

Behavior, physiology and the niche of marine phytoplankton

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Halifax, Nova Scotia, Canada B3H 4J1

*Microbial Oceanography Summer Course:
Genomes to Biomes*

University of Hawai'i
June 27, 2008



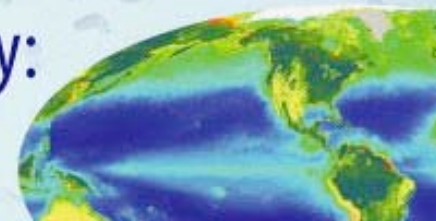
2008 HAWAII SUMMER COURSE ON MICROBIAL OCEANOGRAPHY

AGOURON
INSTITUTE



Microbial Oceanography:
Genomes to Biomes

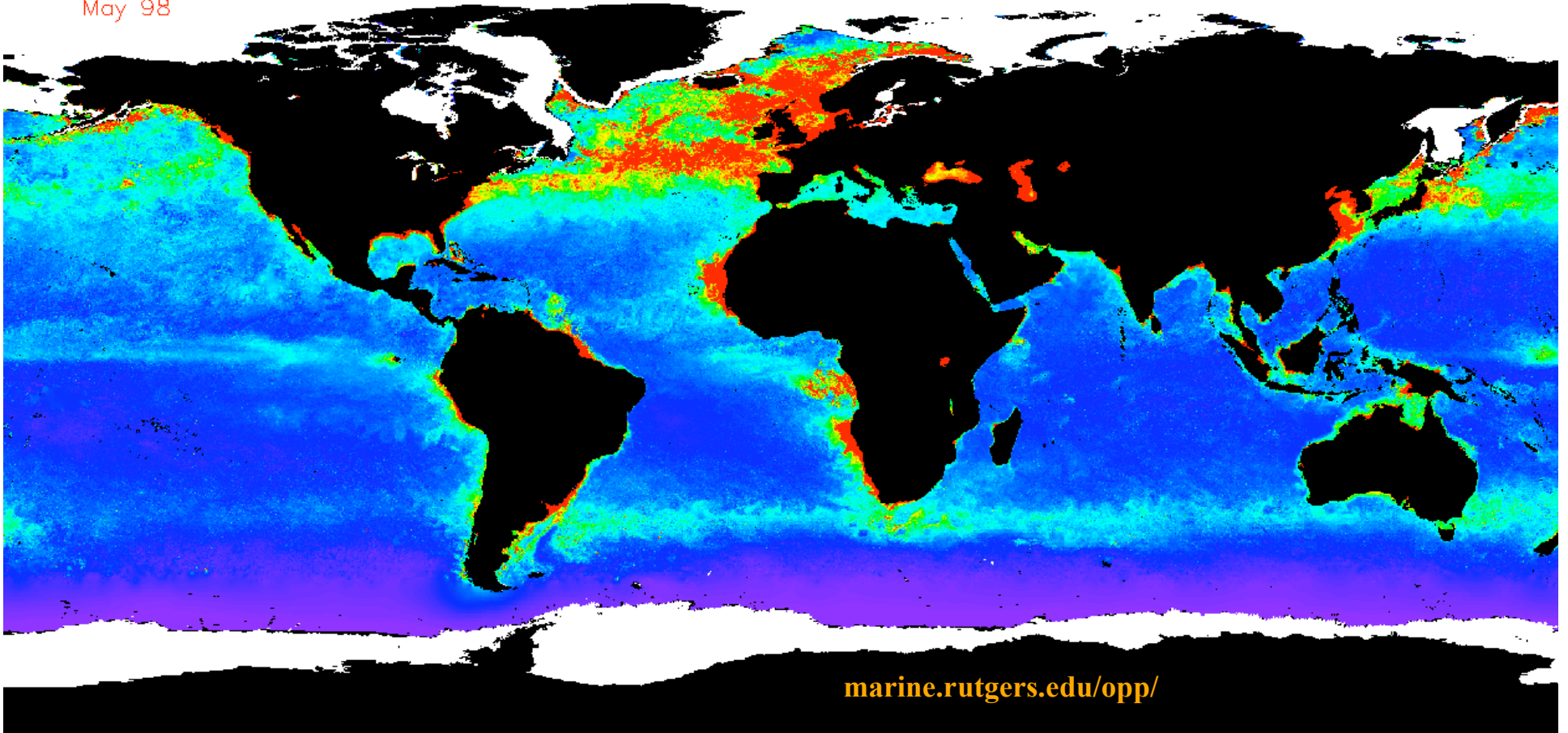
A laboratory-field training course at the University of Hawai'i at Mānoa



A principal goal of microbial oceanography

Describing and explaining the
distributions and *activities* of marine microbes

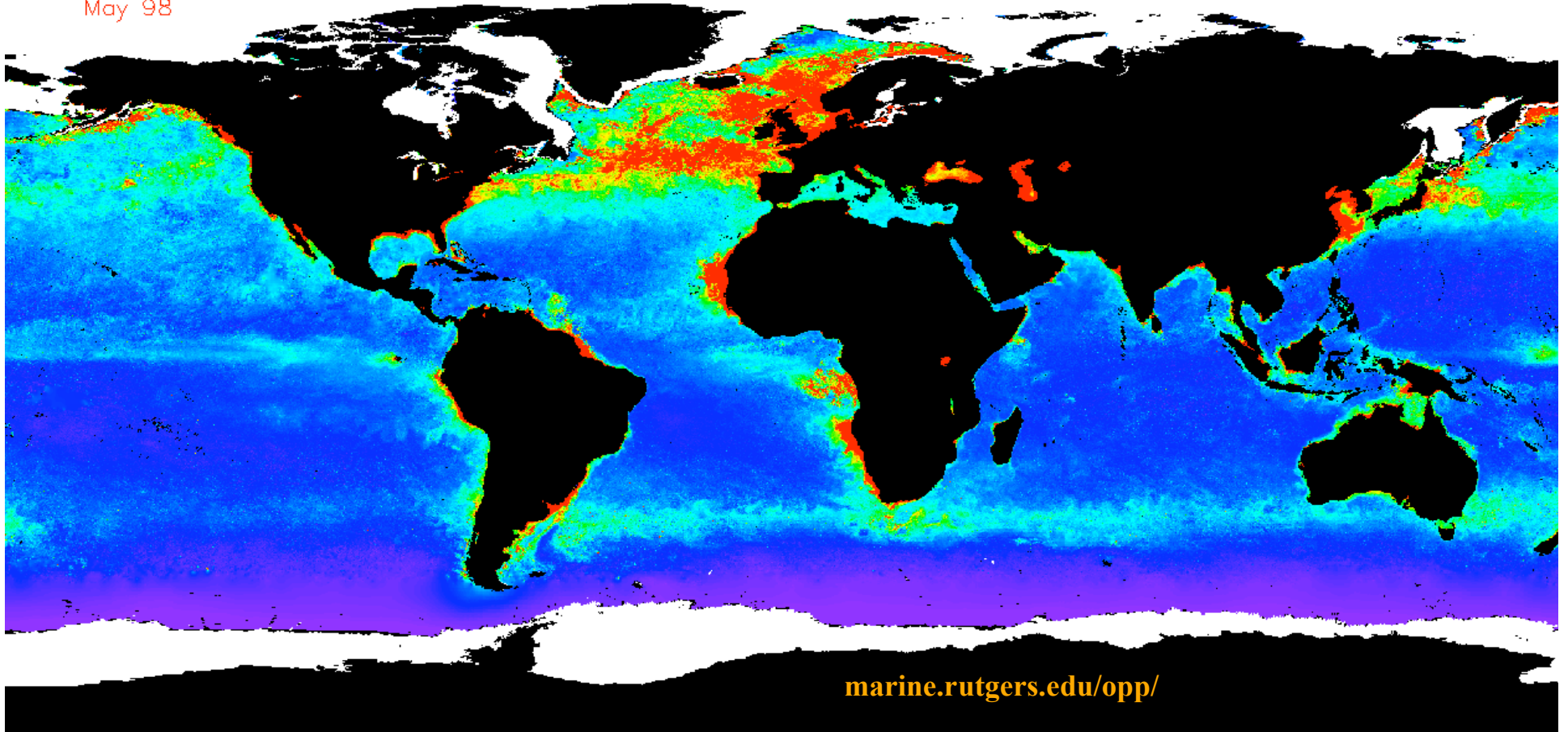
May 98



marine.rutgers.edu/opp/

...and using this information to describe the *causes* and *consequences* of variations in key biogeochemical processes

May 98

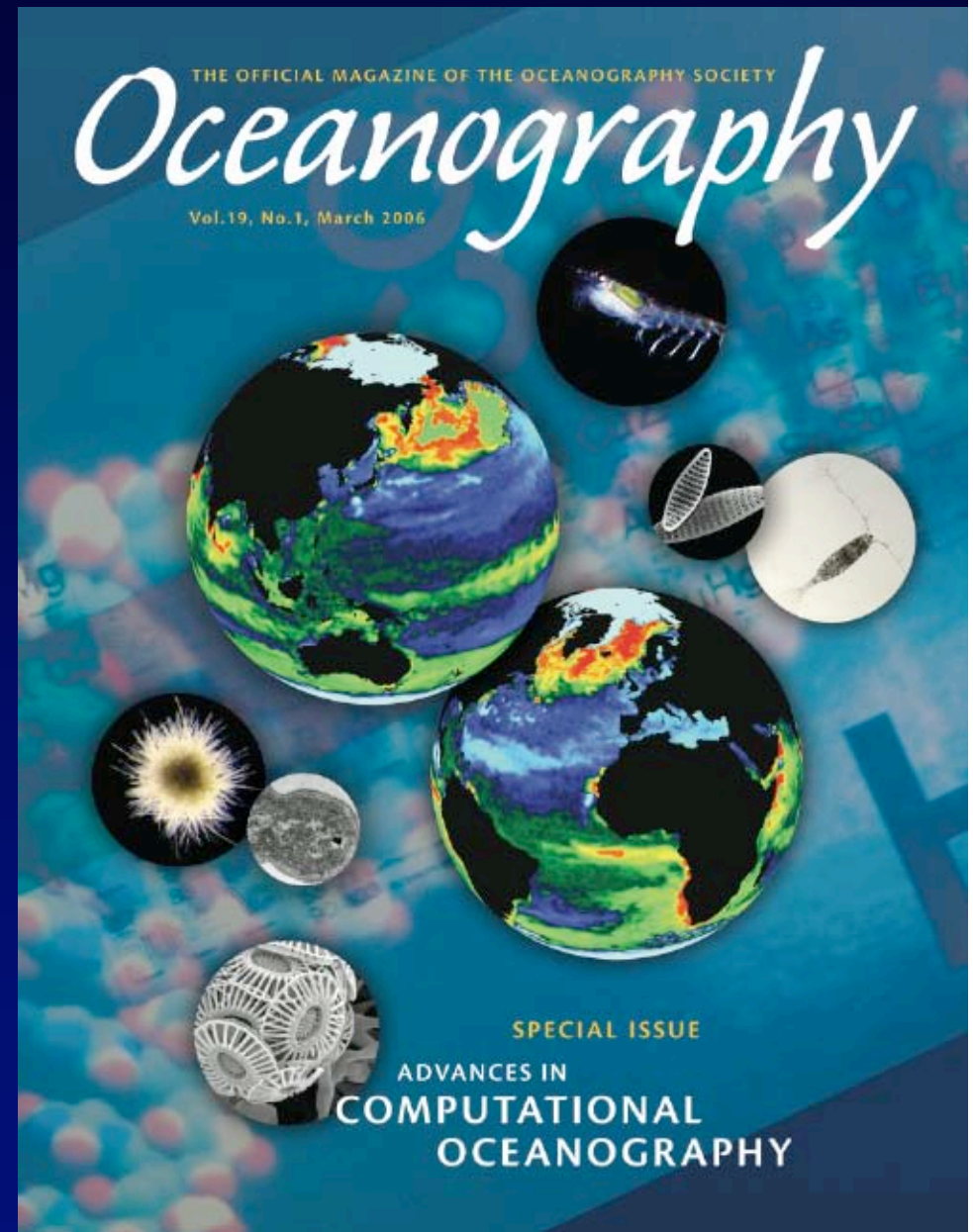


Key biogeochemical processes

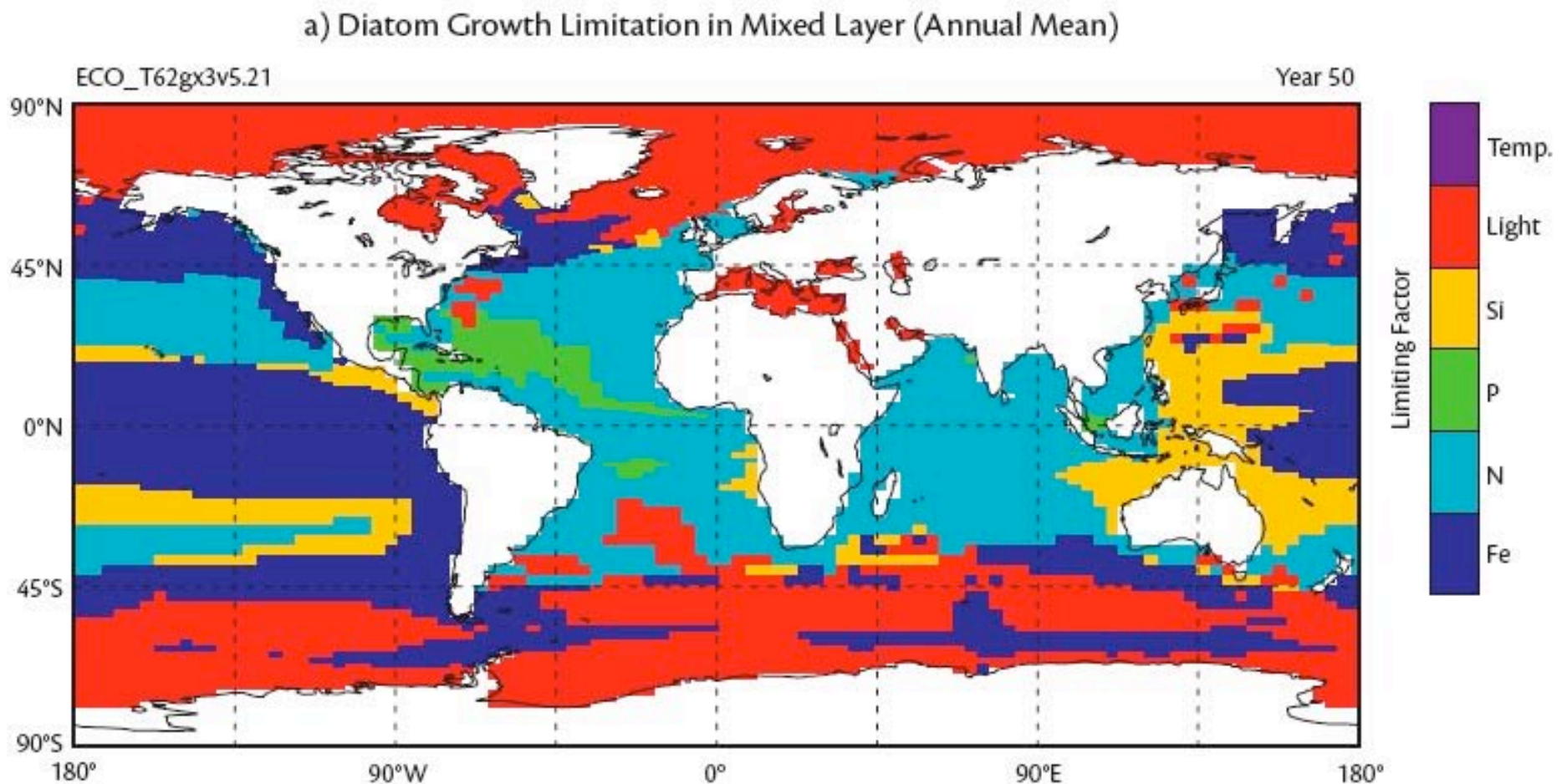
- Primary production
- Nitrogen fixation
- Denitrification
- Trace gas production
- ...and the many other processes that make those processes possible

This can be achieved
only through an
integrated approach

*The role of the oceans in
Earth systems ecology,
and the effects of climate
variability on the ocean
and its ecosystems, can be
understood only by
observing, describing, and
ultimately predicting the
state of the ocean as a
physically forced
ecological and
biogeochemical system.*



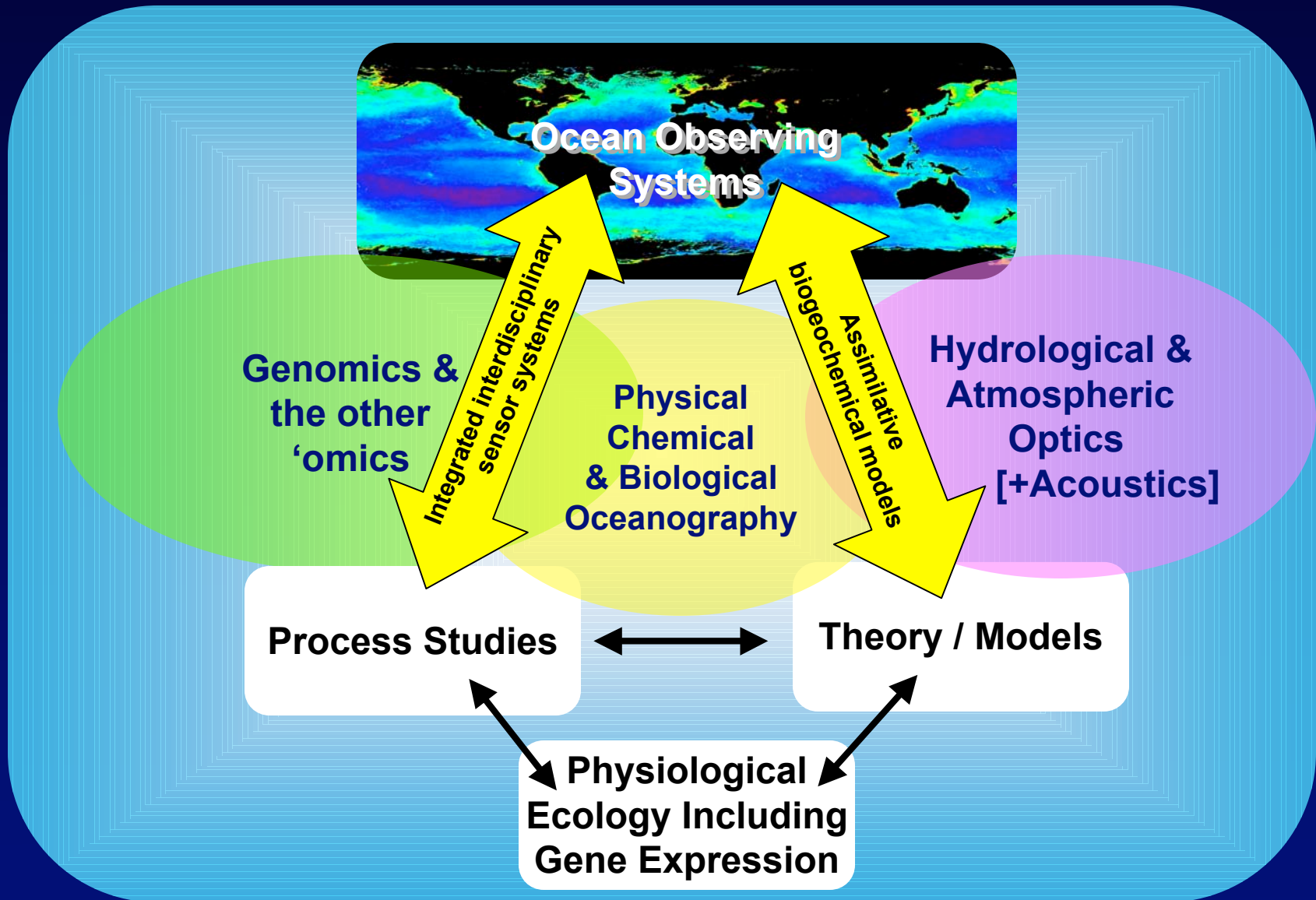
Arguably, this represents the state of the art



PARADIGM Global Biogeochemistry - Ecology - Circulation model (Doney and colleagues)

Rothstein et al. 2006 – Oceanography Magazine

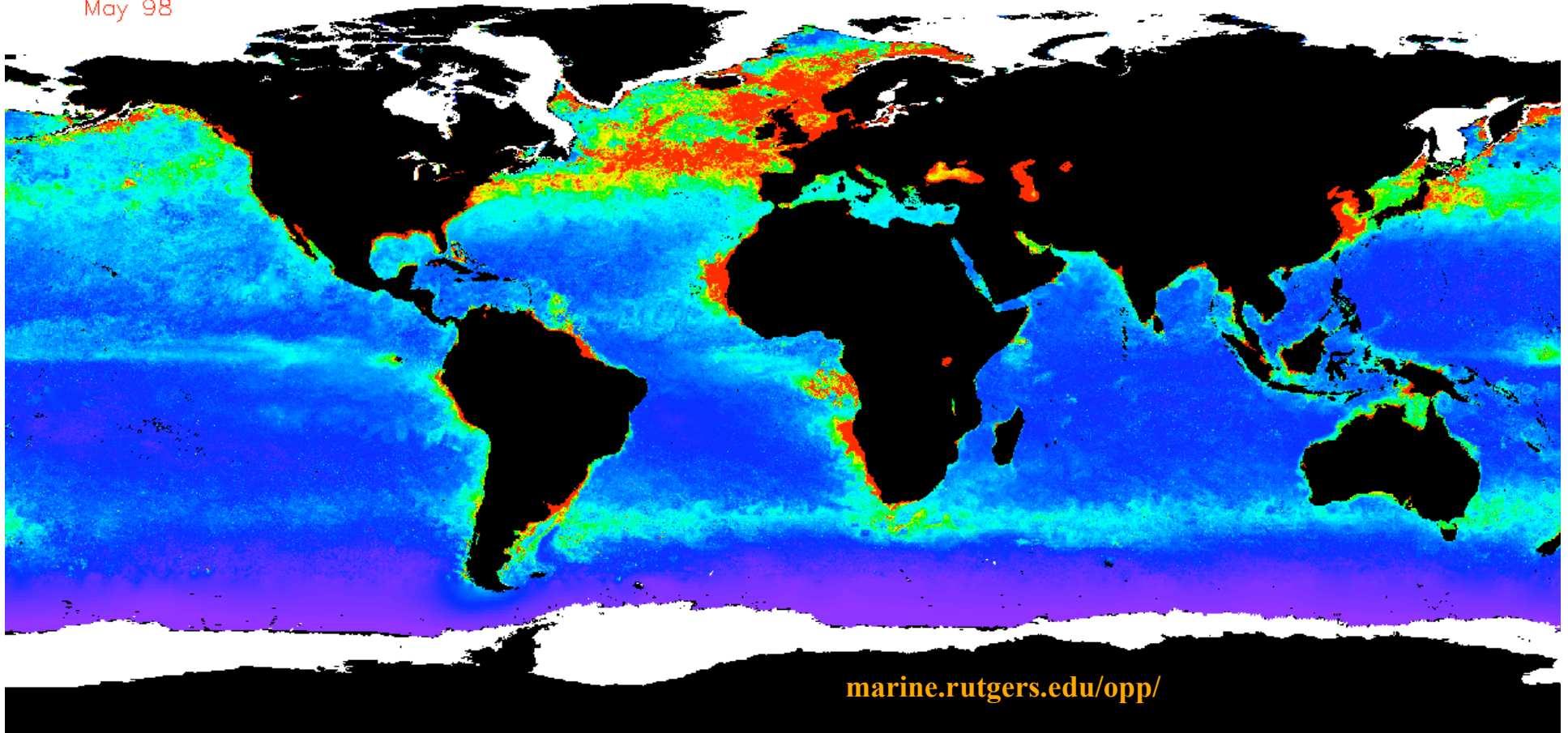
Ultimate Target



Biological oceanography and phytoplankton ecology

Describe the *causes* and *consequences* of variations in primary productivity (and food web structure)

