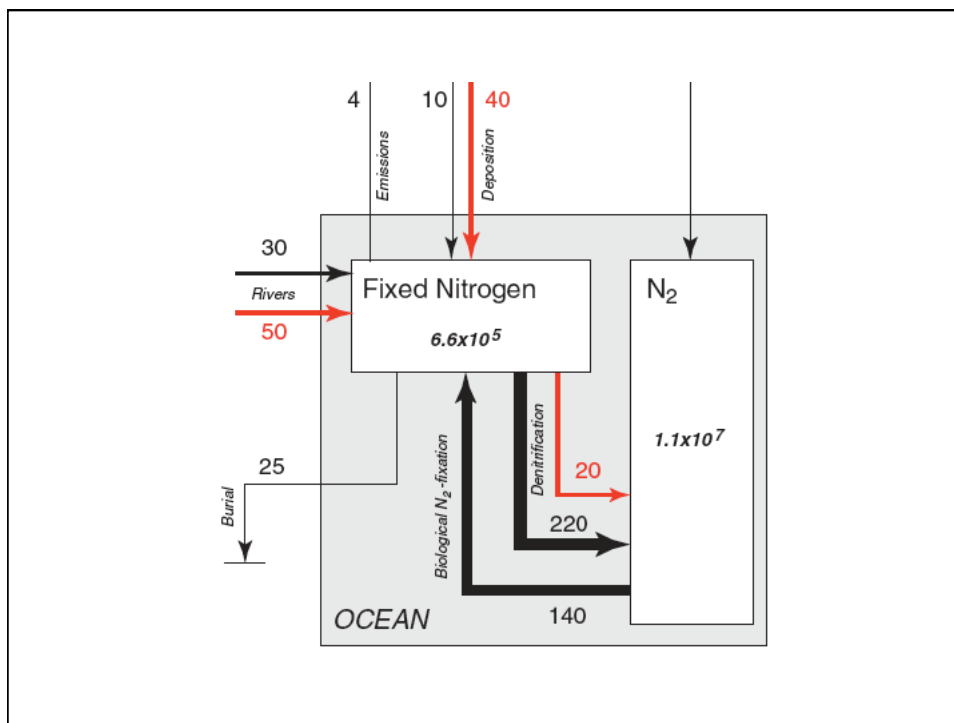
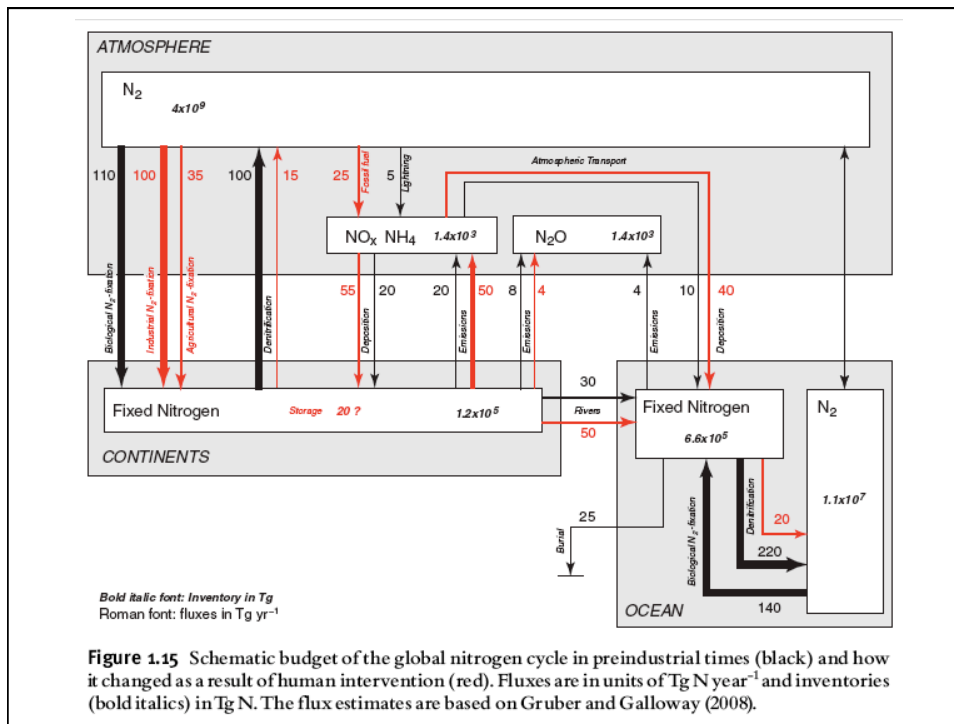
C:N:P = 106: 16: 1