May 98



An overview of established approaches to marine prediction

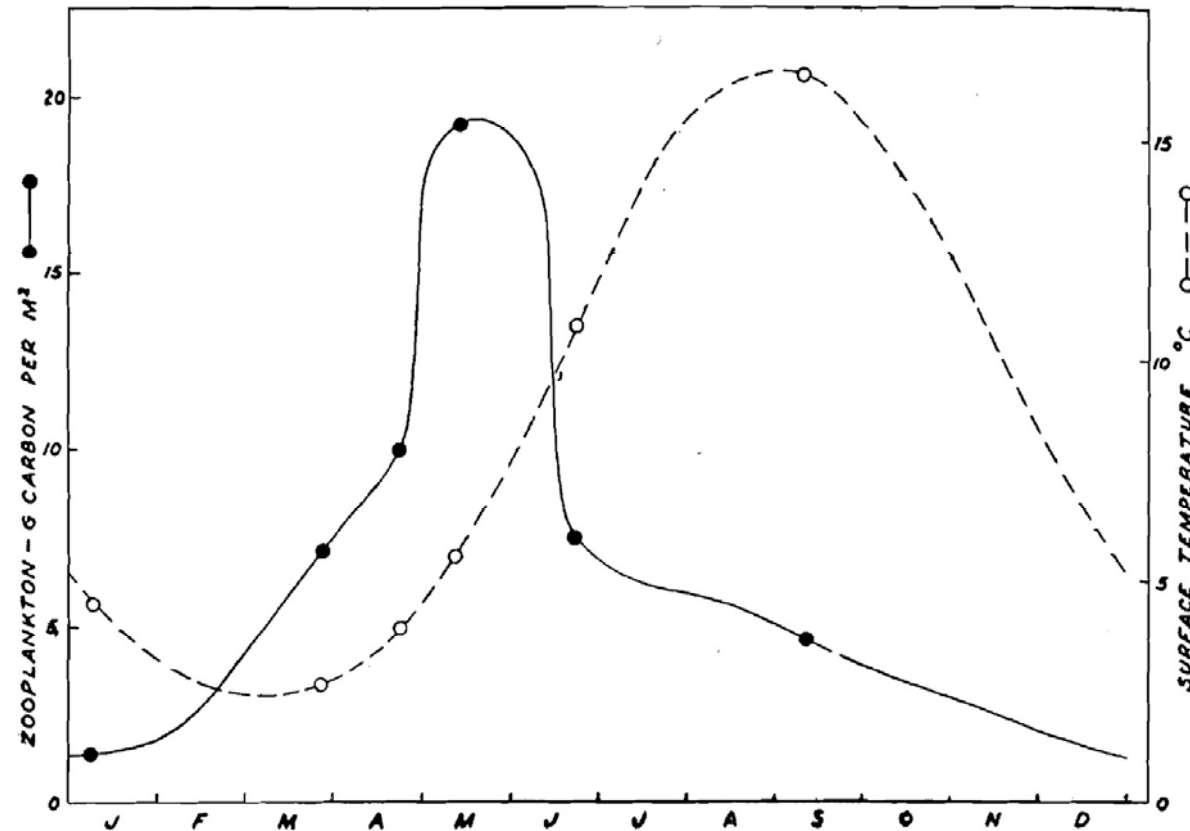
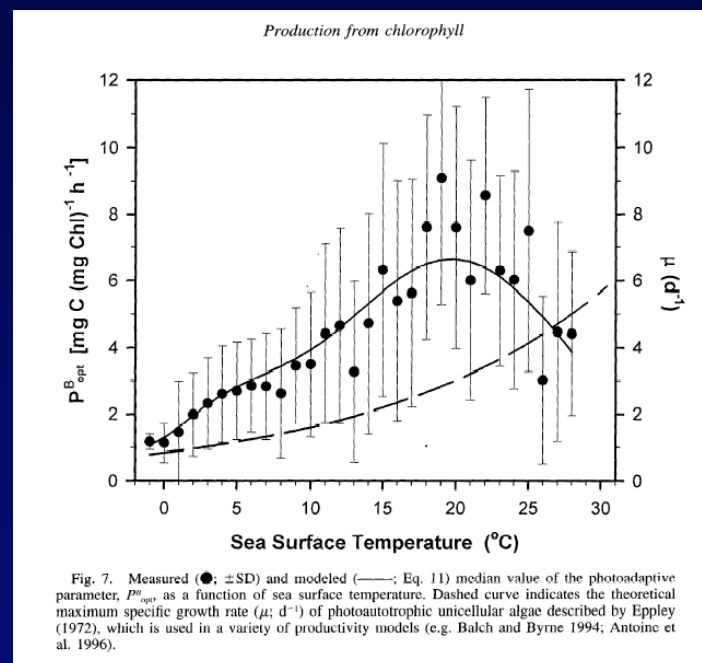
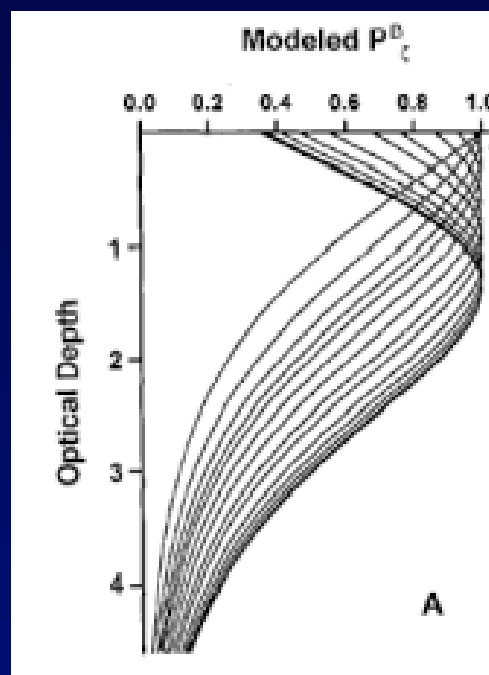
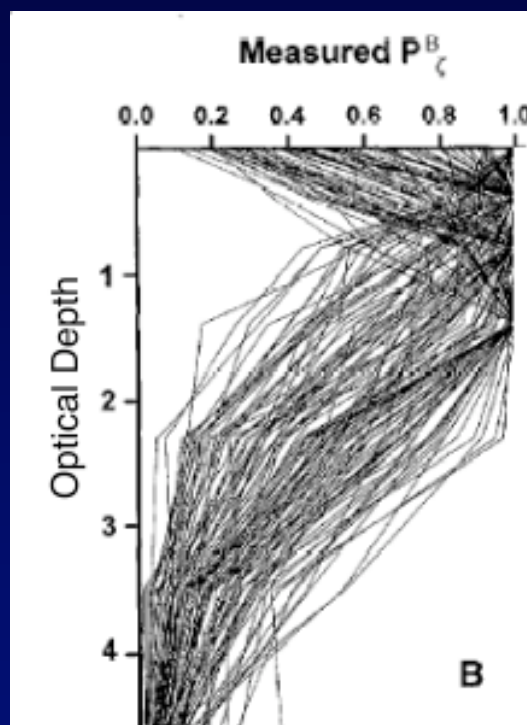


Figure 19. Solid line is the seasonal cycle of zooplankton. Measurements of zooplankton volume by the displacement method are treated by a conversion factor (wt. in g. = 12.5% \times vol. in cc.) to derive a rough estimate of the carbon content. Dotted line is the mean surface temperature.

Approach #1: Observation, analysis, inference (empirical, diagnostic models)



Behrenfeld and Falkowski 1997b L&O

Modeling the pattern in the measurements — not necessarily primary productivity!

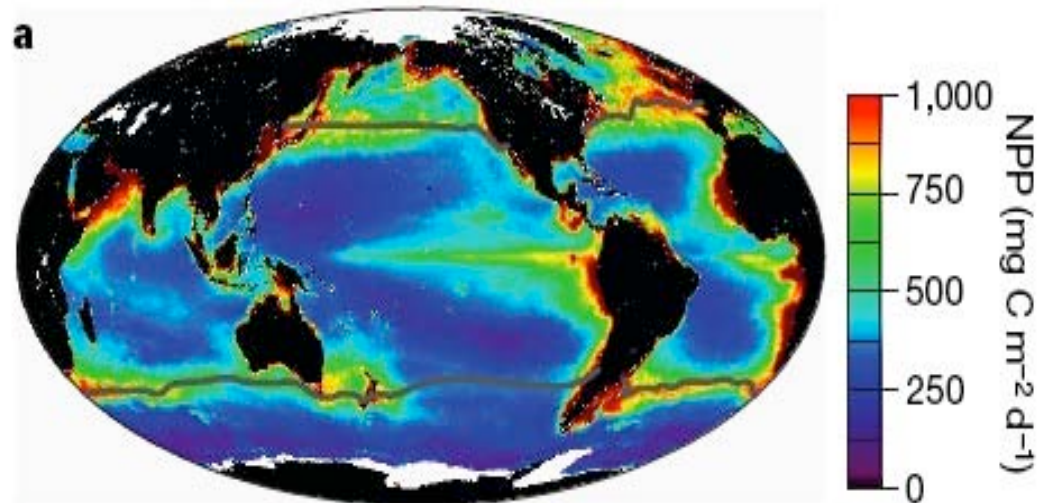
Result: Quantitative predictions that are as good as the data & assumptions that go into them

nature

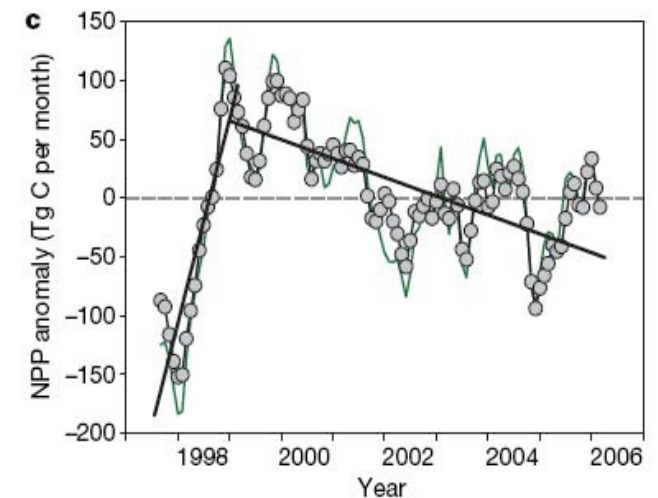
Vol 444 | 7 December 2006 | doi:10.1038/nature05317

Climate-driven trends in contemporary ocean productivity

Michael J. Behrenfeld¹, Robert T. O'Malley¹, David A. Siegel³, Charles R. McClain⁴, Jorge L. Sarmiento⁵, Gene C. Feldman⁴, Allen J. Milligan¹, Paul G. Falkowski⁶, Ricardo M. Letelier² & Emmanuel S. Boss⁷

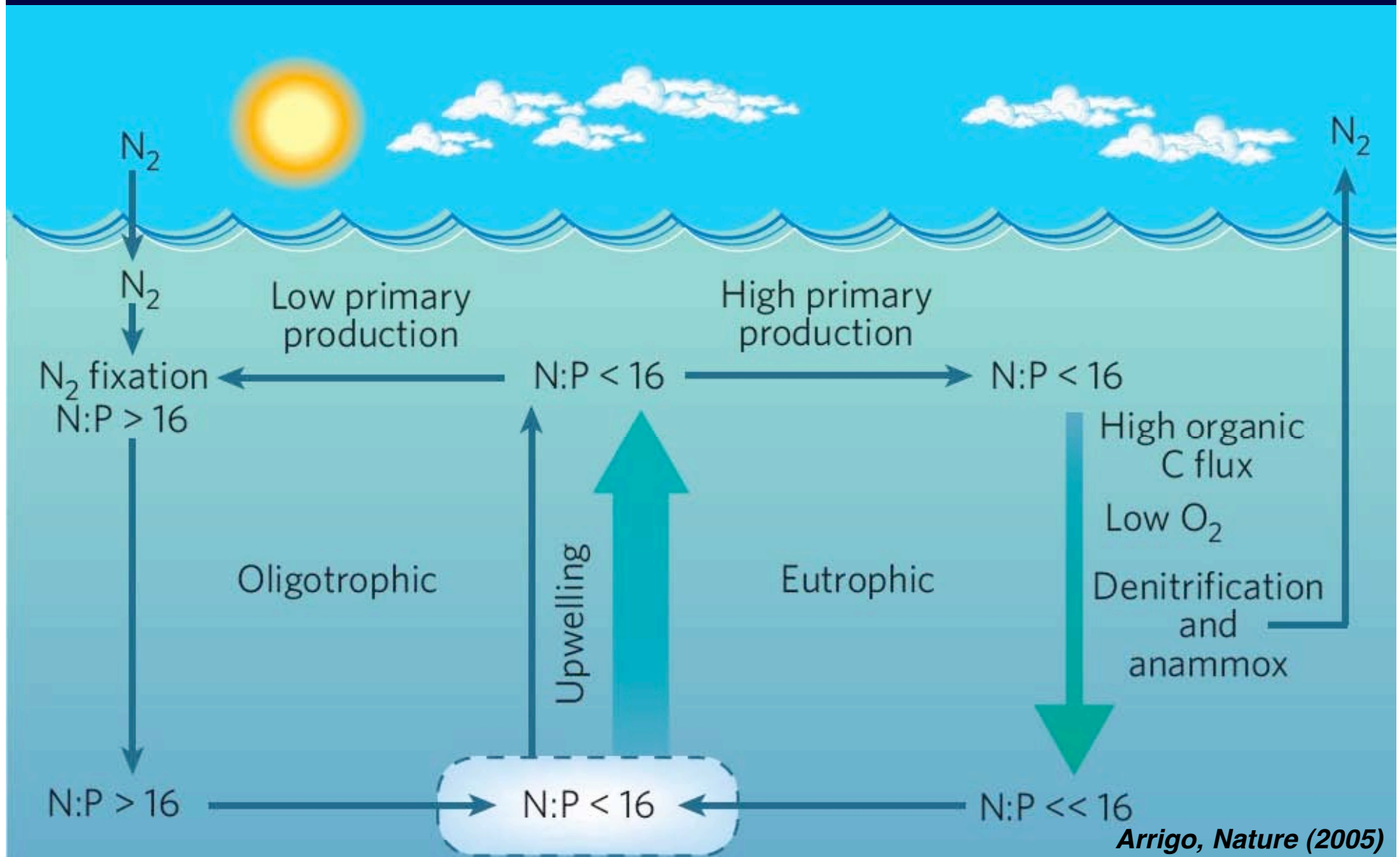


*Inputs/Assumptions of the
Productivity Model(s)*



Validity of the Statistical Analysis

Approach #2: Observation, analysis, inference (qualitative, mechanistic, predictive models)



Arrigo, Nature (2005)

The testing of qualitative, mechanistic, predictive models may be messy

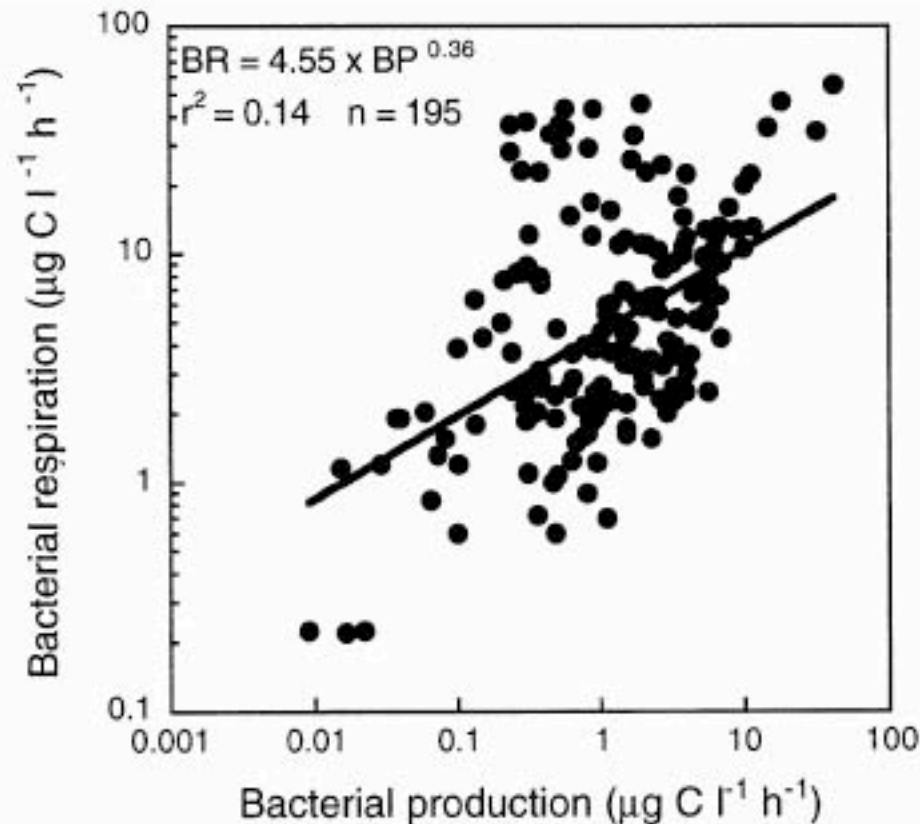


Figure 4. Bacterial respiration as a function of bacterial production in aquatic ecosystems. The data are paired observations of bacterial respiration (*BR*) and production (*BP*); the sources of these data appear in Table 1. The line is the least-squares fit to the log-transformed data.

del Giorgio and Cole (1999)

But predictions can be tested with appropriate observations

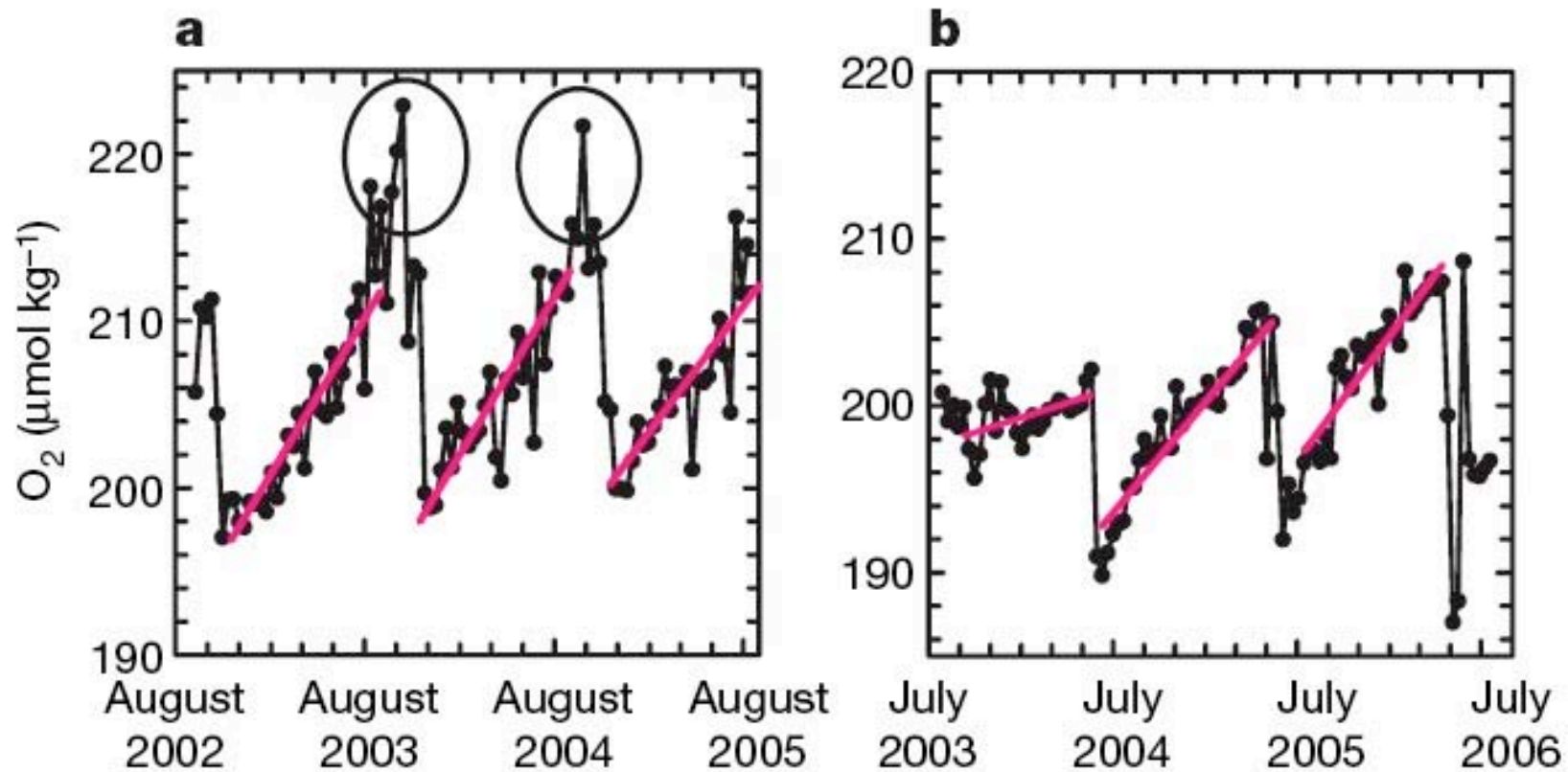


Figure 3 | Oxygen concentrations in the SOM versus time. Oxygen

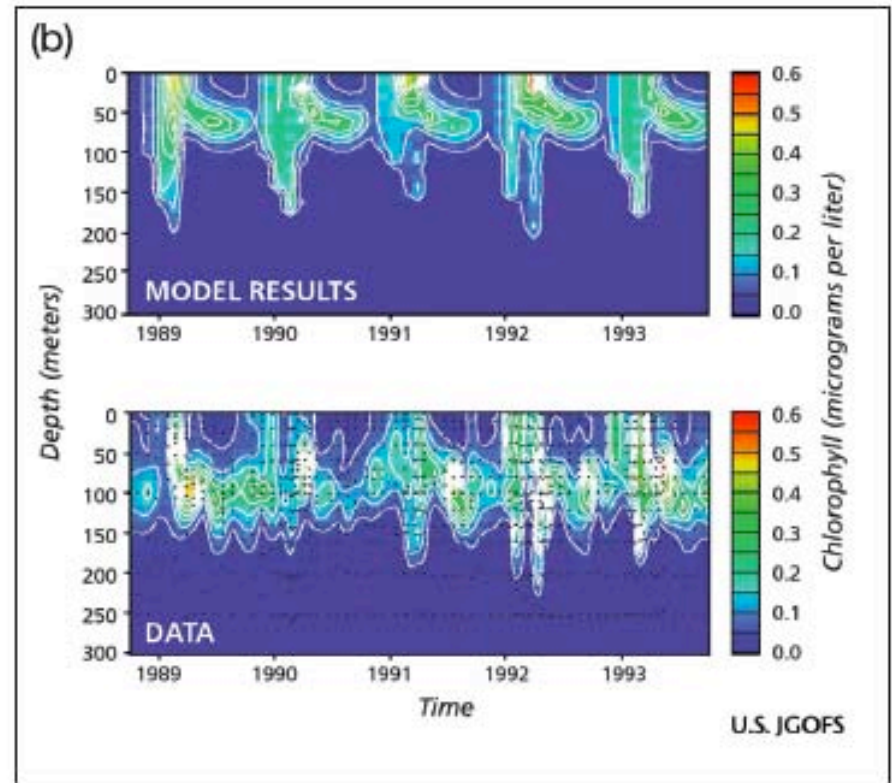
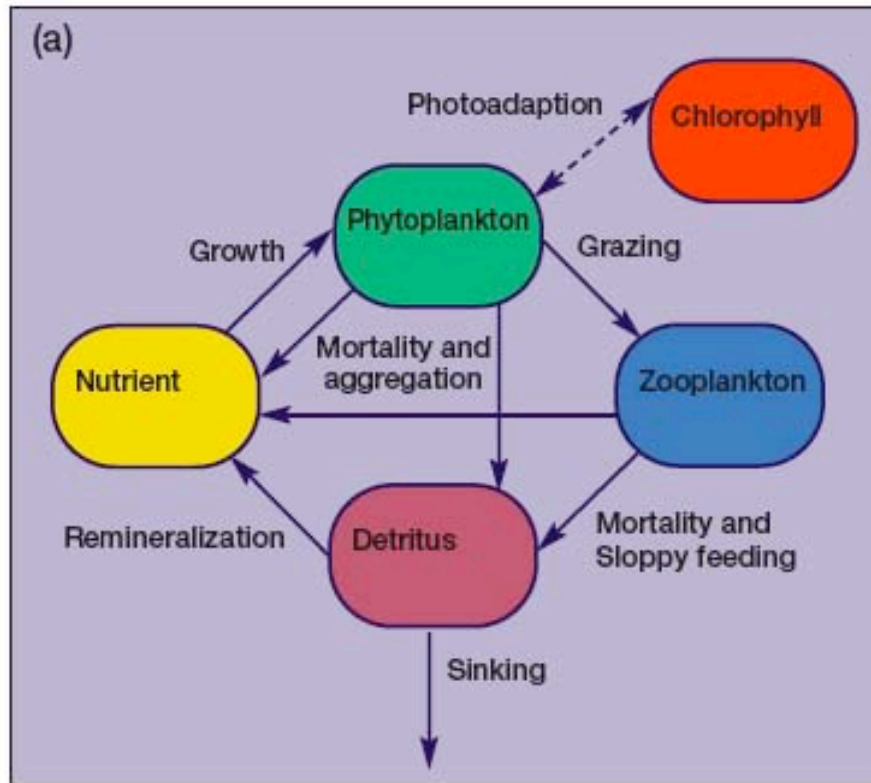
Riser and Johnson Nature 2008

John Cullen: Agouron Institute 2008

Approach #3: Prognostic models (quantitative, mechanistic, predictive models)

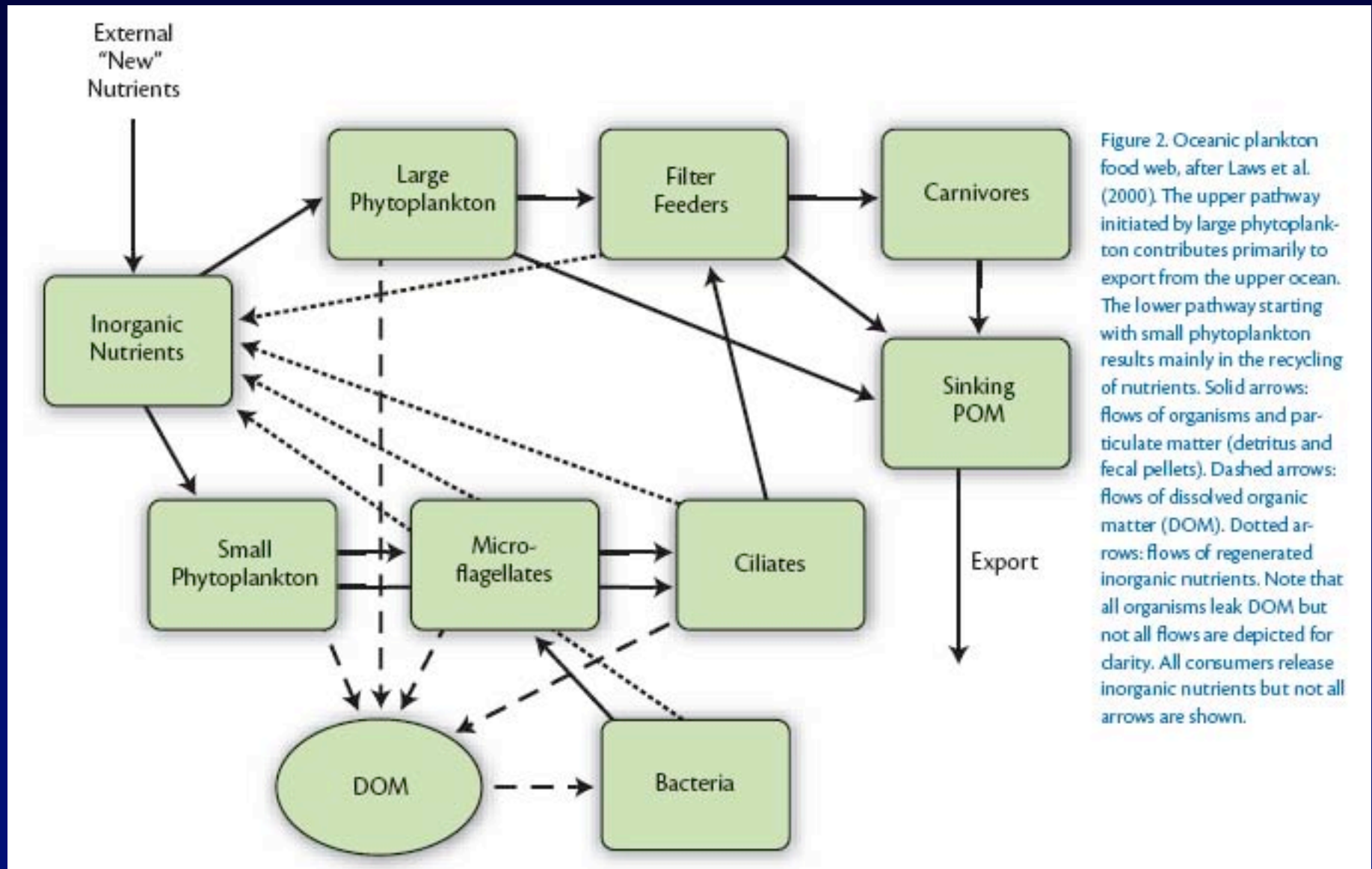
Ocean genomics

SC Doney et al.



Doney, S. C., M. R. Abbott, J. J. Cullen, D. M. Karl, and L. Rothstein. 2004. From genes to ecosystems: the ocean's new frontier. *Frontiers in Ecology and the Environment* 2: 457-466.

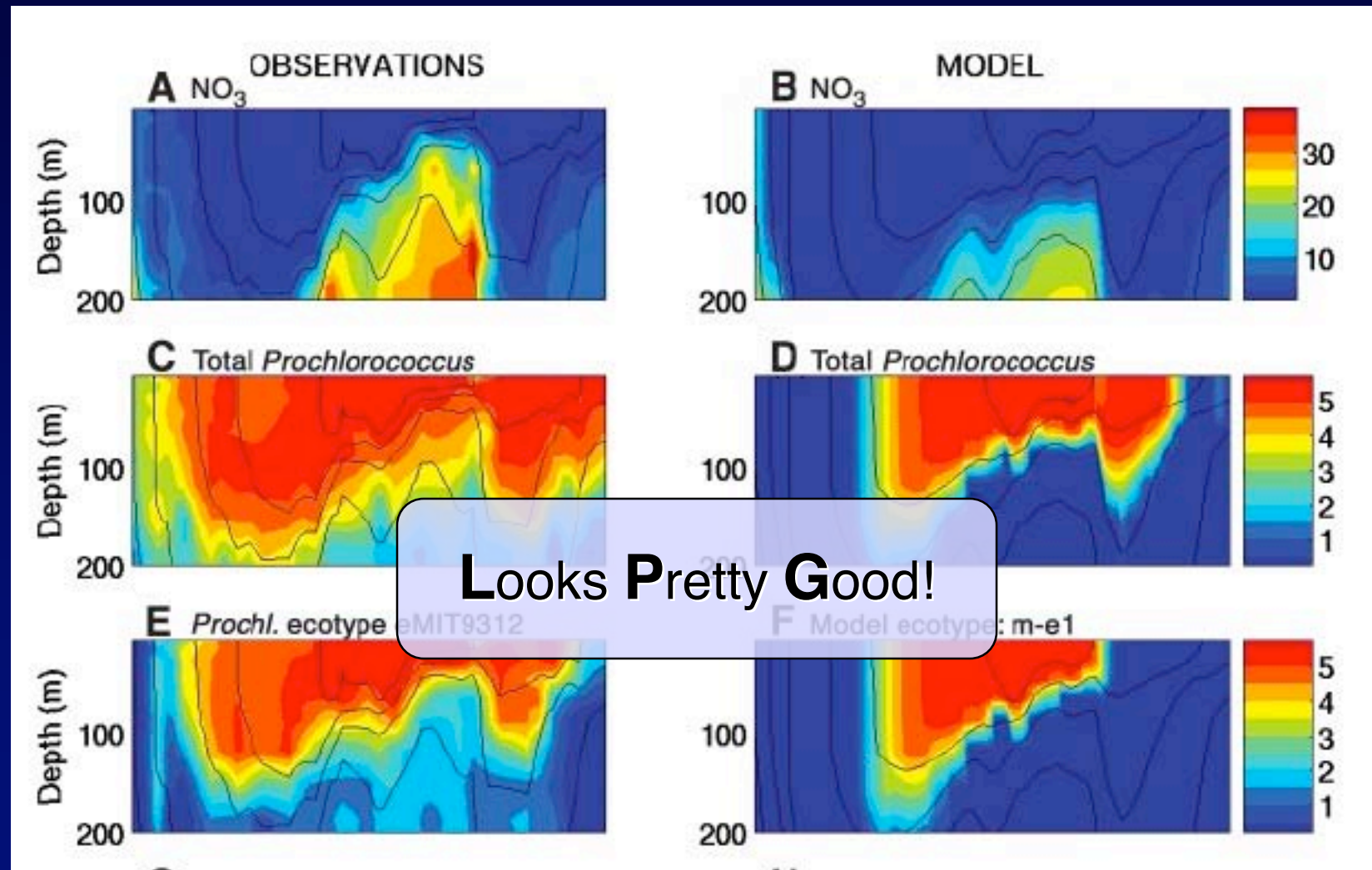
Complexity is added to increase realism and to test hypotheses



Doney, S. C., M. R. Abbott, J. J. Cullen, D. M. Karl, and L. Rothstein. 2004. From genes to ecosystems: the ocean's new frontier. *Frontiers in Ecology and the Environment* 2: 457-466.

John Cullen: Agouron Institute 2008

Conventionally tested by the “LPG” criterion —
but that is changing

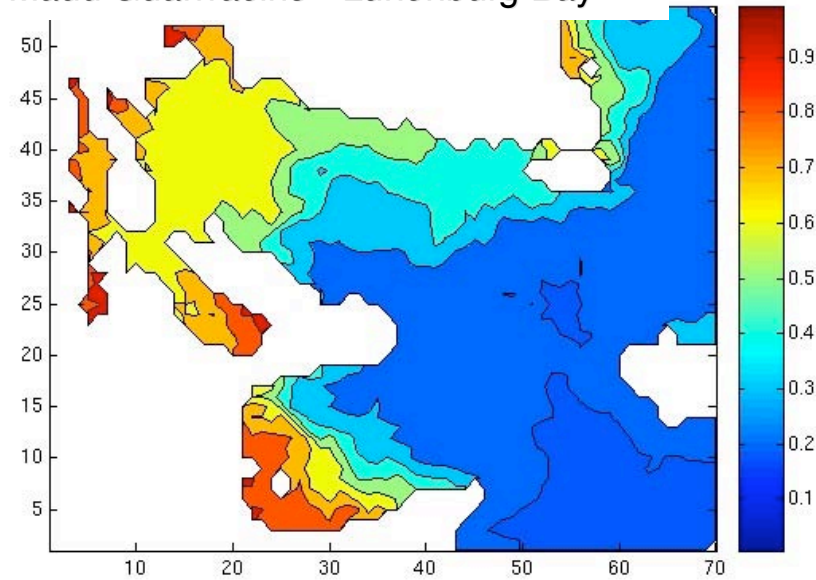


Follows, M. J., S. Dutkiewicz, S. Grant, and S. W. Chisholm. 2007. Emergent biogeography of microbial communities in a model ocean. *Science* 315: 1843-1846.

John Cullen: Agouron Institute 2008

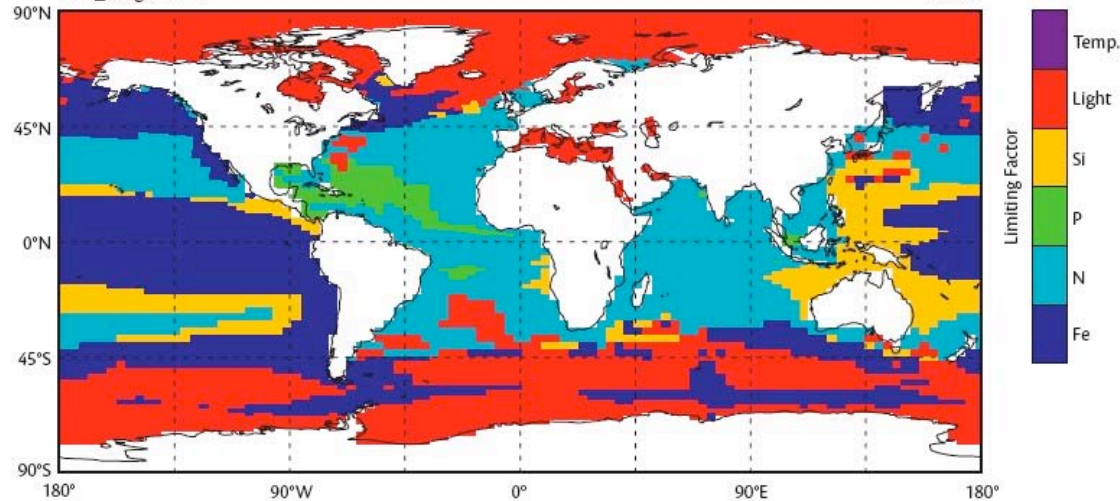
Approaches to Ecosystem Modeling

Maud Guarracino - Lunenburg Bay



ECO_T62gx3v5.21

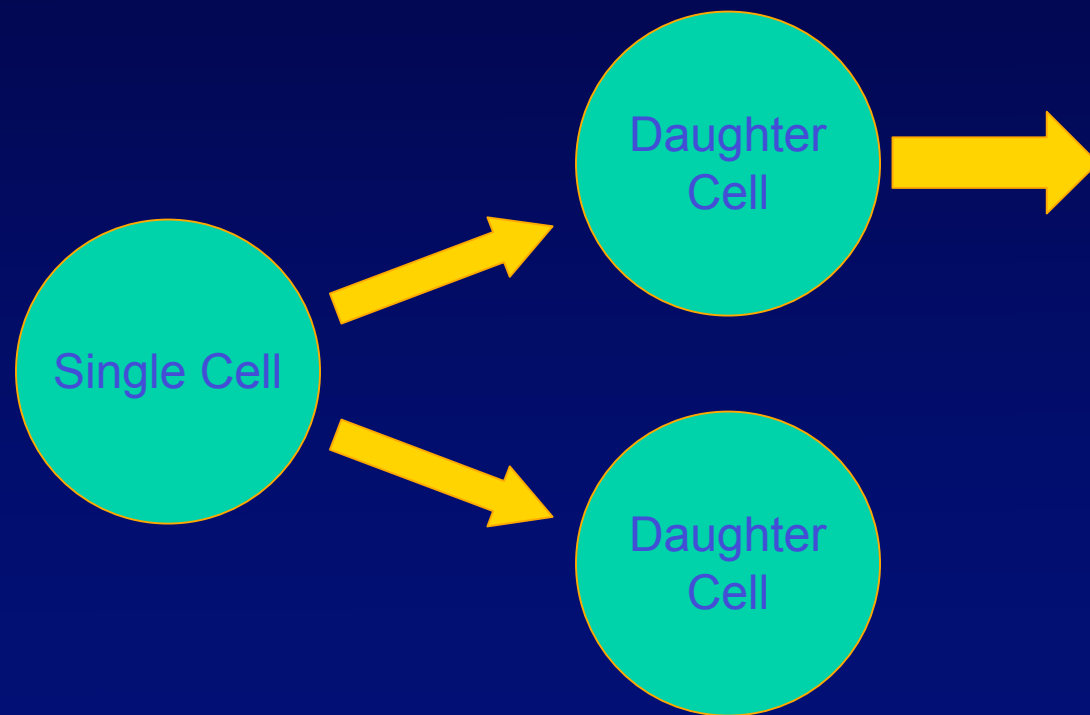
Year 50



Fundamentally, ecosystem models should predict population dynamics

Growth

Loss



Accumulate (Bloom)

Be eaten

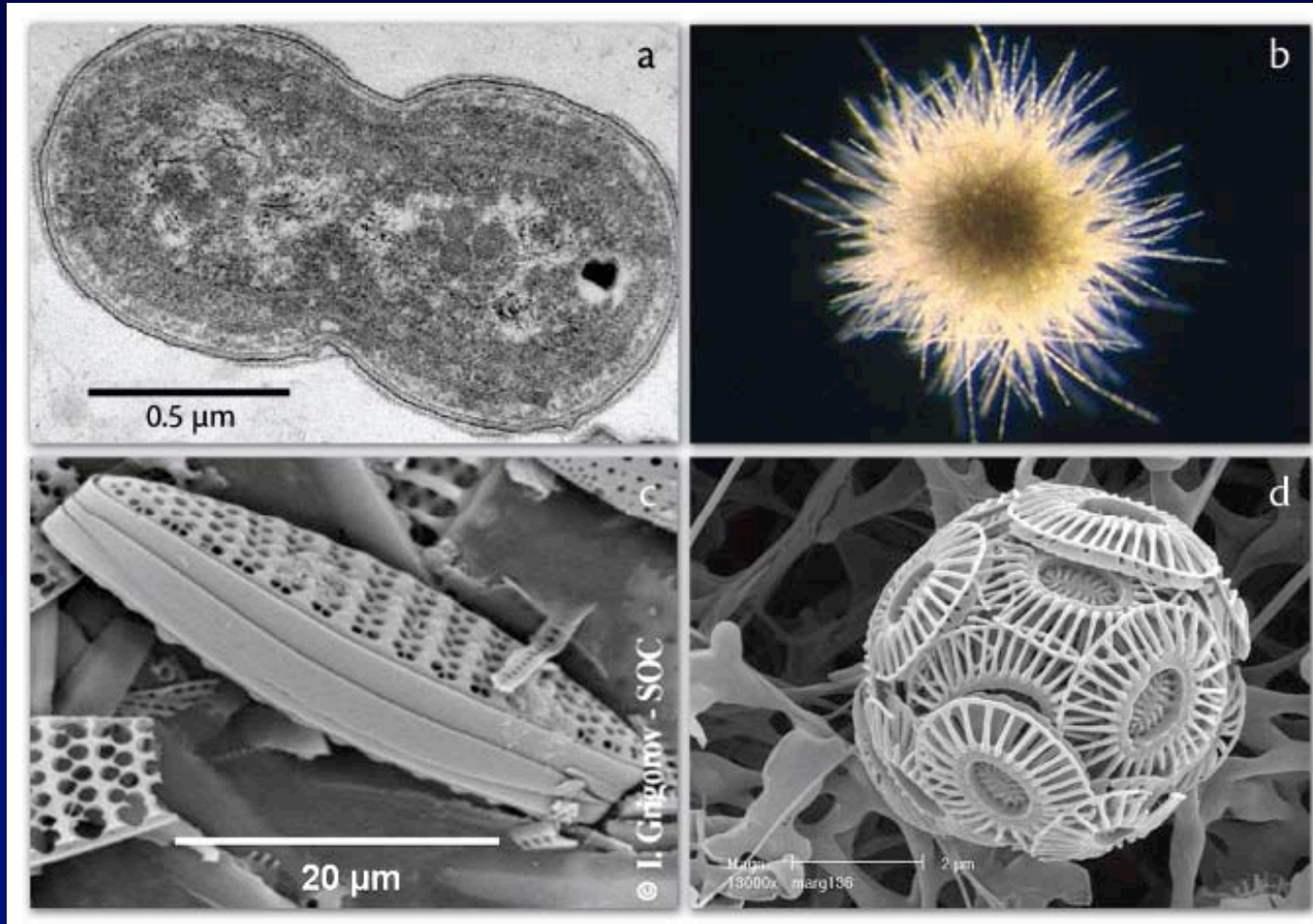
Blow up (viral lysis)

Sink

Die (e.g., apoptosis)

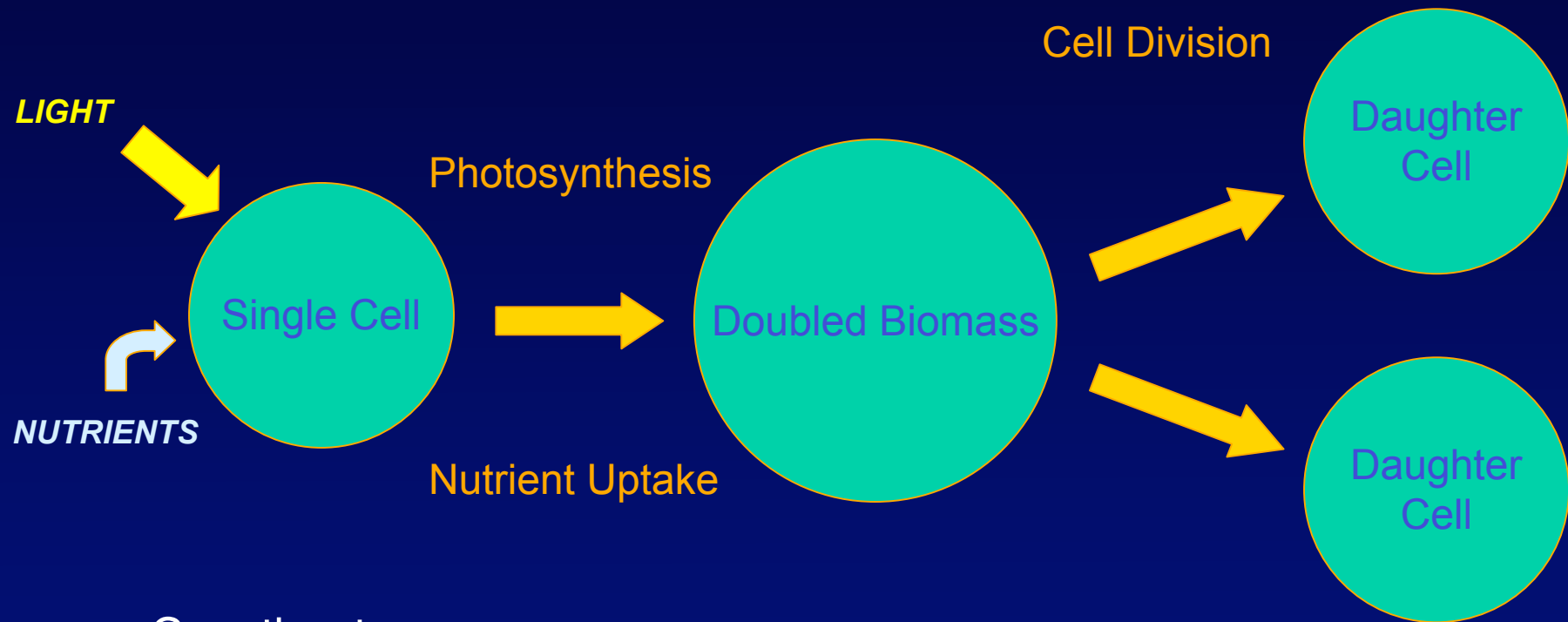
A bit weird, because “population” refers to species, and many species are often lumped

Biogeochemical models must include functional groups



Essential Knowledge:

Environmental Influences on the Growth and Chemical Composition of Phytoplankton



Growth rate

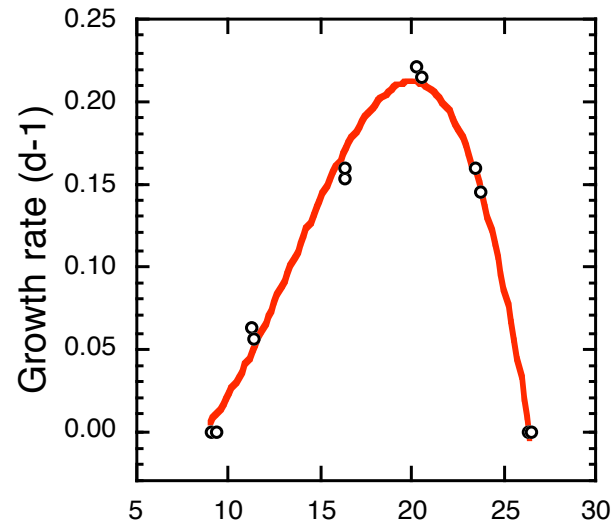
Chemical composition

Biogeochemical transformations

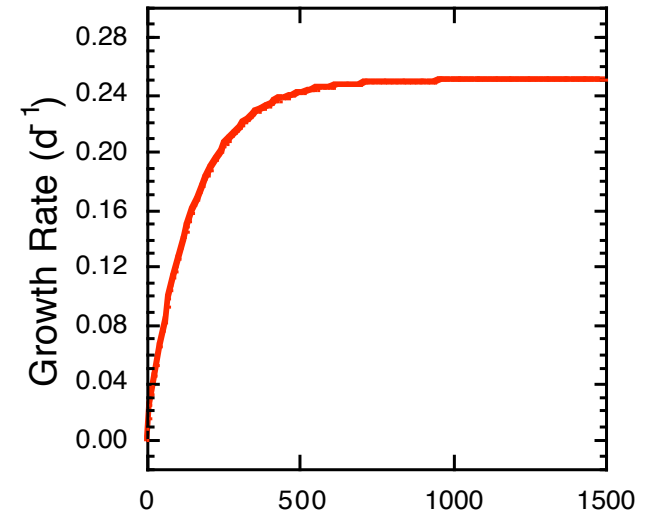
**Critical to
know the
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Influences on
the Growth of
Phytoplankton:**