**Is the amount of N
in the ocean in
steady state?**



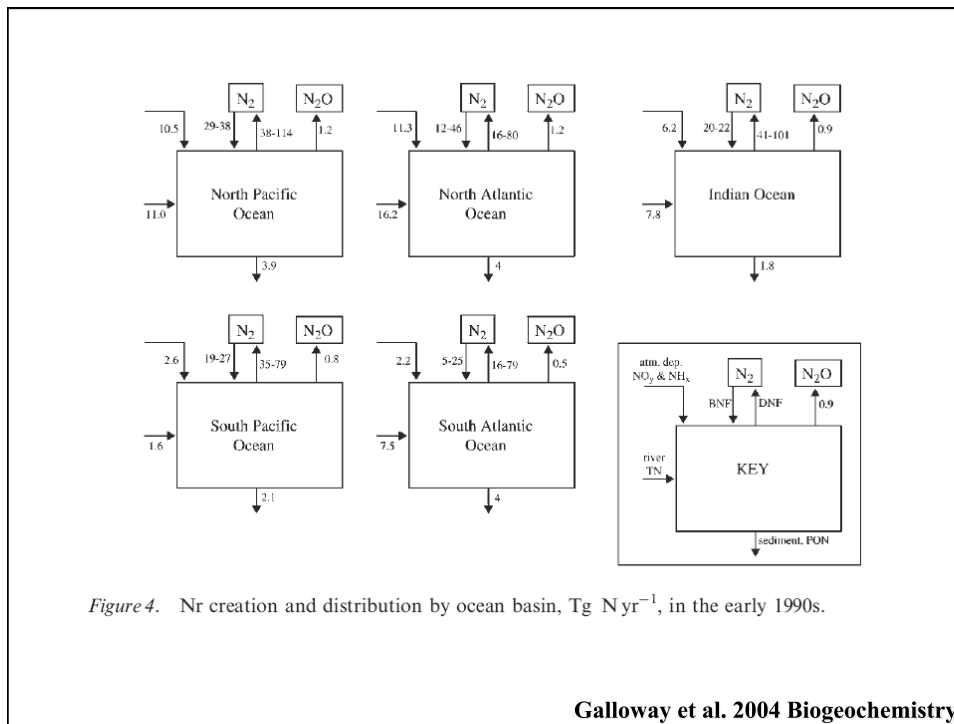


Table 1.3 Present-day (ca 1990) Global marine nitrogen budgets of Codispoti *et al.* (2001), Gruber (2004), and Galloway *et al.* (2004)

Process	Codispoti <i>et al.</i> ^a	Galloway <i>et al.</i> ^{a,b}	Gruber ^a
	2001	2004	2004
	Tg N year ⁻¹	Tg N year ⁻¹	Tg N year ⁻¹
<i>Sources</i>			
Pelagic N ₂ fixation	117	106	120 ± 50
Benthic N ₂ fixation	15	15	15 ± 10
River input (DON)	34	18 ^c	35 ± 10
River input (PON)	42	30 ^c	45 ± 10
Atmospheric deposition	86	33	50 ± 20
Total sources	294	202	265 ± 55
<i>Sinks</i>			
Organic N export	1		1
Benthic denitrification	300	206	180 ± 50
Water column denitrification	150	116	65 ± 20
Sediment Burial	25	16	25 ± 10
N ₂ O loss to atmosphere	6	4	4 ± 2
Total sinks	482	342	275 ± 55

^a See the original publications for details, e.g., Galloway *et al.* (2004), and Codispoti *et al.* (2001).
^b Listed are the central values reported by Galloway *et al.* (2004) (see Table 1.1 and Fig. 1.1 of their publication).
^c Galloway *et al.* (2004) lists only the total river flux. I assumed that about two thirds of the total is PON, and one third is DON.

Gruber 2008 N in the Marine Env.

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Tomorrow.....