**Temperature
Light
Daylength
Nutrients**

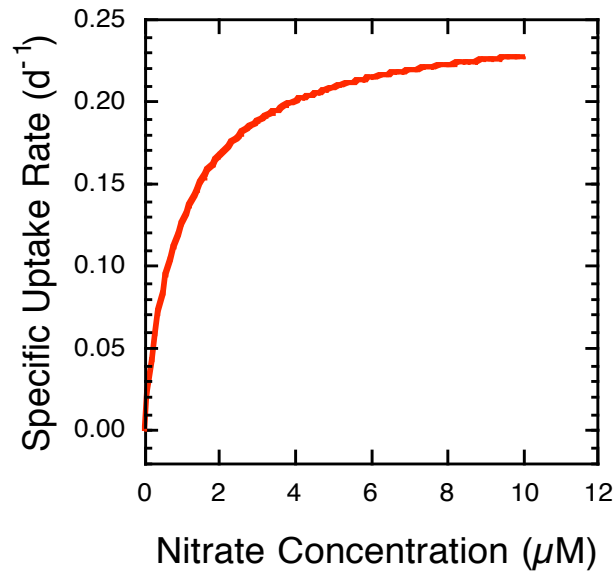
Alexandrium ostenfeldii



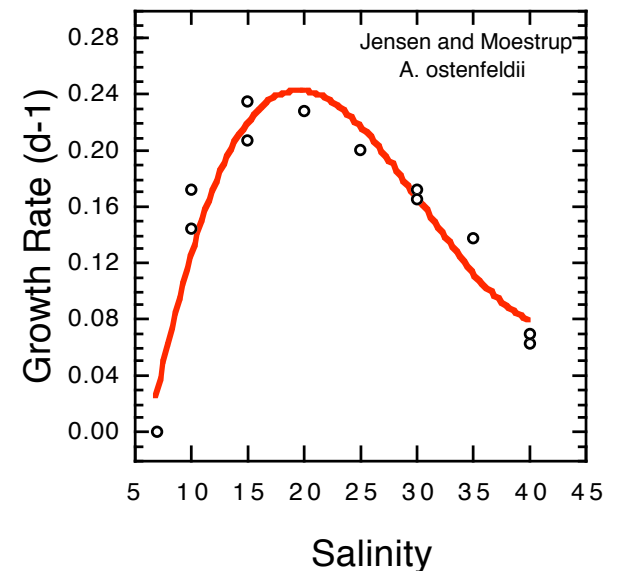
Growth vs Irradiance



Nutrient Uptake Kinetics

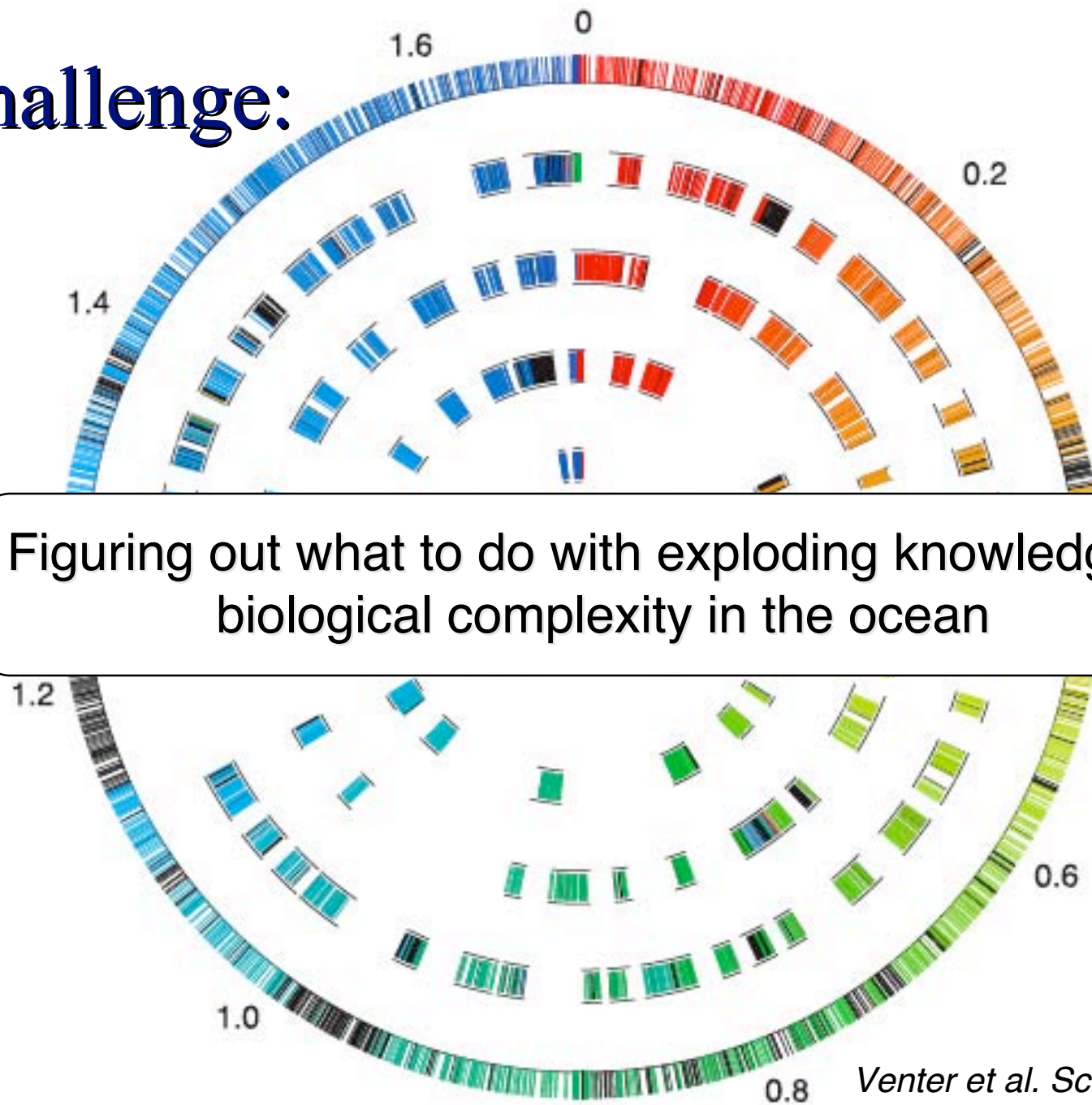


Effect of Salinity

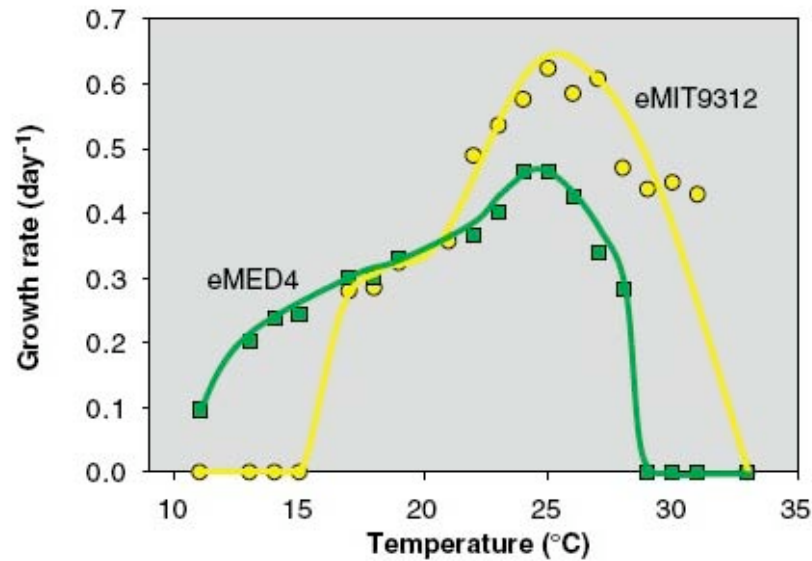


Challenge:

Figuring out what to do with exploding knowledge of biological complexity in the ocean



Venter et al. Science 2004

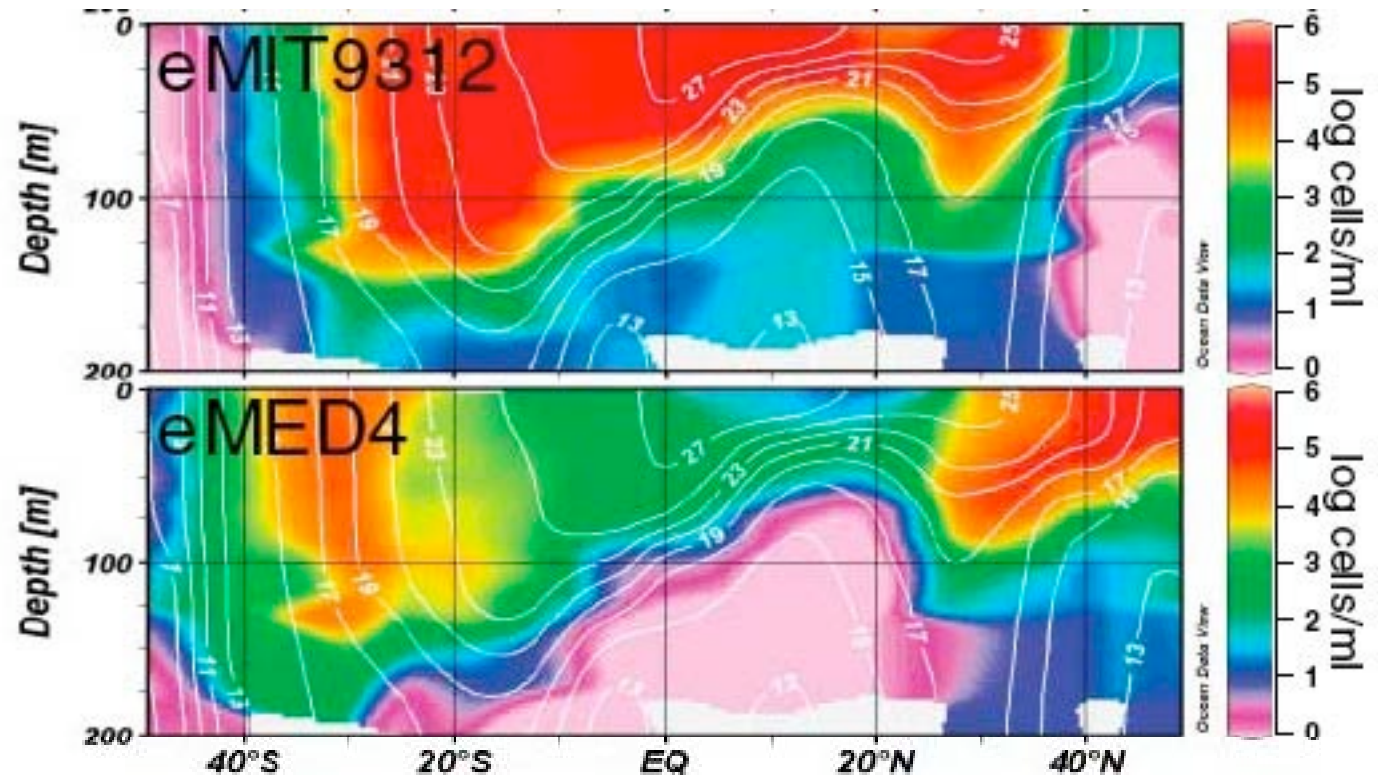


Growth vs Temperature

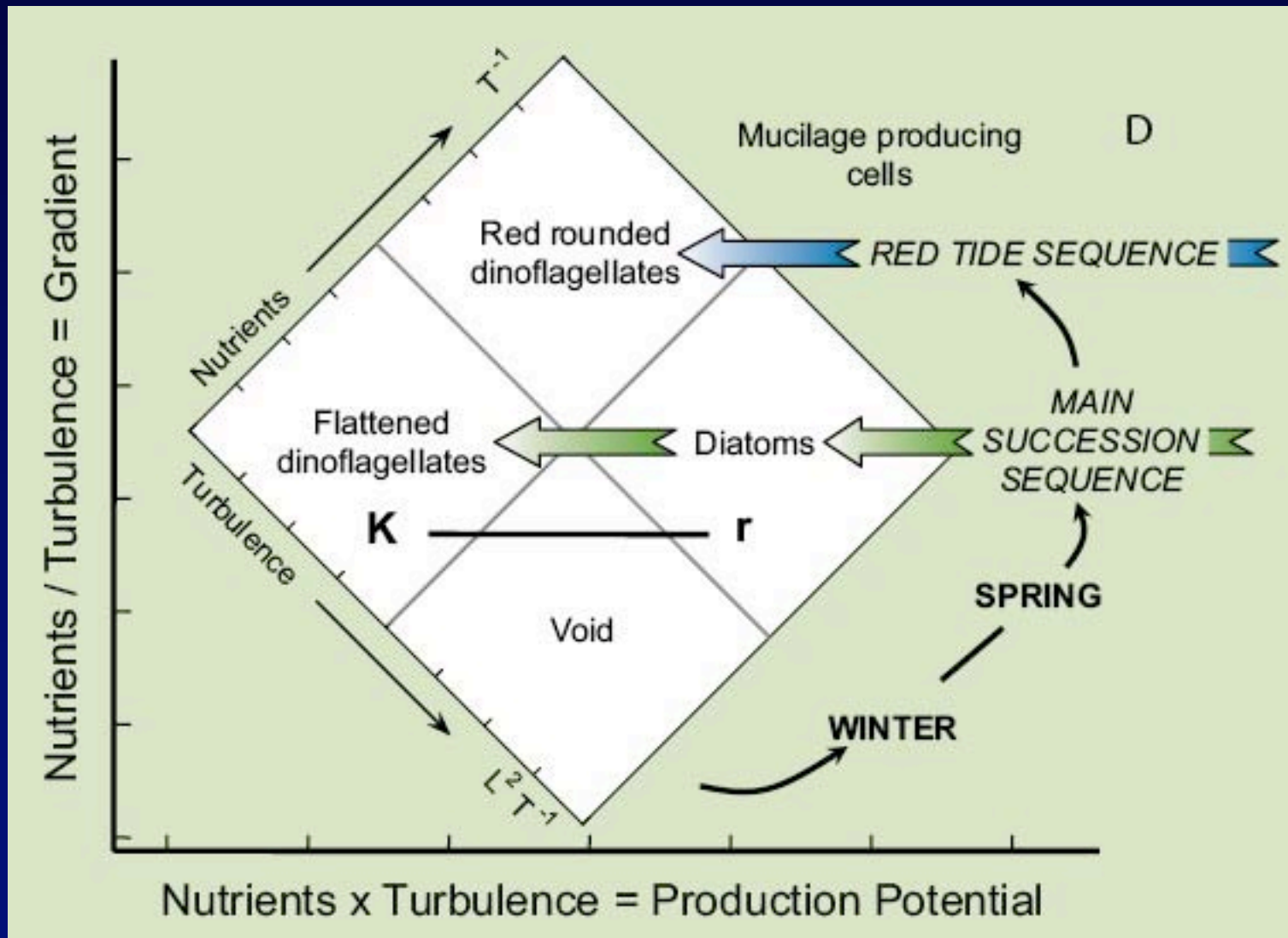
SCIENCE VOL 311 24 MARCH 2006 1737

Niche Partitioning Among *Prochlorococcus* Ecotypes Along Ocean-Scale Environmental Gradients

Zackary I. Johnson,^{1,2*} Erik R. Zinser,^{1,3*} Allison Coe,¹ Nathan P. McNulty,¹
E. Malcolm S. Woodward,⁴ Sallie W. Chisholm^{1†}



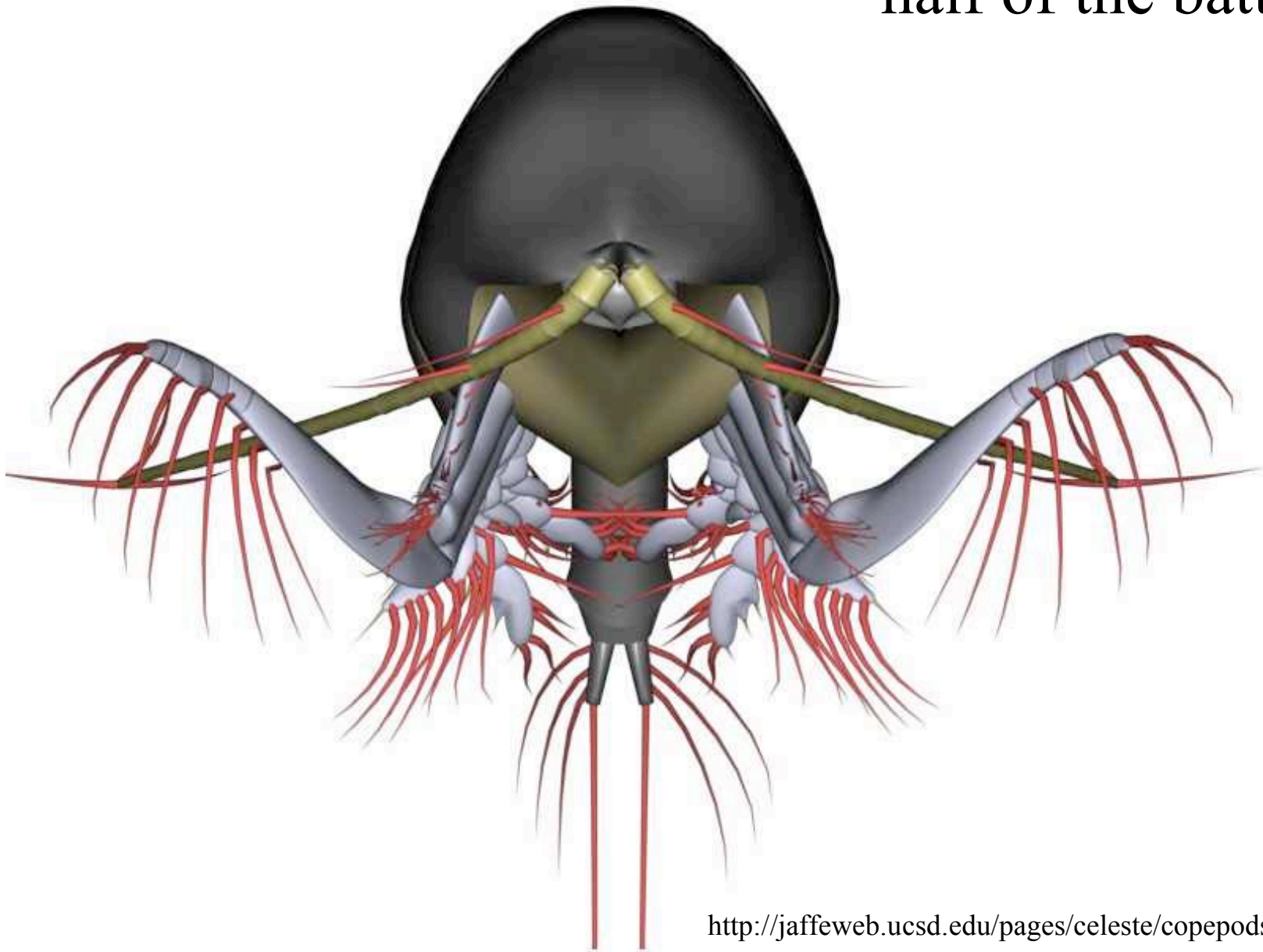
Many other things define the growth-niche of marine phytoplankton



From Margalef et al. 1979 in Cullen et al. Oceanography Mag. 2007

John Cullen: Agouron Institute 2008

And cell division is only
half of the battle!



<http://jaffeweb.ucsd.edu/pages/celeste/copepods.html>

Reduction of loss can be as good as an increase of growth rate

Architecture and material properties of diatom shells provide effective mechanical protection

Christian E. Hamm*, Rudolf Merkel†‡, Olaf Springer§, Piotr Jurkojc§, Christian Maier†, Kathrin Pechtel† & Victor Smetacek*

NATURE | VOL 421 | 20 FEBRUARY 2003 | www.nature.com/nature

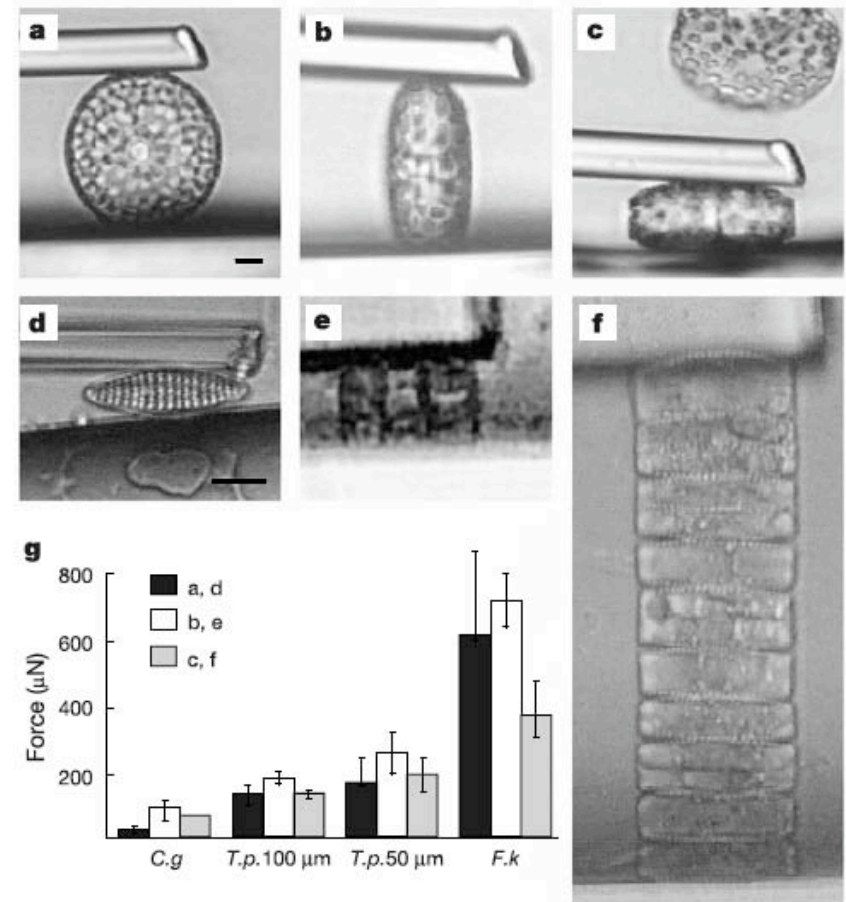
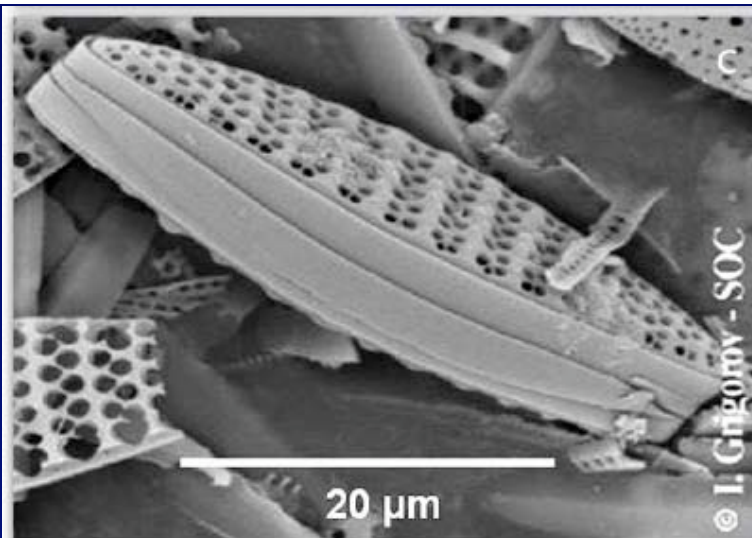
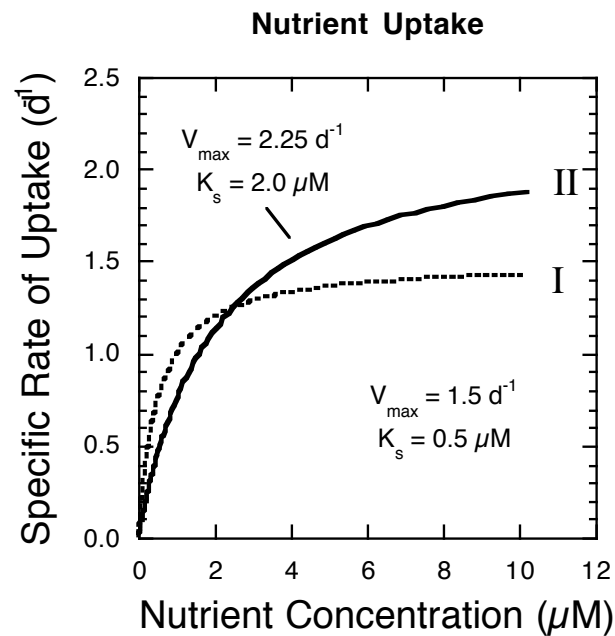
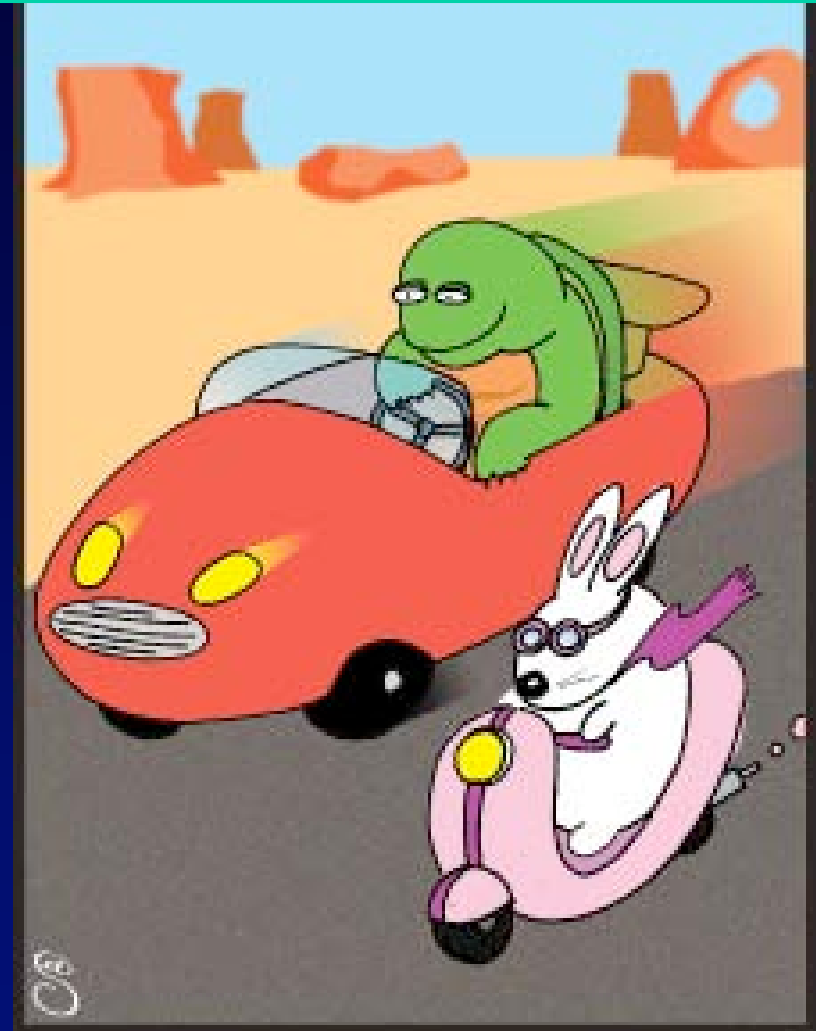


Figure 1 Glass needle tests: Live single cells of *T. punctigera* (a–c) and *F. kerguelensis* (d–f), in chains (e, f). Pressures applied along the girdle bands, (a, d), across the girdle bands (b, e), and across the centre of the valves (c, f). **g**, Forces necessary to break *Coscinodiscus granii* (C.g.), *Thalassiosira punctigera* (T.p.) with diameters of 100 and 50 μm , and *Fragilaropsis kerguelensis* (F.k.). *C. granii* has a geometry similar to that of *T. punctigera*. Scale bars, 10 μm .

So rapid growth is not
the only strategy for
survival / selection

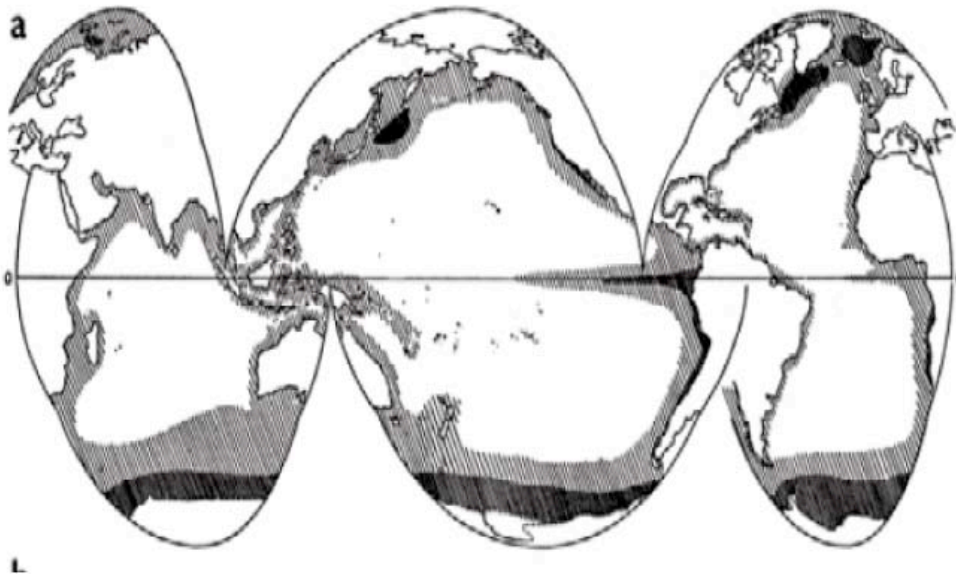


It's the gene, stupid!

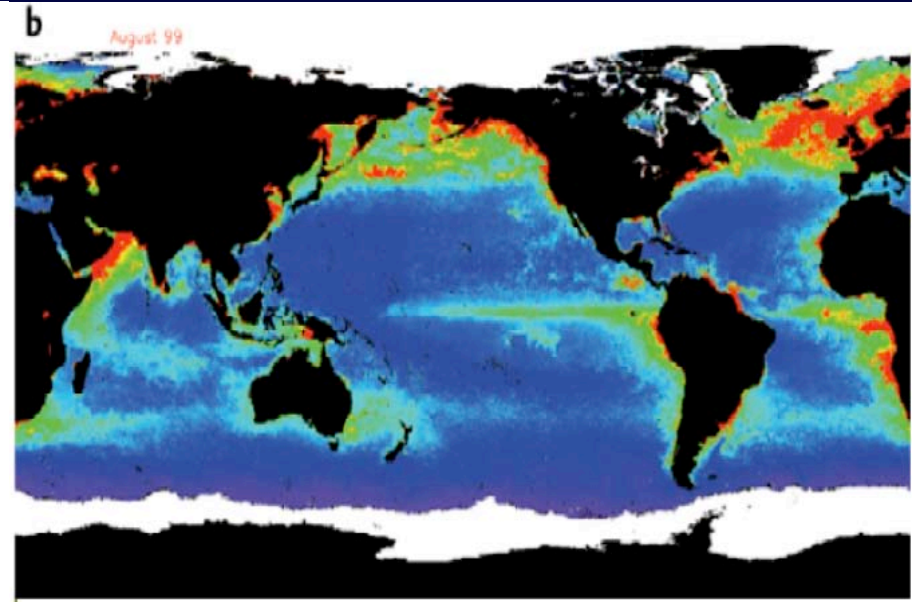


www.andyslocum.com/images/tortoise&hare.jpg

Top-down or bottom-up control?



Sverdrup's (1955) map of productivity based on vertical convection, upwelling and turbulent diffusion

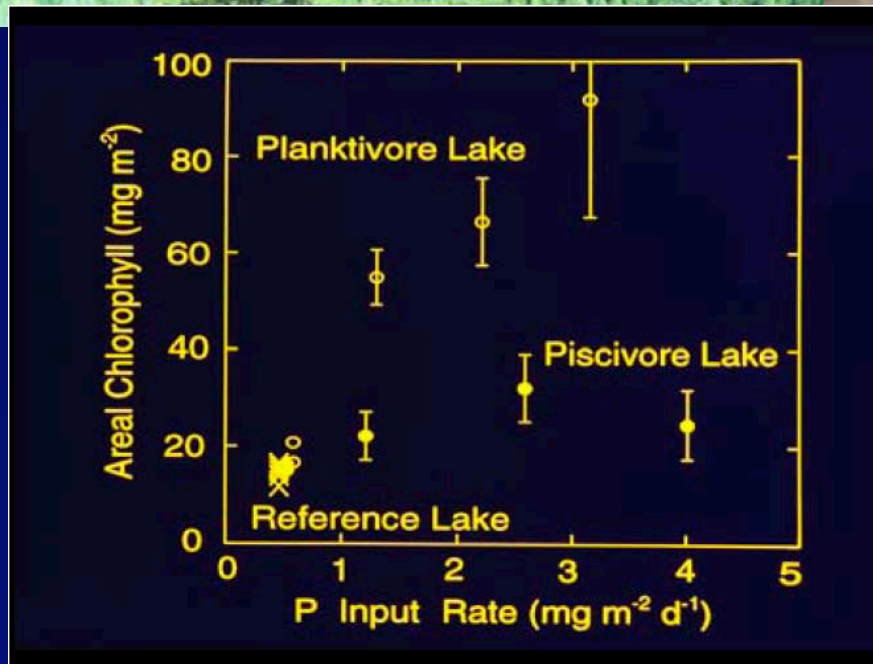


Global productivity estimated from remote sensing (Falkowski et al. 1998)

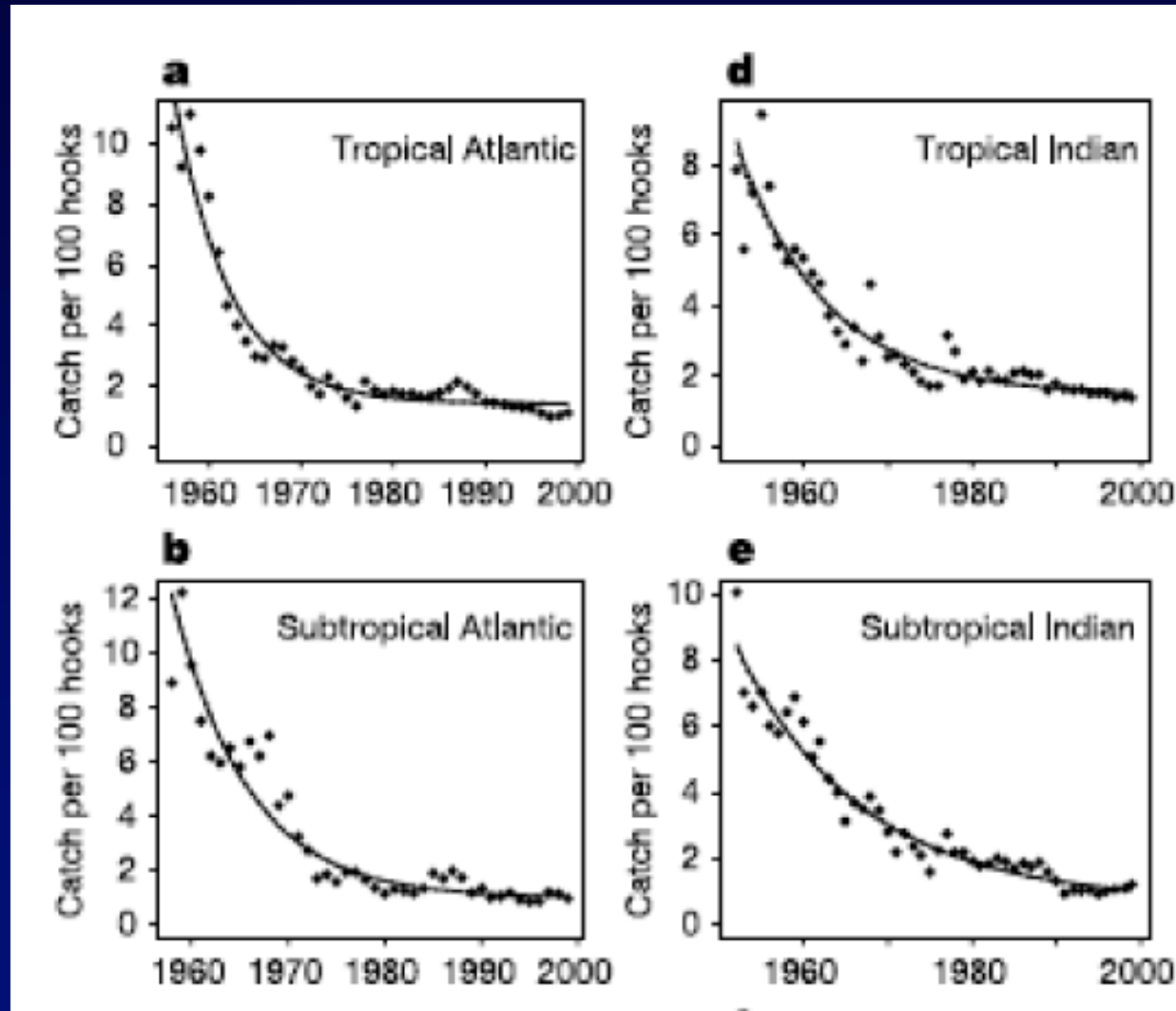
As presented by John McGowan (Oceanography Mag., 2004)

John Cullen: Agouron Institute 2008

Top-down control



Global test of the top-down hypothesis?



Decline of fish stocks since 1960

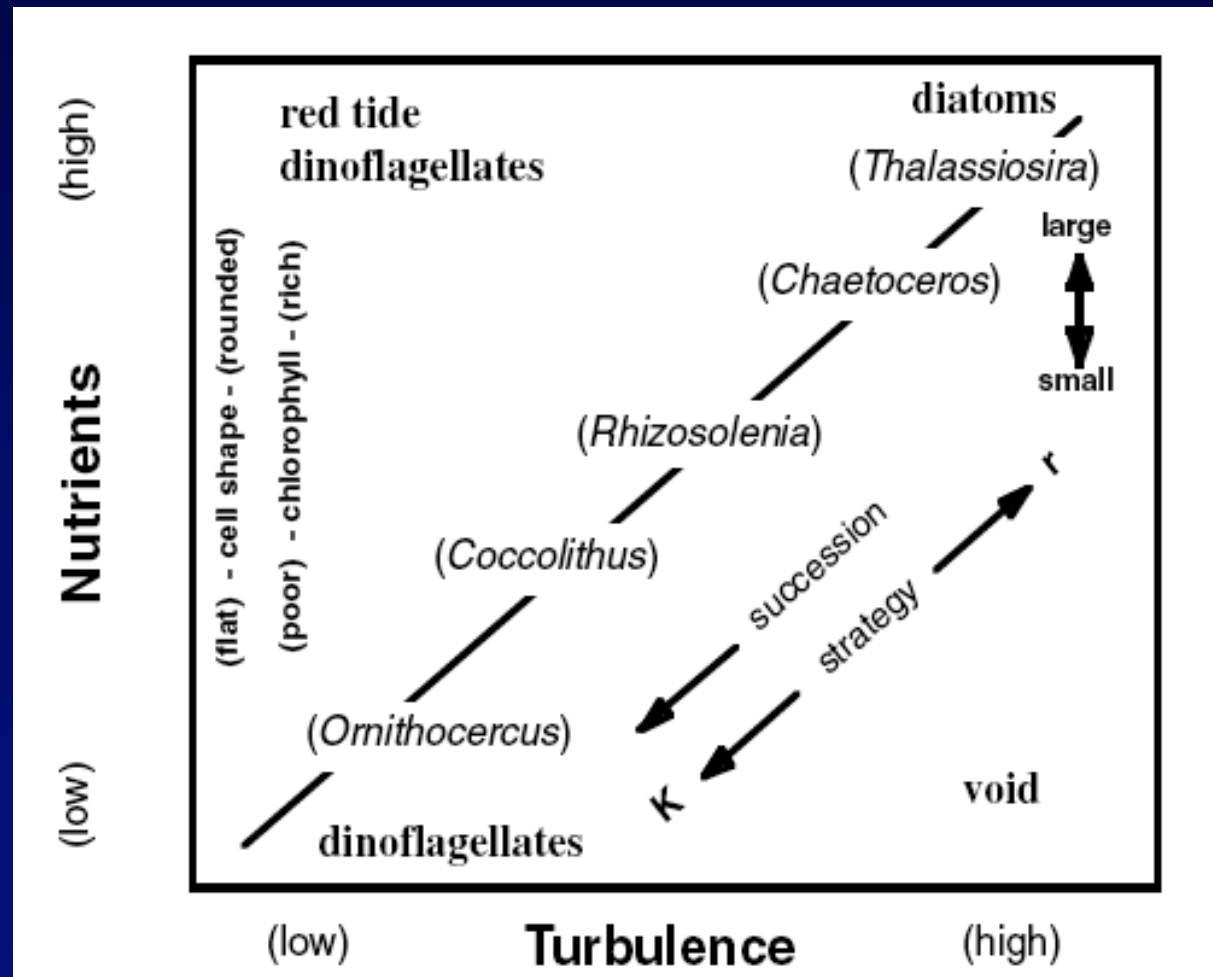
Myers and Worm Nature 2003

Bottom-up processes

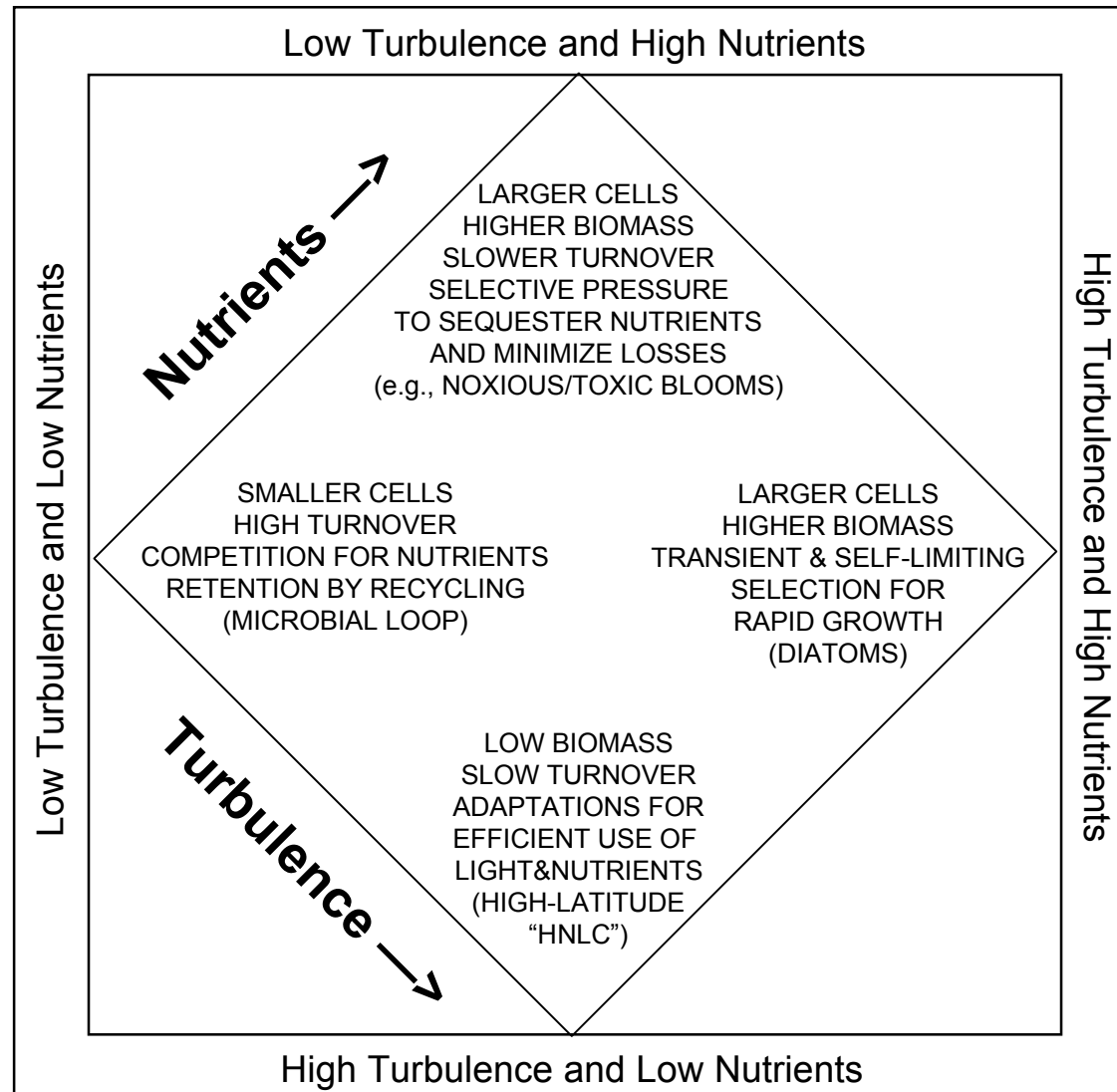
It can be argued that a similarly parsimonious set of factors determines the distribution of pelagic biomes, each with its characteristic flora and fauna... ***Copepods and whales do not determine which groups of plants will flourish; like the phytoplankton, they are themselves expressions of the regional physical oceanographic regime.***

(Alan Longhurst's section of Cullen et al., 2002, *The Sea*)

A Tool for Making Sense of Physically Forced Ecosystem Dynamics: Margalef's Mandala



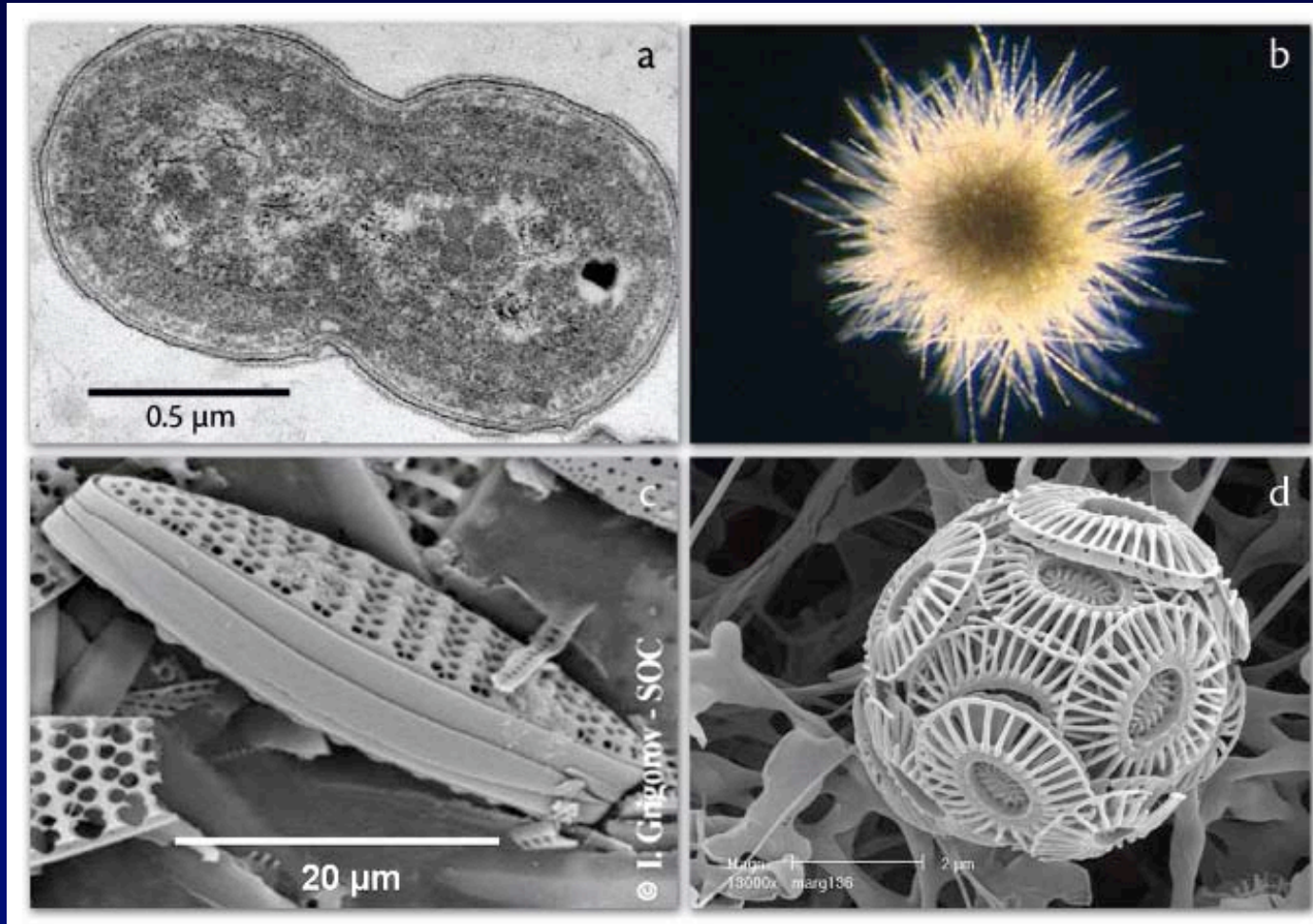
Event-Scale Forcing —>
<— Succession



Potential for Production and Export —>

Cullen et al. 2002, The Sea

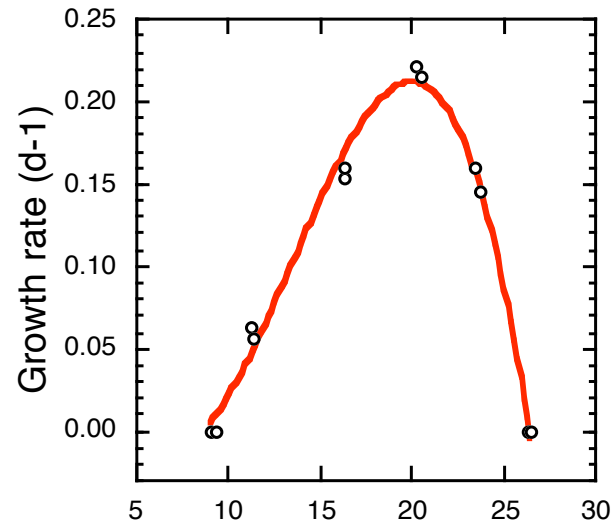
The challenge: quantitative description of the niches of functional groups



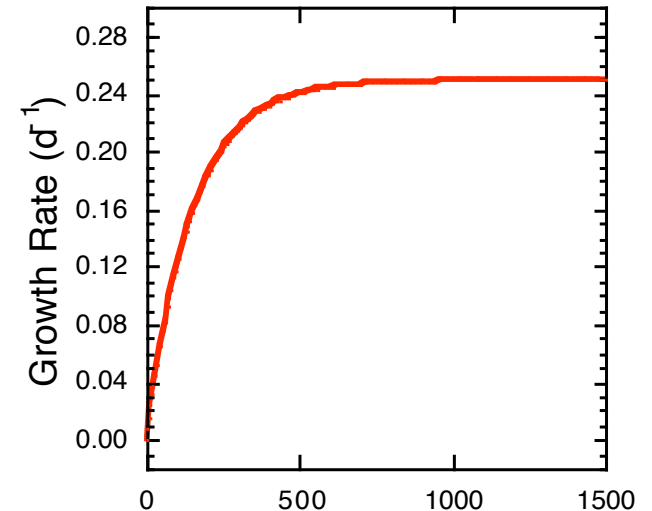
**Critical to
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**Temperature
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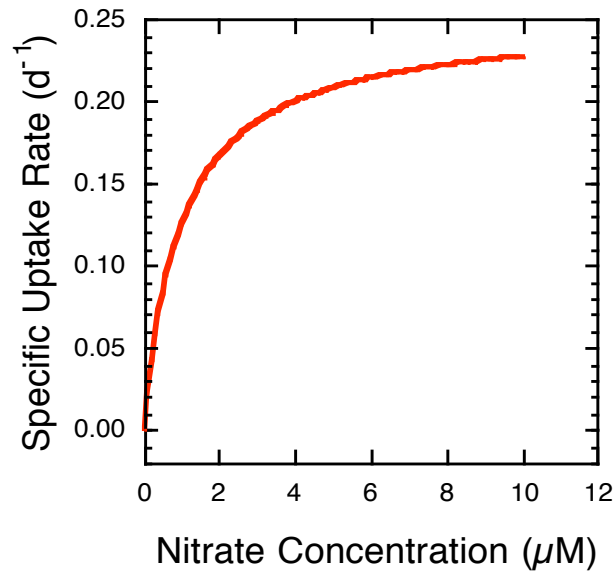
Alexandrium ostenfeldii



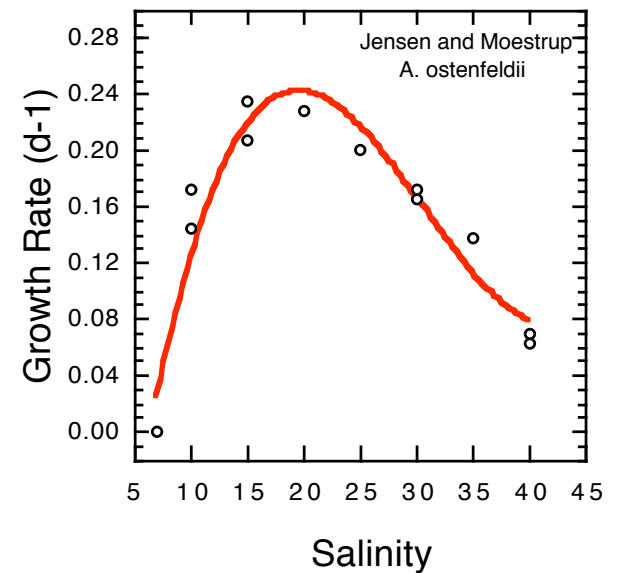
Growth vs Irradiance



Nutrient Uptake Kinetics

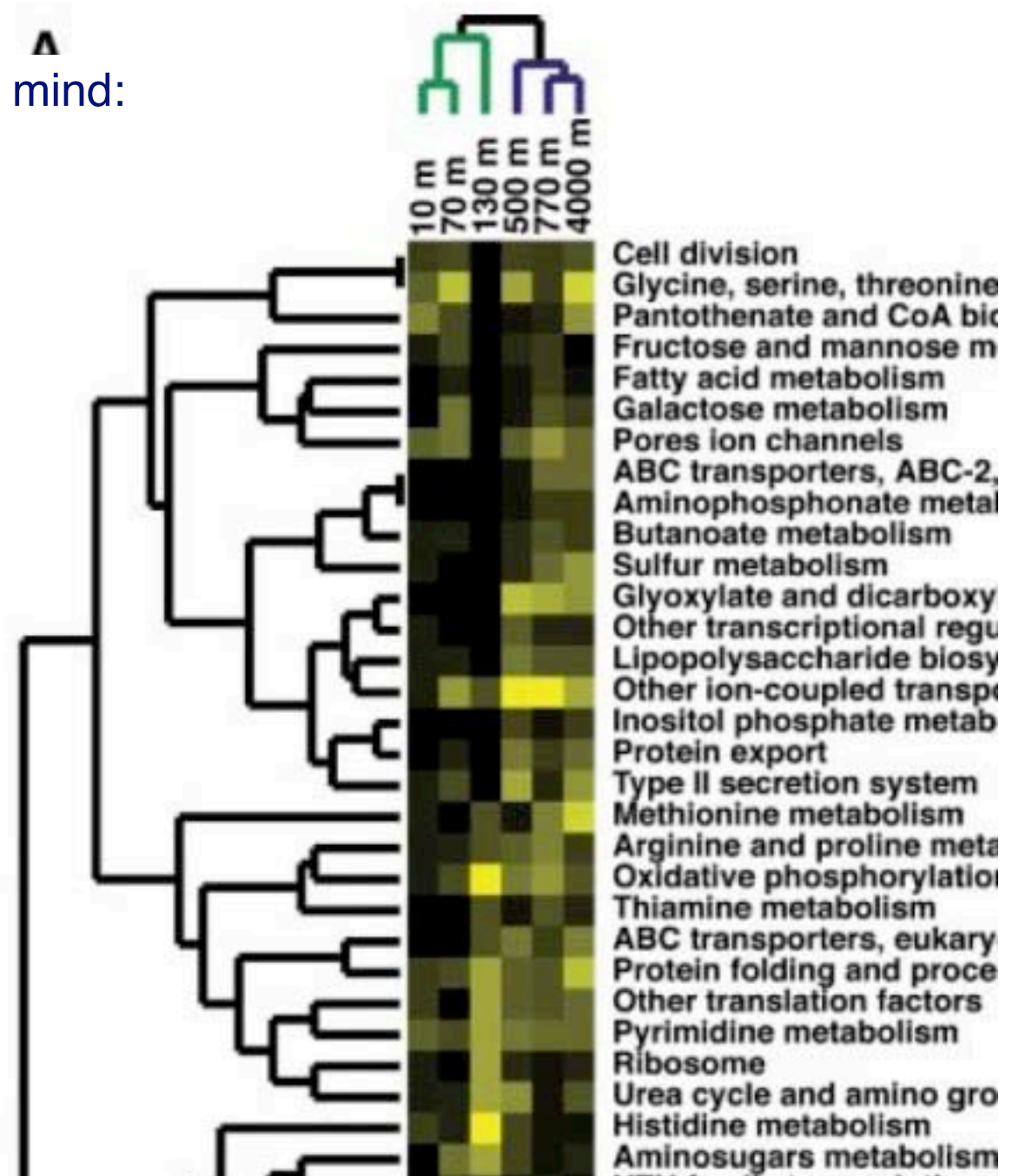


Effect of Salinity



Keep in the back of your mind:

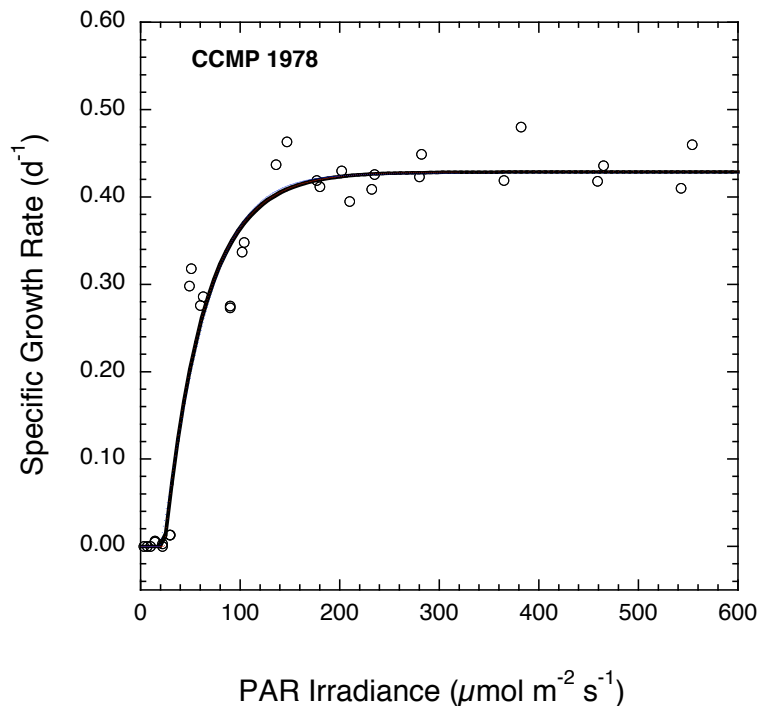
***Surely, all
this means
something!***



Ecosystem modeling ground zero:

Acclimated growth rate: genotypic

Cullen et al. in prep



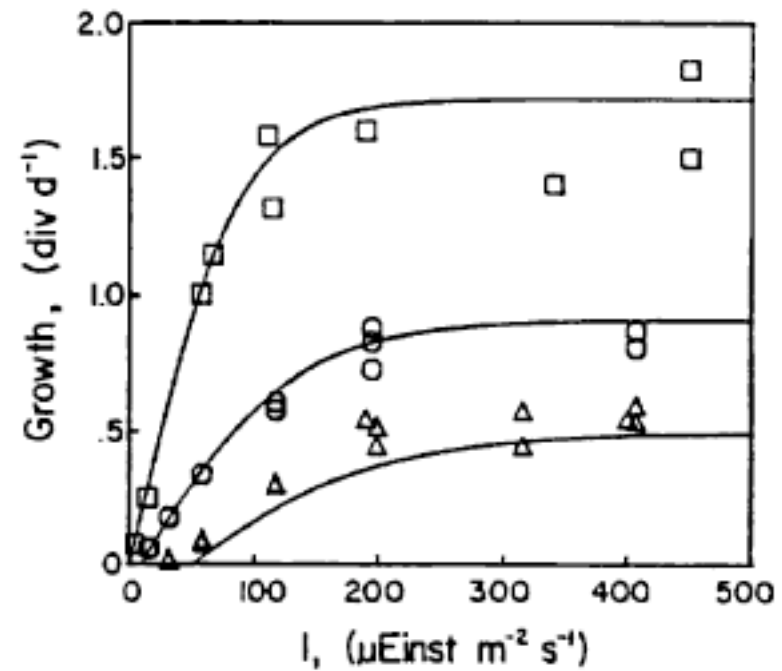
$$\mu(E) = (\mu_{\max}) \left(1 - e^{\frac{-(E-K_C)}{(K_E-K_C)}}\right) \quad \text{if } E > K_C$$
$$\mu(E) = 0 \quad \text{if } E \leq K_C$$

Species evolve different functions
through **adaptation** and selection

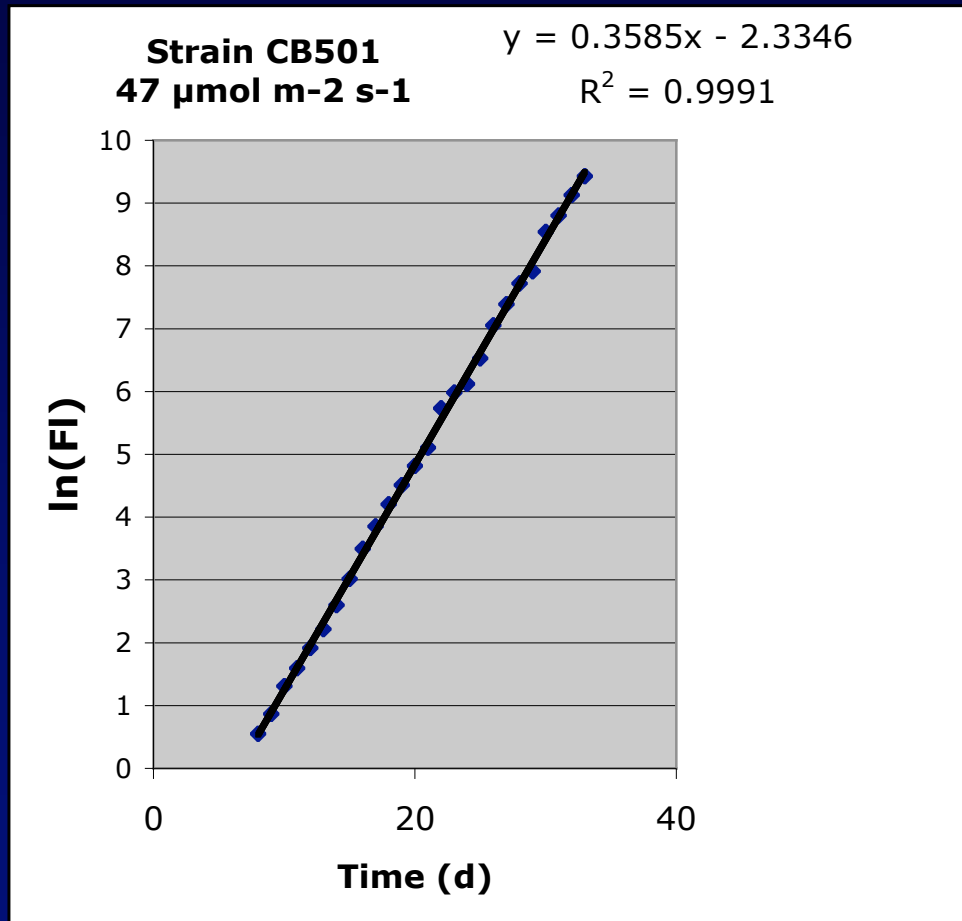
Journal of Plankton Research Vol.9 no.3 pp.459-482, 1987

On the causes of interspecific differences in the growth-irradiance relationship for phytoplankton. Part I. A comparative study of the growth-irradiance relationship of three marine phytoplankton species: *Skeletonema costatum*, *Olisthodiscus luteus* and *Gonyaulax tamarensis*

Christopher Langdon¹



μ as a function of T and E may not be relevant for describing growth rate in dynamic environments



Alexandrium fundyense

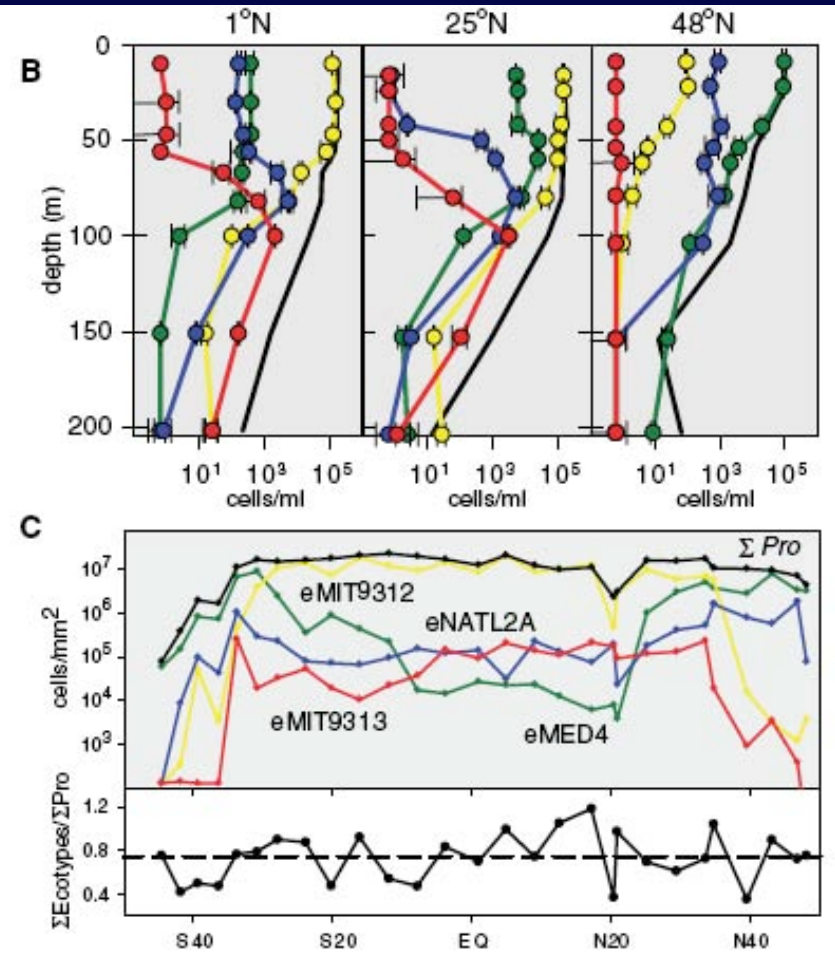
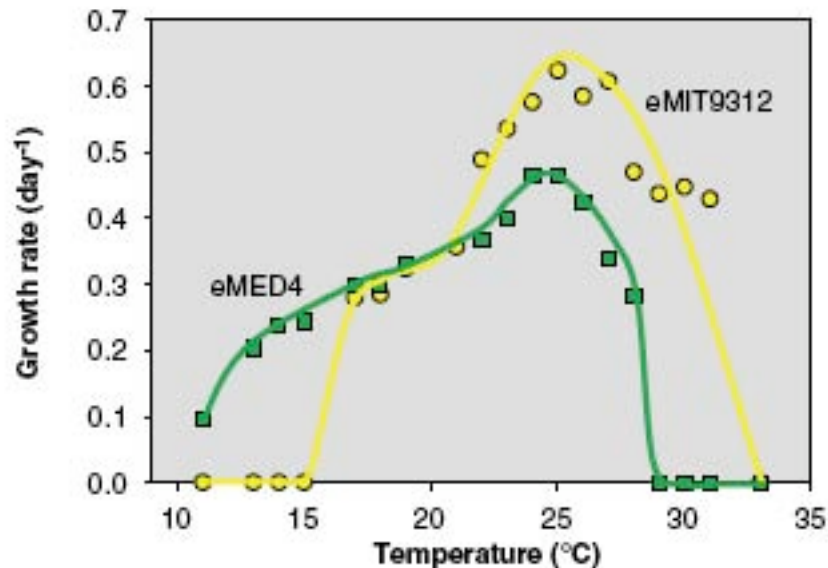
Growth determined using the method of Brand and Guillard

Brand, L. E. and Guillard, R. R. L. (1981). A method for the rapid and precise determination of acclimated phytoplankton reproduction rates. *J. Plankton Res.* 3: 191-201.

But it is an excellent start for identifying niches

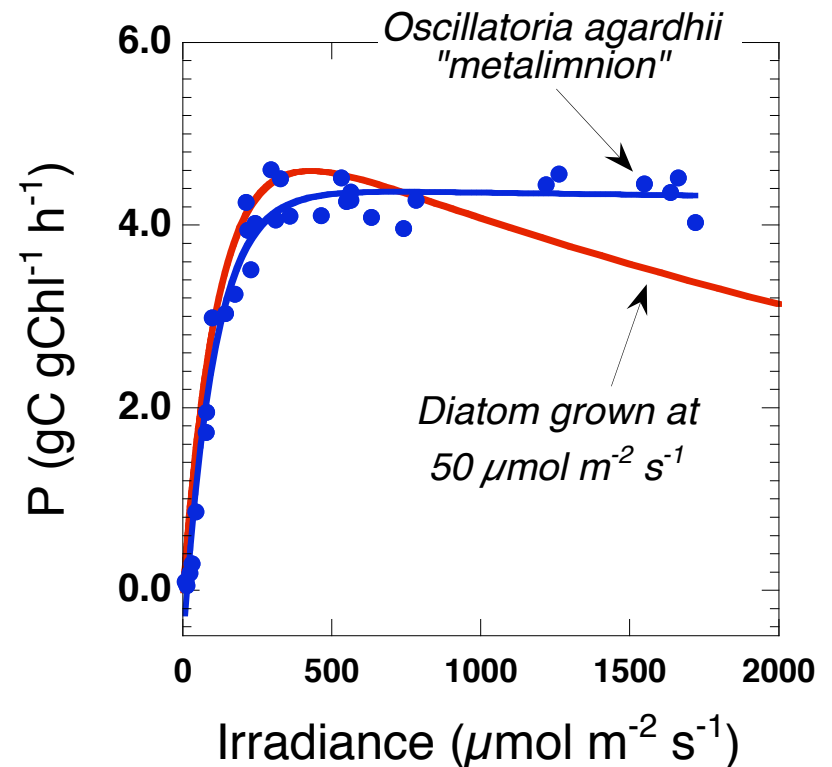
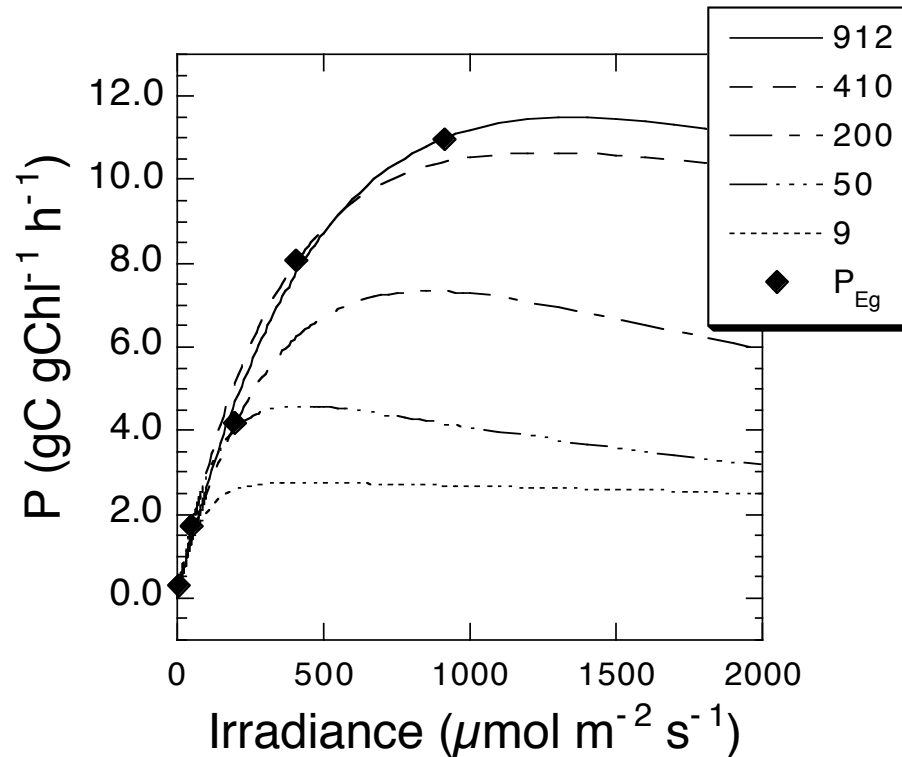
Niche Partitioning Among *Prochlorococcus* Ecotypes Along Ocean-Scale Environmental Gradients

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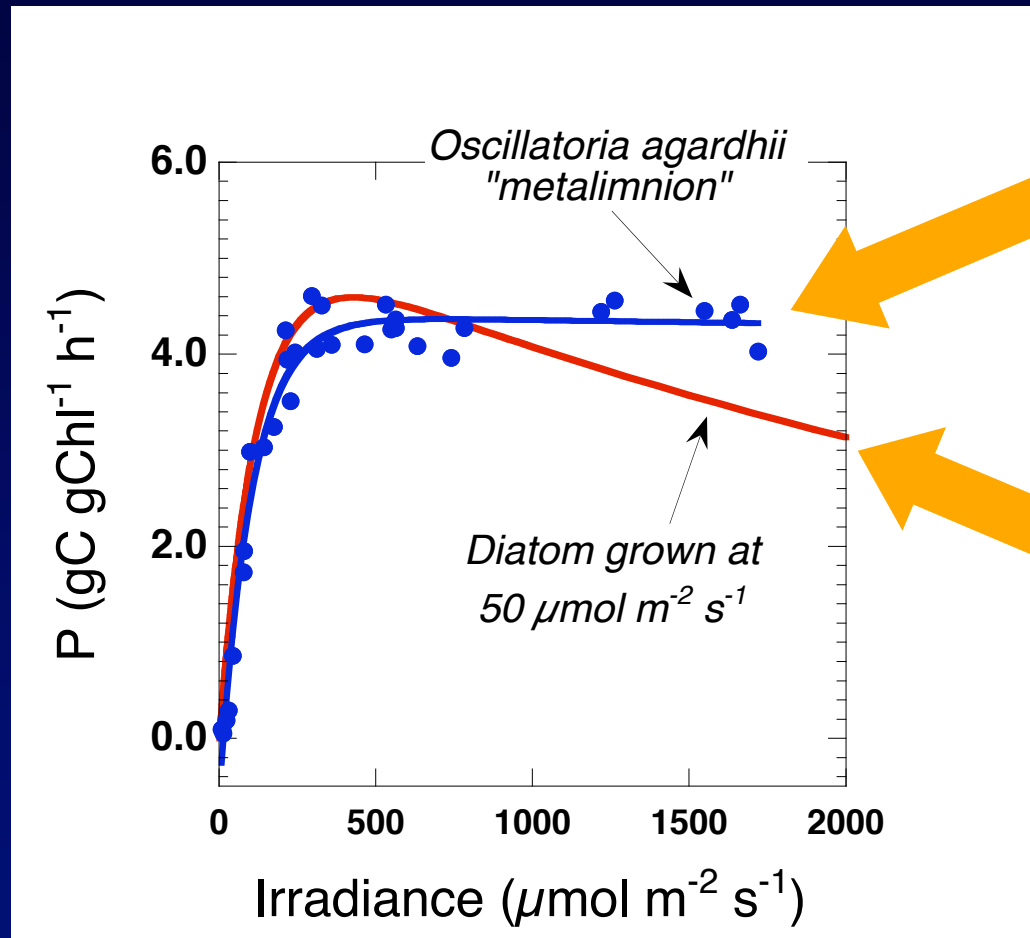
SCIENCE VOL 311 24 MARCH 2006

Photosynthesis vs Irradiance: phenotypic



Species develop different functions through **acclimation**

Photosynthesis vs Irradiance: phenotypic



Layer former

Mixer

Strategies to respond to environmental variability are adaptations

Adaptations to oceanic vs coastal conditions

Vol. 41: 275–282, 1987

MARINE ECOLOGY – PROGRESS SERIES
Mar. Ecol. Prog. Ser.

Published December 14

Thalassiosira oceanica and *T. pseudonana*: two different photoadaptational responses

E. Sakshaug¹, S. Demers², C. M. Yentsch³

Photosynthetic architecture differs in coastal and oceanic diatoms

Robert F. Strzepek¹⁺ & Paul J. Harrison²⁺

NATURE | VOL 431 | 7 OCTOBER 2004 | www.nature.com/nature

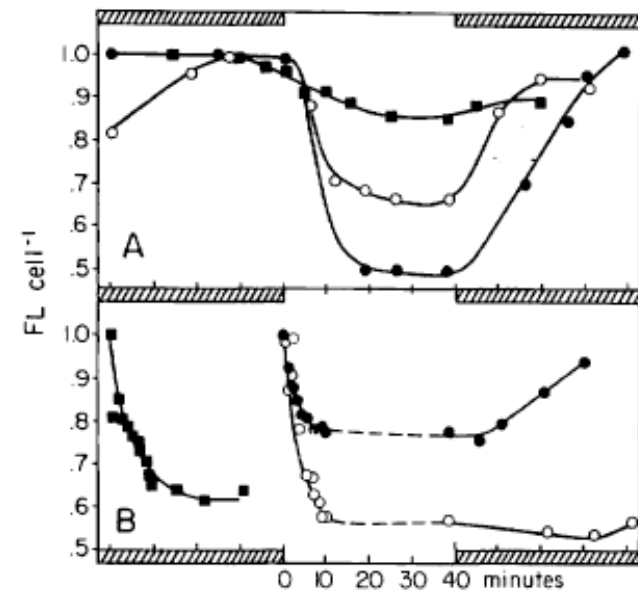


Fig. 5. *Thalassiosira pseudonana* and *T. oceanica*. Short-term changes of *in vivo* fluorescence per cell normalized to the highest observed value in each experiment. (A) *T. pseudonana* at 600 (○) or 800 μmol m⁻² s⁻¹ (●) exposed to 2800, and then to 600 or 800 μmol m⁻² s⁻¹ and *T. oceanica* (■) at 800 μmol m⁻² s⁻¹ exposed to 2800 and then to 800 μmol m⁻² s⁻¹. (B) Changes in *in vivo* fluorescence per cell of *T. pseudonana* after addition of gramicidin; (■) cultures at 800 μmol m⁻² s⁻¹; (○) cultures at 2800 μmol m⁻² s⁻¹ followed by darkness; (●) control (no gramicidin), cultures at 800 μmol m⁻² s⁻¹ followed by darkness. Horizontal bars: period of low light or darkness according to descriptions above

Oceanic species has much reduced capability for regulation

Lean but valiant oceanic survivor

Rough and ready coastal mixer

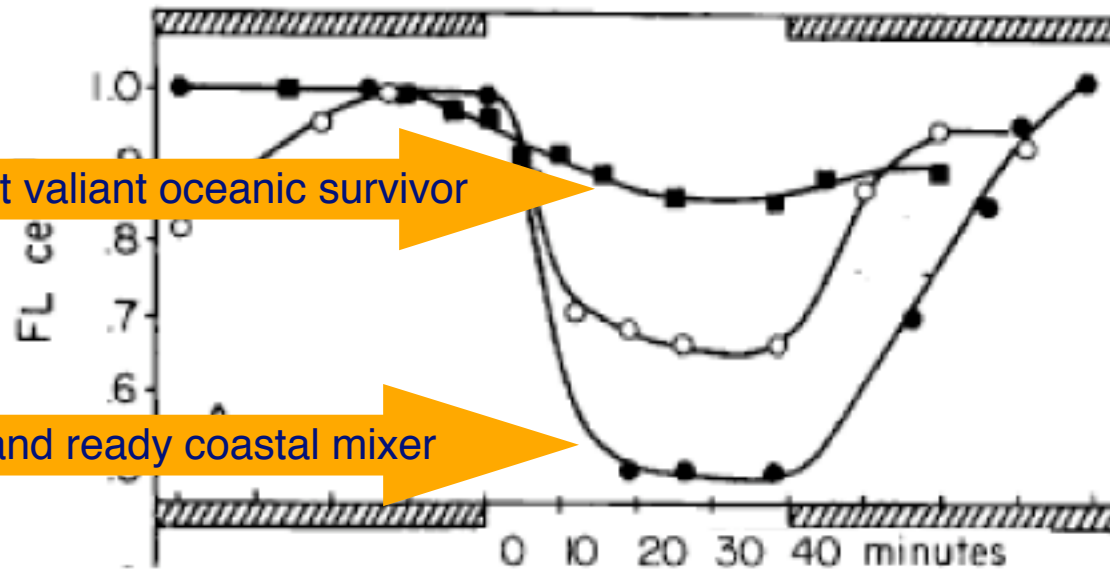
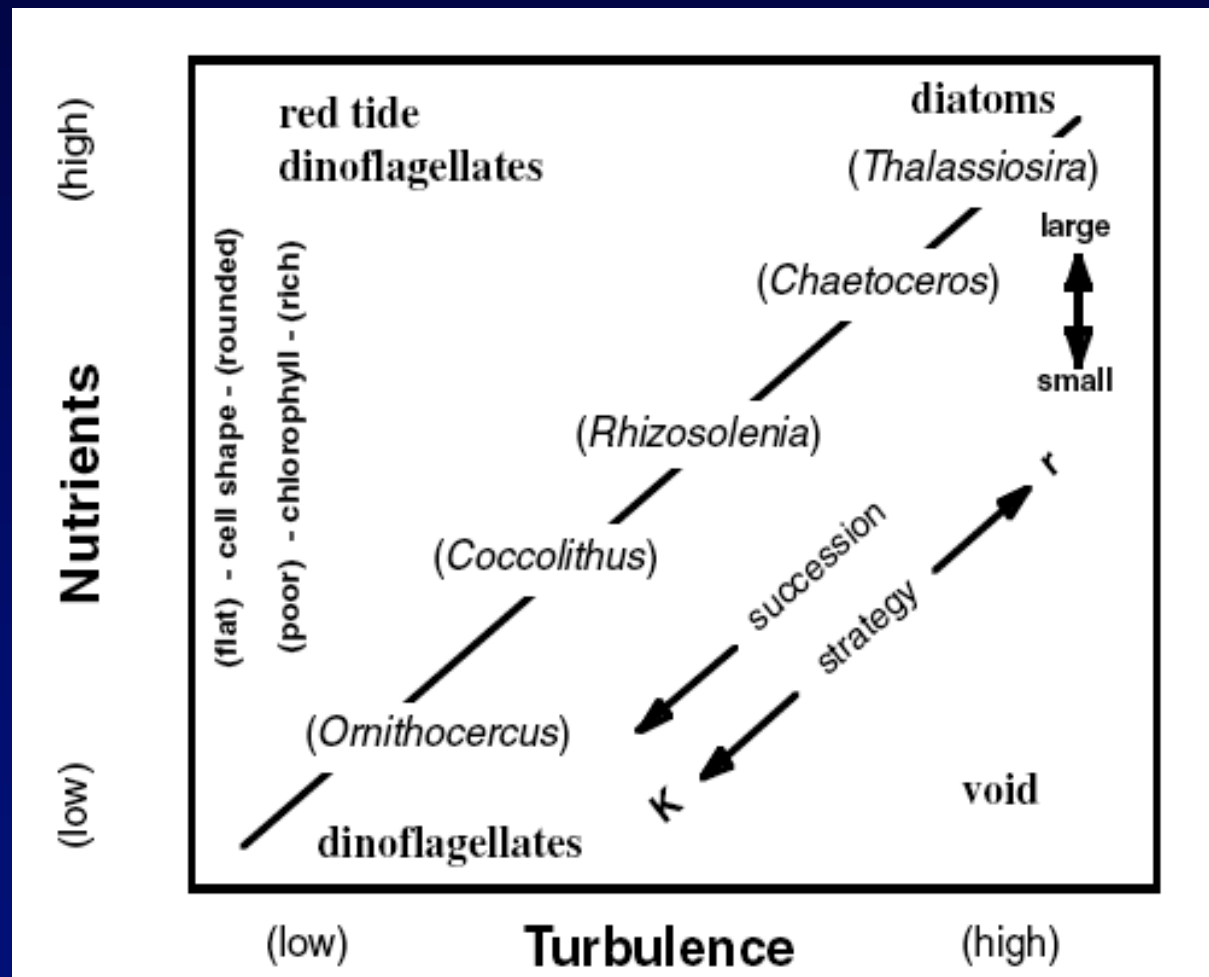
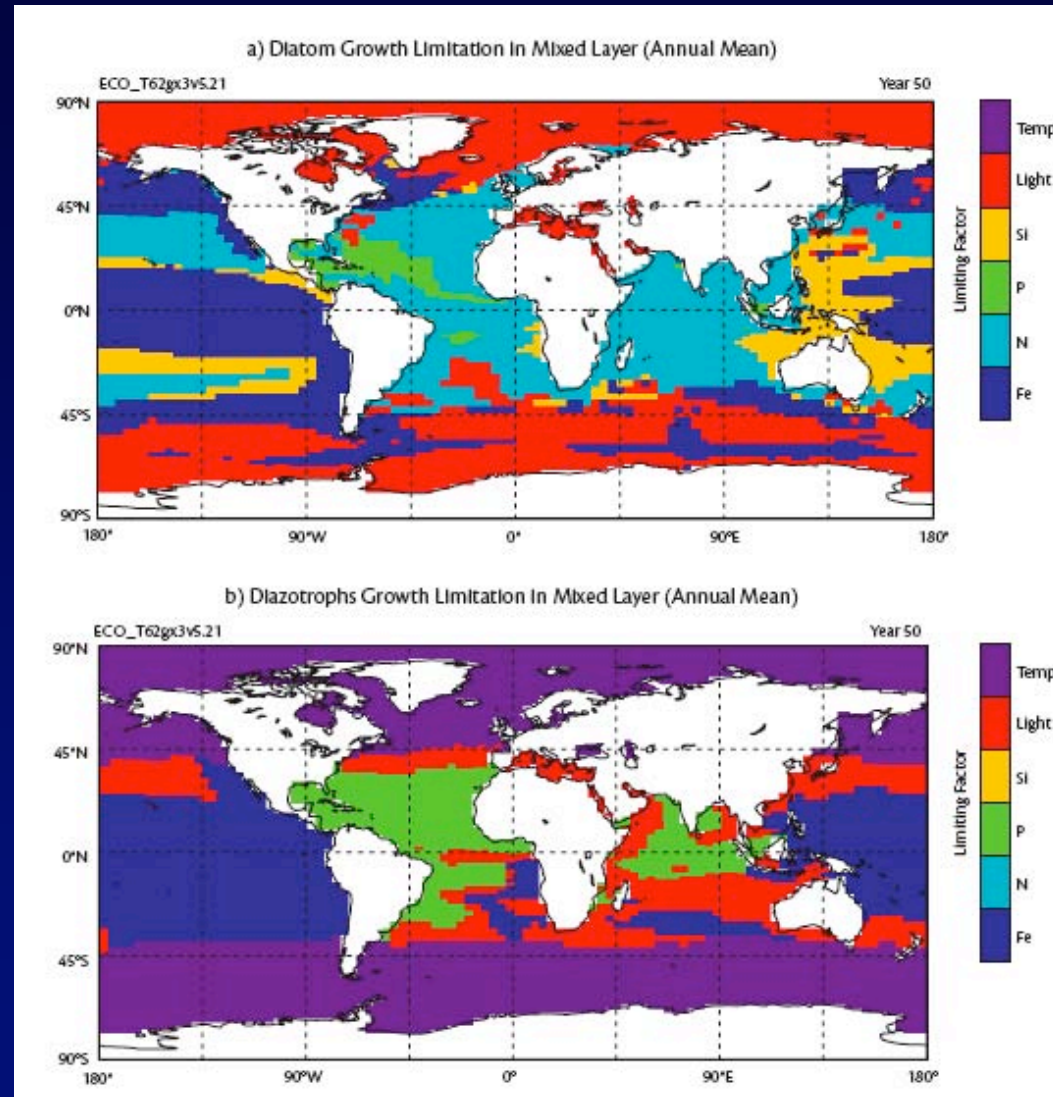


Fig. 5. *Thalassiosira pseudonana* and *T. oceanica*. Short-term changes of *in vivo* fluorescence per cell normalized to the highest observed value in each experiment. (A) *T. pseudonana* at 600 (○) or 800 $\mu\text{mol m}^{-2} \text{s}^{-1}$ (●) exposed to 2800, and then to 600 or 800 $\mu\text{mol m}^{-2} \text{s}^{-1}$ and *T. oceanica* (■) at 800 $\mu\text{mol m}^{-2} \text{s}^{-1}$ exposed to 2800 and then to 800 $\mu\text{mol m}^{-2} \text{s}^{-1}$.

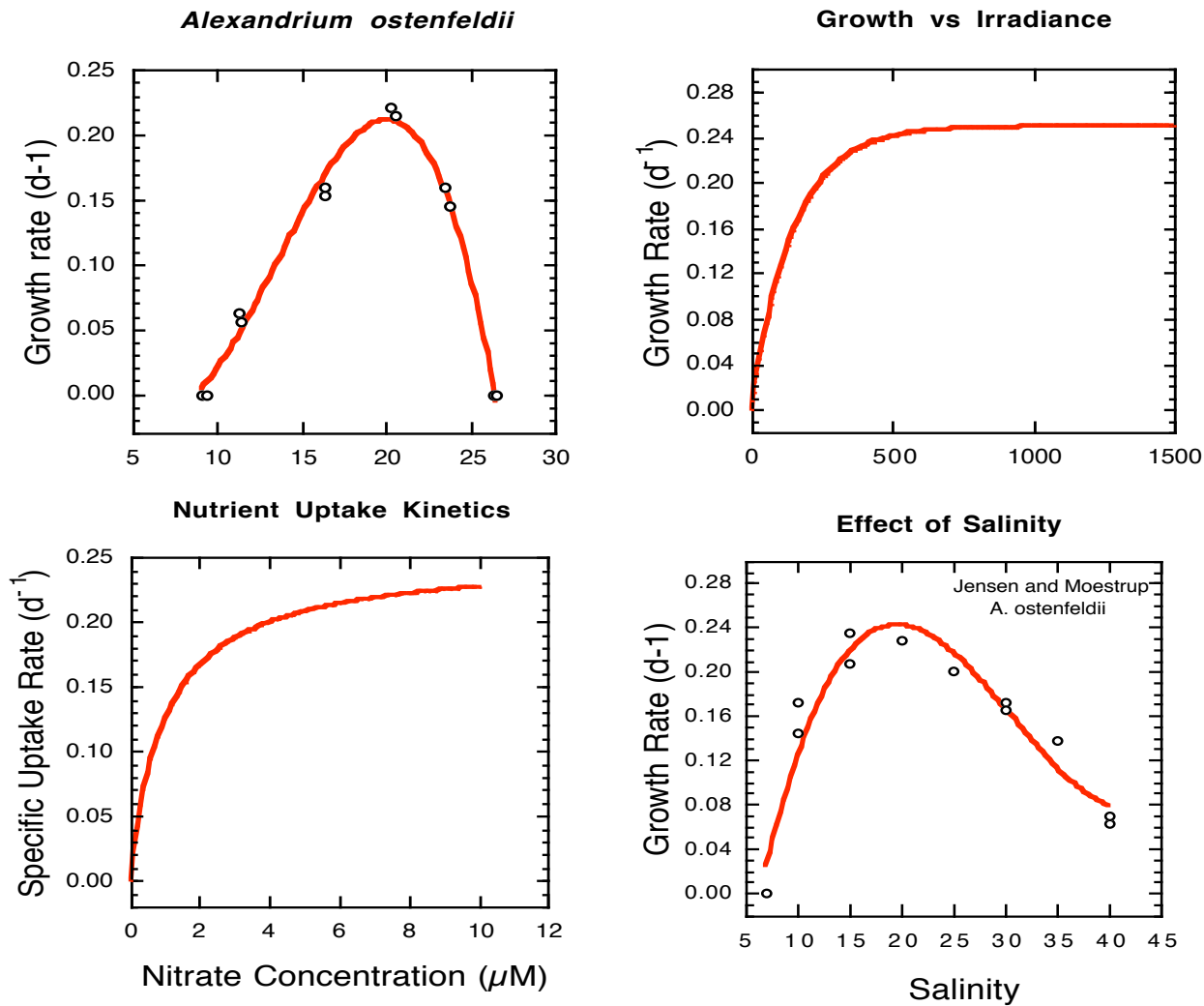
Niches abound!



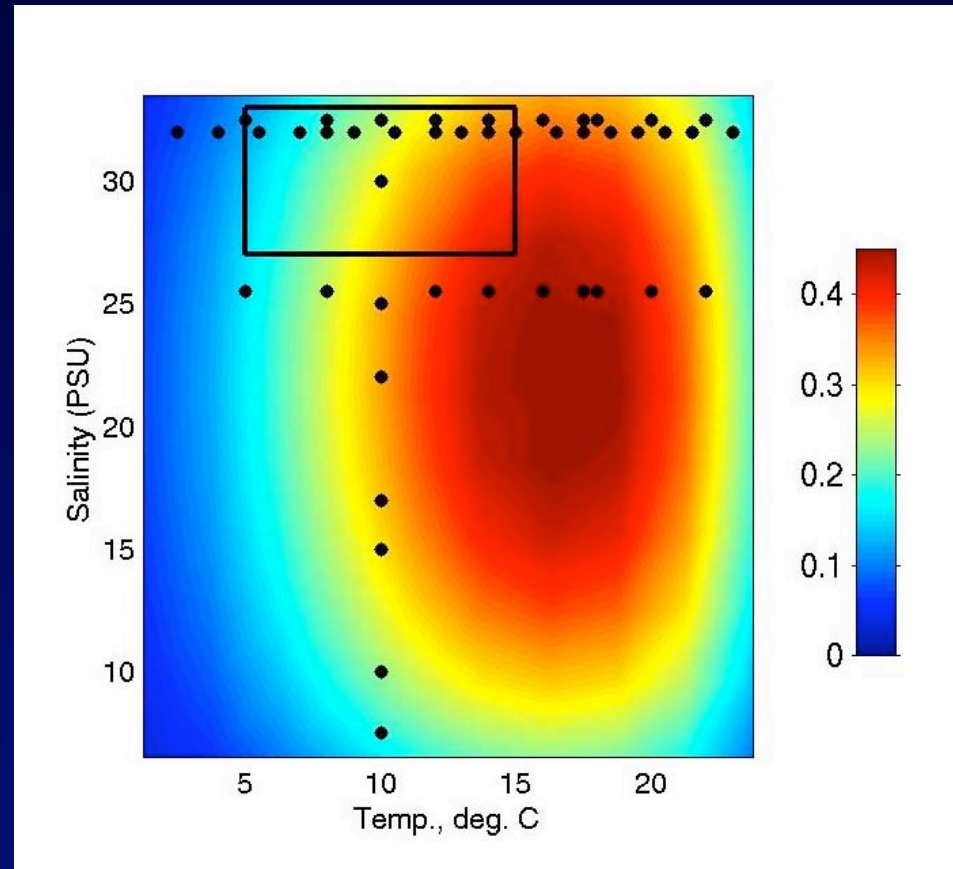
But what about quantitative prediction?



Growth must be described as a function of environmental conditions



Functions can be Developed



McGillicuddy,
Stock, Anderson,
and Signell
WHOI

$$\mu = f(D, E, N, T)$$

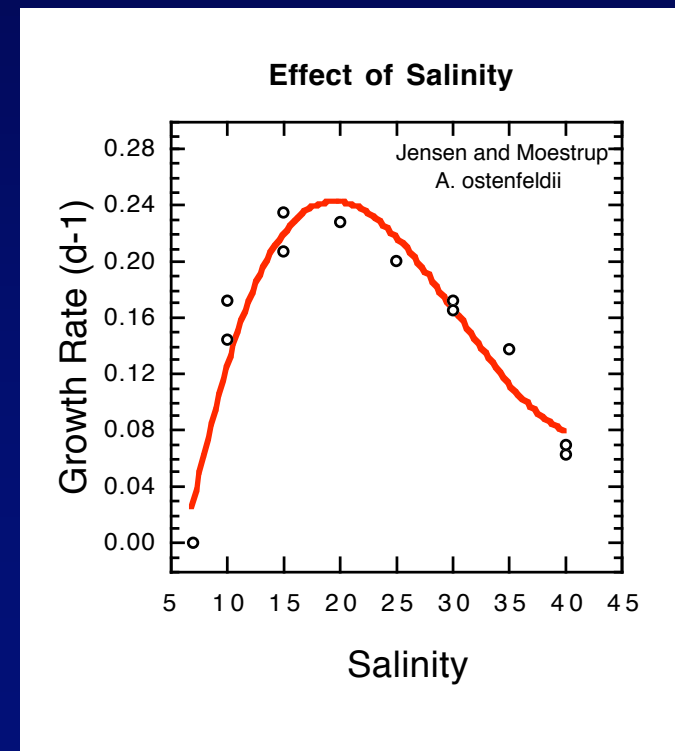
(Daylength, Irradiance, Temperature, Nutrients)

...but

- Requires a huge amount of work with cultures
- Algae should be acclimated to each set of conditions
 - This can require several weeks
- Conditions in nature are almost never so stable
 - Phytoplankton are subject to vertical mixing
 - Vertical migration
- All combinations of *Daylength*, *Irradiance*, *Nutrients* and *Temperature* are essentially impossible to test

Summary: μ as a function of environmental conditions

Good for identifying environmental ranges and optima



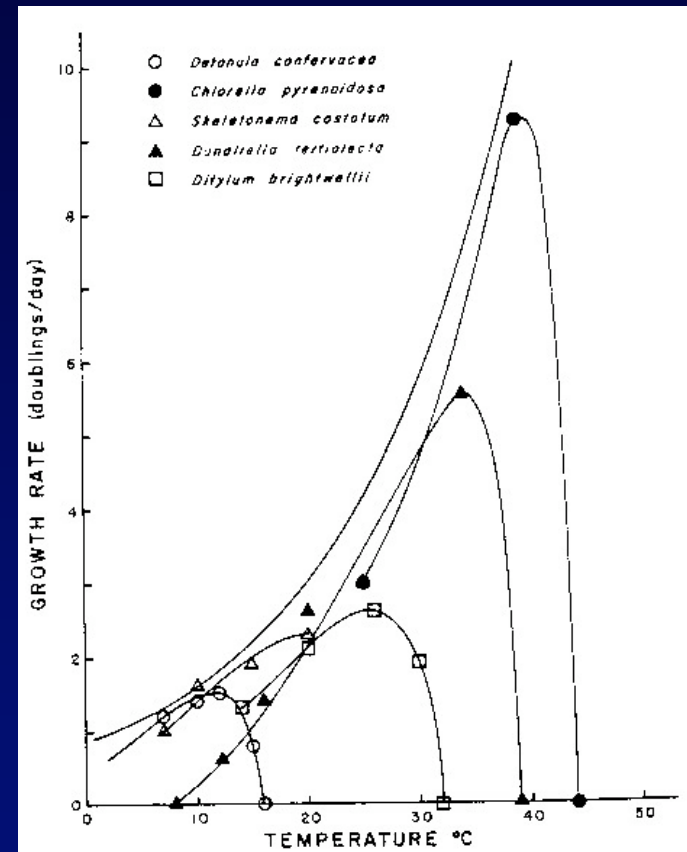
Summary: μ as a function of environmental conditions

Excellent for describing differences between species

(and variations among strains of the same species)

Ancient example:

Gallagher, J. C. (1982). Physiological variation and electrophoretic banding patterns of genetically different seasonal populations of *Skeletonema costatum* (Bacillariophyceae). *J. Phycol.* 18: 148-162.



Even more ancient:

Eppley, R. W. 1972. Temperature and phytoplankton growth in the sea. *Fish. Bull.* 70: 1063-1085.

Can we assume that adaptation provides all the raw genetic material for aggregate super-bugs?

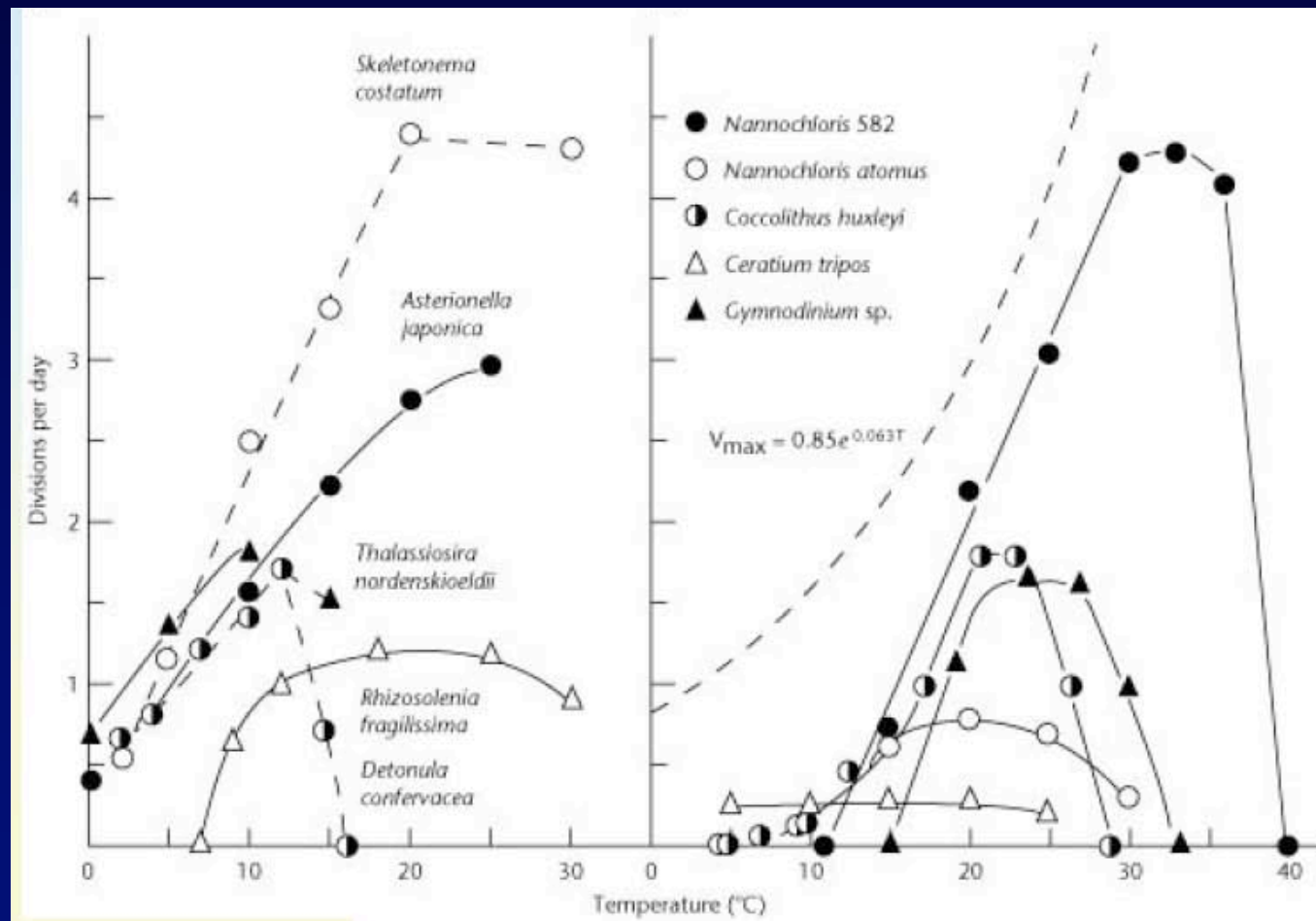


Figure from C.B. Miller, "Biological Oceanography" after Smayda, 1976

see Eppley, R. W. 1972. Temperature and phytoplankton growth in the sea. Fish. Bull. 70: 1063-1085.

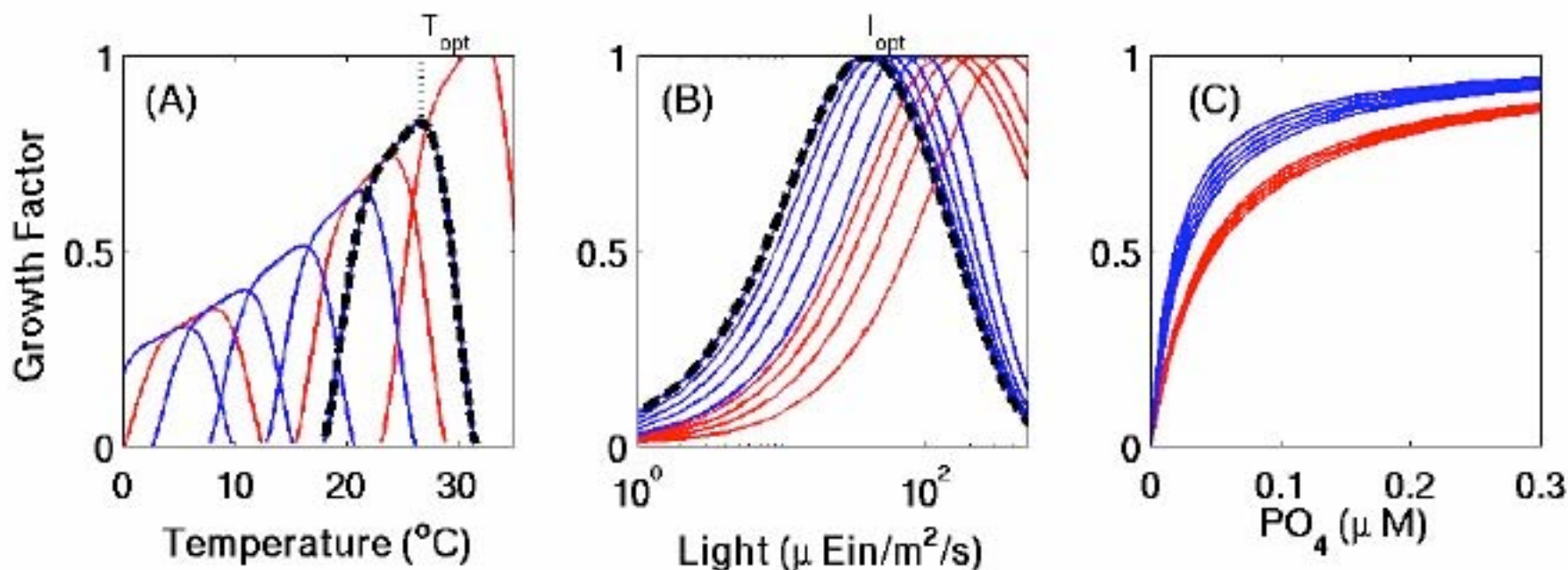
A novel approach

Emergent Biogeography of Microbial Communities in a Model Ocean

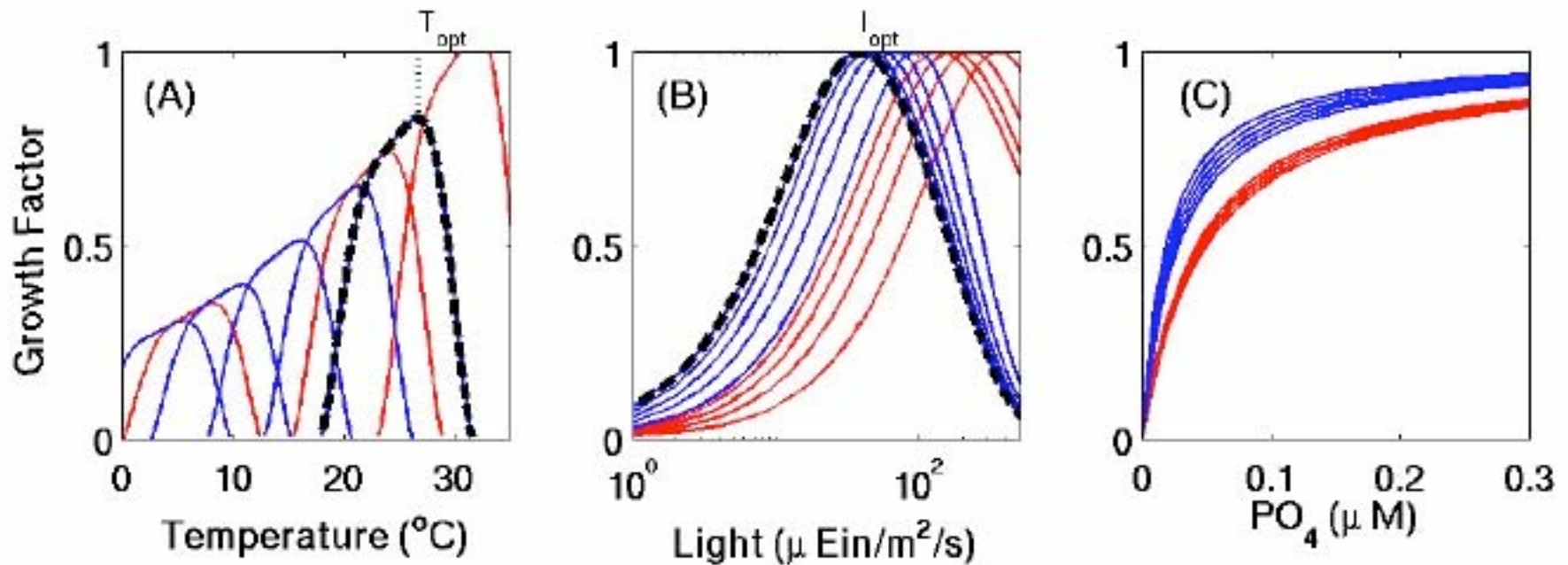
Michael J. Follows,^{1*} Stephanie Dutkiewicz,¹ Scott Grant,^{1,2} Sallie W. Chisholm³

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Everything (well, a lot of things) is everywhere and the environment selects



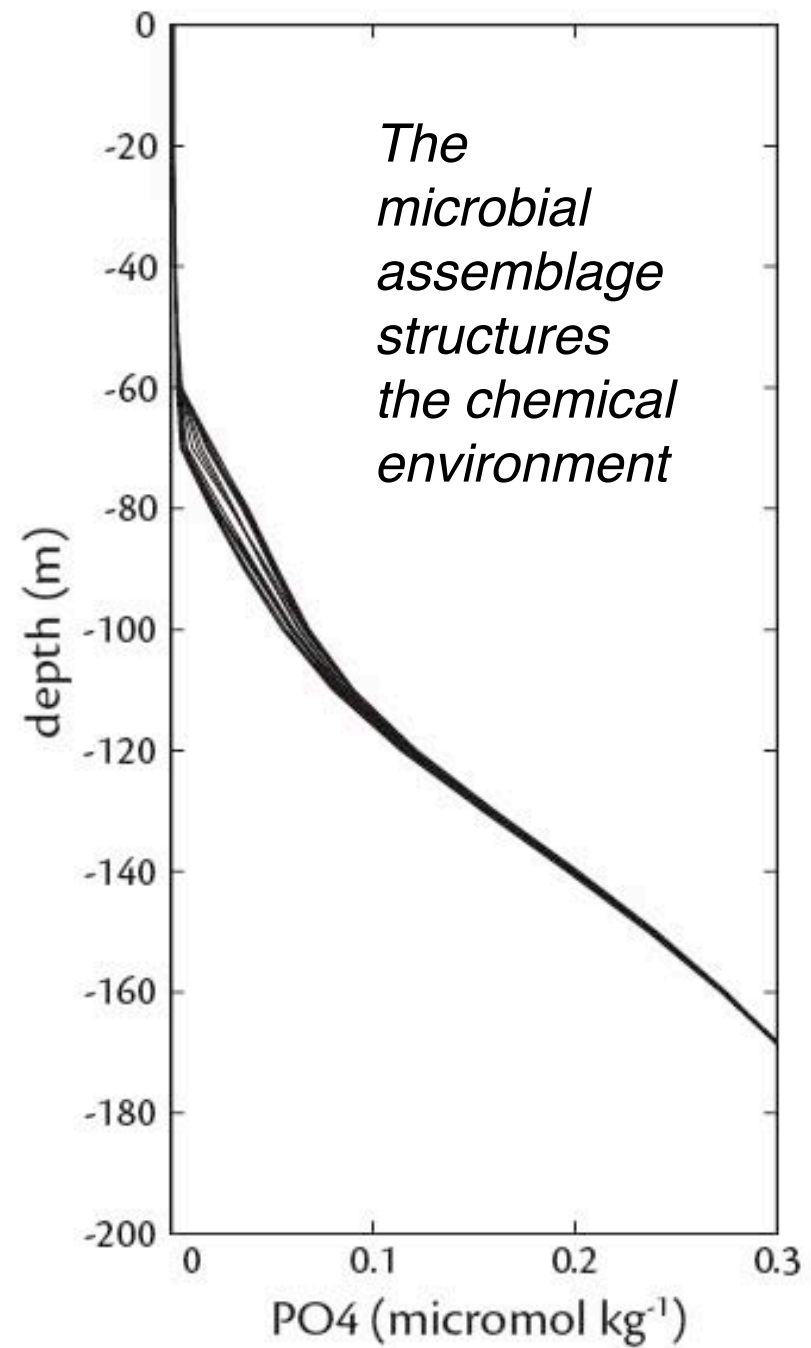
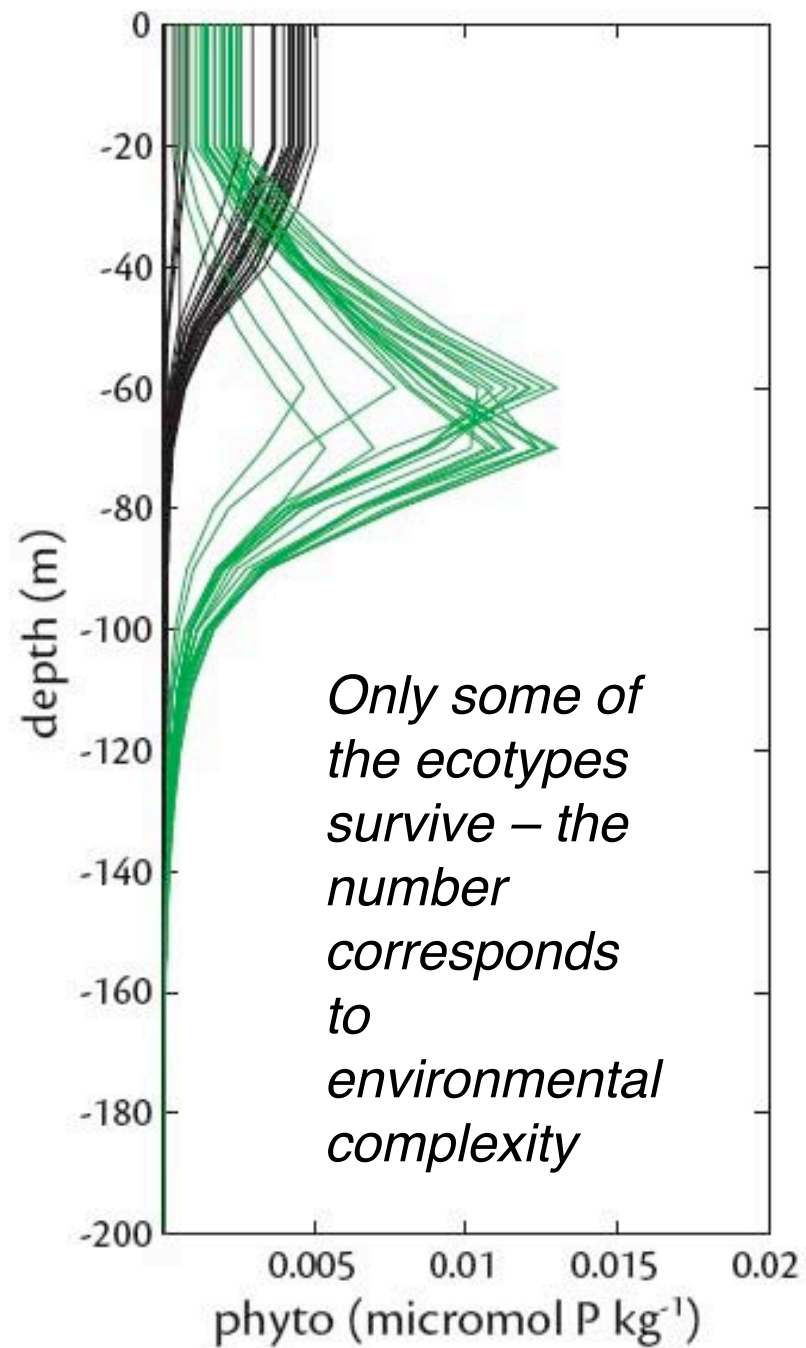
Emergent Biogeography of Microbial Communities in a Model Ocean

Michael J. Follows,^{1*} Stephanie Dutkiewicz,¹ Scott Grant,^{1,2} Sallie W. Chisholm³

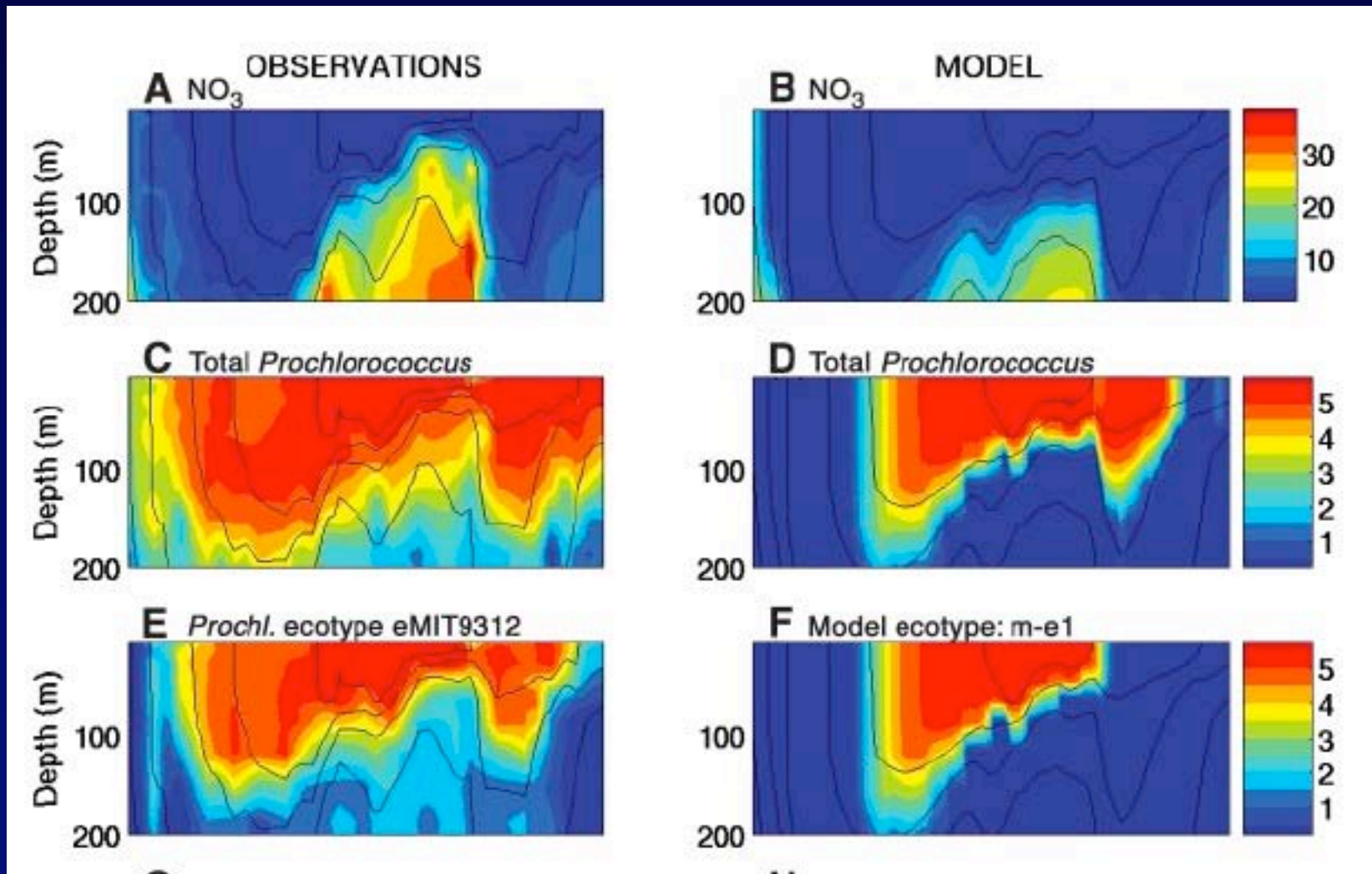
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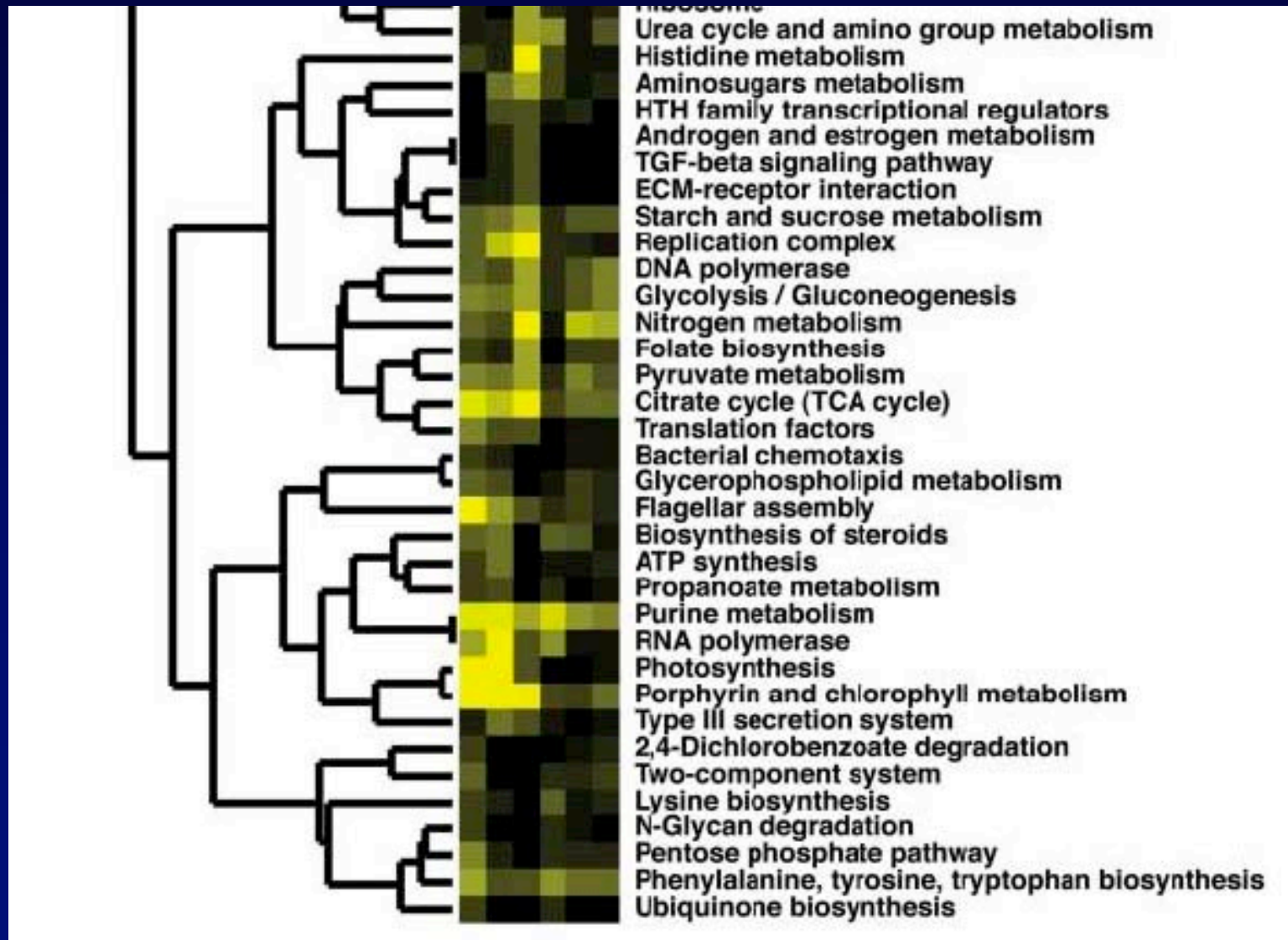
Natural selection *in silico*



A test of our understanding of what structures marine ecosystems (LPG)

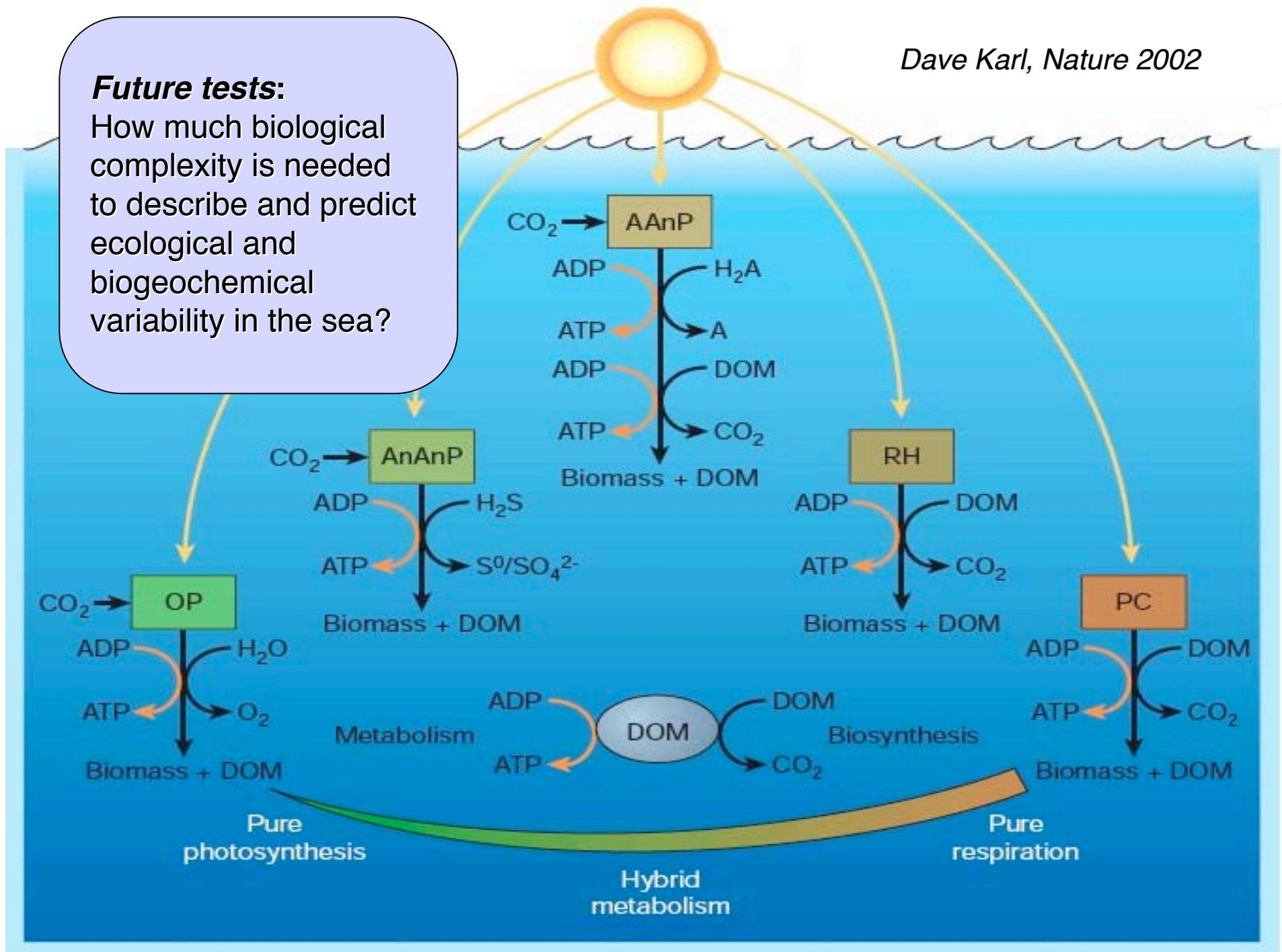


Future steps: Assembly and natural selection of microbial communities guided by metagenomics



Future tests:

How much biological complexity is needed to describe and predict ecological and biogeochemical variability in the sea?



Conclusions

Relationships between environmental variability and microbial diversity must be described and ultimately predicted to understand the ecology and biogeochemistry of the sea

Niches abound:

- wide range of environmental tolerances

- specialized life-styles (nutrient requirements, alternate modes of photosynthesis)

- physiological plasticity vs specialization for stable environments

Complexity will never be fully described with numerical models

The degree of model complexity can and should be related to the ecological/biogeochemical question and its scale.

This can be done — but models must be verified by measurements!

Mahalo!